

Systematic Review

The Role of Platelet Concentrates and Growth Factors in Facial Rejuvenation: A Systematic Review with Case Series

Giuseppina Malcangi ^{1,†}, Angelo Michele Inchingolo ^{1,*+‡}, Alessio Danilo Inchingolo ^{1,†}, Laura Ferrante ¹, Giulia Latini ¹, Irma Trilli ¹, Paola Nardelli ¹, Marialuisa Longo ¹, Andrea Palermo ², Francesco Inchingolo ^{1,*‡} and Gianna Dipalma ^{1,‡}

¹ Department of Interdisciplinary Medicine, University of Bari “Aldo Moro”, 70124 Bari, Italy; giuseppinamalcangi@libero.it (G.M.); ad.inchingolo@libero.it (A.D.I.); laura.ferrante79@virgilio.it (L.F.); dr.giulia.latini@gmail.com (G.L.); trilliirma@gmail.com (I.T.); dr.nardelli.paola@gmail.com (P.N.); dott.marialuisa.longo@gmail.com (M.L.); giannadipalma@tiscali.it (G.D.)

² Department of Experimental Medicine, University of Salento, 73100 Lecce, Italy; andrea.palermo@unisalento.it

* Correspondence: angelo.inchingolo@gmail.com (A.M.I.); francesco.inchingolo@uniba.it (F.I.)

† These authors contributed equally to this work as co-first.

‡ These authors contributed equally to this work as co-last.

Abstract: *Background and objectives:* Due to the regeneration potential of growth factors (GFs) and platelet concentrates (PCs), facial rejuvenation has been a major area of attention in esthetic medicine. The effectiveness and safety of PCs and GFs in promoting face rejuvenation are examined in this systematic review, which is complemented by a case series. GFs are essential for collagen production and dermal matrix remodeling, while PCs, like Platelet-Rich Plasma (PRP), are abundant in bioactive chemicals that promote tissue healing and cellular regeneration. *Materials and Methods:* A comprehensive literature search was performed on PubMed, Web of Science, and Scopus, focusing on human clinical trials published between February 2019 and February 2024 related to PRP and facial esthetics. *Results:* Thirteen studies met the inclusion criteria and were analyzed. *Conclusions:* The review summarizes the most recent data on patient outcomes, treatment regimens, and possible hazards. The case series that goes with it shows real-world examples of how to improve skin elasticity, texture, and general facial appearance with little negative side effects. These results highlight the potential use of PCs and GFs as minimally invasive procedures.

Keywords: PRP; facial esthetic; photoaging; rejuvenation; skin; growth factors; regenerative medicine; wrinkles; skin degradation; skin aging



Academic Editor: Māra Pilmane

Received: 17 November 2024

Revised: 29 December 2024

Accepted: 30 December 2024

Published: 7 January 2025

Citation: Malcangi, G.; Inchingolo, A.M.; Ferrante, L.; Latini, G.; Trilli, I.; Nardelli, P.; Longo, M.; Palermo, A.; Inchingolo, F.; et al. The Role of Platelet Concentrates and Growth Factors in Facial Rejuvenation: A Systematic Review with Case Series. *Medicina* **2025**, *61*, 84. <https://doi.org/10.3390/medicina61010084>

Copyright: © 2025 by the authors. Published by MDPI on behalf of the Lithuanian University of Health Sciences. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The ‘healthy and youthful appearance of the skin is closely determined by a system of balance between anabolism and catabolism of extracellular matrix (ECM) proteins. The ‘progressive alteration of these metabolic processes results in decreased microvascularization, reduced production of collagen and elastin, increased degradation of them, and consequent production of thinner and disorganized fibers [1]. This imbalance results in the appearance of wrinkles, furrows, altered pigmentation, loss of elasticity, texture, and skin radiance, better defined as skin aging [2–4].

Two biologically distinct aging processes are recognized: intrinsic (reduction in hydration, thinning of the dermis due to loss of collagen, and degradation of the elas-

tic fiber network) and extrinsic (environmental factors, photoaging due to exposure to ultraviolet rays) [5–8].

In both aging processes, there is an increase in matrix metalloproteinases (MMP)-1, MMP-2, and MMP-9 that cause degradation of dermal connective tissue fibers and matrix proteins [9–12].

Photoaging by ultraviolet (UV) radiation manifests a greater presence of MMPs with more pronounced connective tissue alterations, given by a greater space present between connective tissue fibers and a greater reduction in vascularization, the latter probably due to the damage and thus the difficulty of the connective tissue no longer being able to maintain the normal structure and function of the vascular system in the damaged skin [13–17].

Understanding the biological mechanisms of changes in skin angiogenesis and photoaging skin processes, thinking about using a therapeutic agent that can block degradation processes and activate regenerative processes, could promote esthetic regenerative treatments of the face and beyond [18–22].

Considering the tissue rejuvenation process comparable to the metabolic processes triggered in wound healing processes, autologous platelet concentrates (PC) were considered [23–33].

The concentration of platelets in the human blood of a healthy patient is 150,000–350,000/microliter. PCs contain about 1,000,000 of them per microliter in a minimal volume of plasma [34,35].

PCs and autologous growth factors (AGFs), obtained by different processes and techniques of centrifugation of patient blood sampling, have been considered to slow down skin aging processes and stimulate cellular activity, improving the esthetics of the face, neck, decolletage, and even skin of the hands, areas more easily marked by aging [18,36–39]. The use of PCs and AGFs has evolved in use over the years. The first application was used in maxillofacial surgery in 1994, in the form of platelet gel (referred to as Toyapongsaki) in maxillofacial surgery as a bone mass thickener in a mandibular excision, to be later applied in many branches of specialty medicine [40–43].

To date, several forms of GF use are known and have been refined over the years. Modifying preparation methods, based on leukocyte concentrations and on ‘fibrin architecture, the different categories are distinguished: fibrin glue (Tissucol Baxter 1994), Platelet-Concentrated cPRP (Marx-Garg 1998), Platelet-Rich Plasma (PRP), Plasma Rich in Growth Factor (PRGF 1998 Anitua), Platelet-Rich Fibrin (PRF 2001 J. Choukroun) to the best-performing PCs in regenerative medicine with Concentrated Growth Factors (CGF Corigliano et al., 2006), Liquid Phase Concentrated Growth Factors (LPCGF) and Activated Plasma Albumin Gel (APAG) [44–47].

The reparative function of platelets has long been known through the degranulation of alpha granules, which are considered the most important growth factors (GF), such as platelet-derived growth factors (PDGF α , PDGF β , PDGF δ), endothelial growth factor (VEGF), transforming growth factor β 1 and β 2 (TGF- β 1 and TGF- β 2), TNF- α , basic fibroblast growth factor (bFGF), and epithelial growth factor (EGF), all of which play an important role in the early phase of the wound healing process, accelerating it [48–51].

In the scientific literature, there is evidence of the multiplicity of PRP preparation techniques, administration intervals, and sites treated with a finding of varying results between them, although, in general, its use manifested improvement in the parts of the face, hands, and decollete treated (Figure 1) [34,52–54].

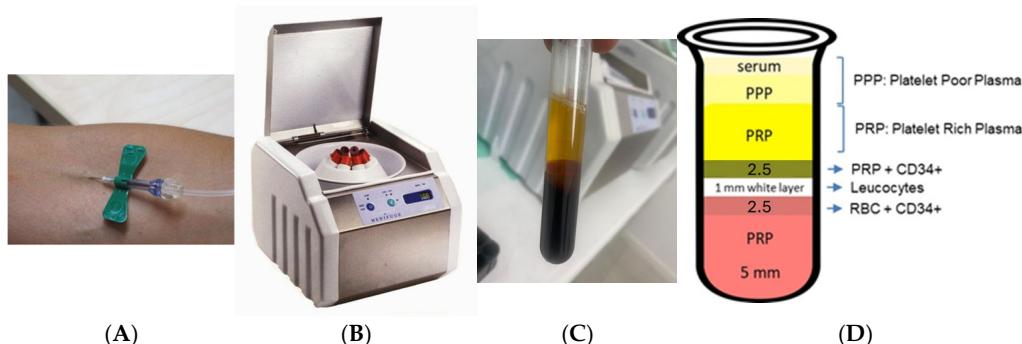


Figure 1. PRP preparation process. Venous blood is taken from the patient (A) and centrifuged with the Medifuge 200 device Silfradent® (B) to divide it into three distinct layers (C,D): red blood cells (RBC) at the base, PRP at the top, and the buffy coat in between. This illustration provides a rough breakdown of cell types within the buffy coat, emphasizing the arrangement of neutrophils, eosinophils, basophils, monocytes, lymphocytes, and platelets [55,56].

PRF is an evolution of PRP and PRGF, in solid (gel) or liquid form, obtained by a single centrifugation without the addition of anticoagulants or platelet activators, hence completely autologous [57–60]. Lower relative centrifugal force (RCF), shorter centrifugation time, and the use of plastic tubes create an injectable liquid product (I-PRF) capable of capturing more platelets and leukocytes that, by remaining trapped in the gelled fibrin matrix after inoculation, can slowly release GF [48,61–64].

Leukocytes also stimulate fibroblast propagation, enhance anti-inflammatory effects, angiogenesis, and production of proteins (e.g., procollagen) necessary for structure and remodeling of the extracellular matrix [46,65–67].

The bioregenerative results of both objective and subjective areas of facial skin treated with PRF are found to be very high performing with a reduction in wrinkles and pigmentation and increased skin brightness and elasticity [68–76].

The latest tissue bioengineering studies have focused on evaluating the speed, time, and ‘angle’ of centrifugation of venous blood sampling to obtain a product with greater reparative and regenerative predictability [77–80].

PPP is the top part of the centrifugate and contains a much higher amount of fibrinogen, albumin, white blood cells, and soluble cytokines [81–84]. Its use in esthetic medicine manifested immediate and lasting effects on fine wrinkles, with greater efficacy on dark circles and infraorbital folds by acting as a scaffold for the sustained release of GF than PRP, which, by stimulating the release of fibroblasts and acting as an anti-inflammatory, improved skin smoothness and texture [85–88].

Heating PPP (2 mL) to 75 °C for 10 min with a dedicated frequency heater (APAG Silfradent®) produces gelled PPP, which, when mixed with CD34+ (0.5 mL) and PRP (0.5 mL), produces APAG, an autologous filler enriched with stem and GF. APAG produces better-performing results, combining the immediate smoothing effect of the gel on wrinkles with prolonged cellular activation over time due to the presence of stem and GF (Figure 2) [89–92].

The technique on the use of PC and GF, derived from autologous products, appears to be safe. The appearance of small hematomas, the use of needles for product inoculation or microneedling, appear to be the only aspects of discomfort for the patient [93,94].

The purpose of this work was to be able to evaluate the use of GFs and PCs as a viable therapeutic choice in skin rejuvenation [95–97].

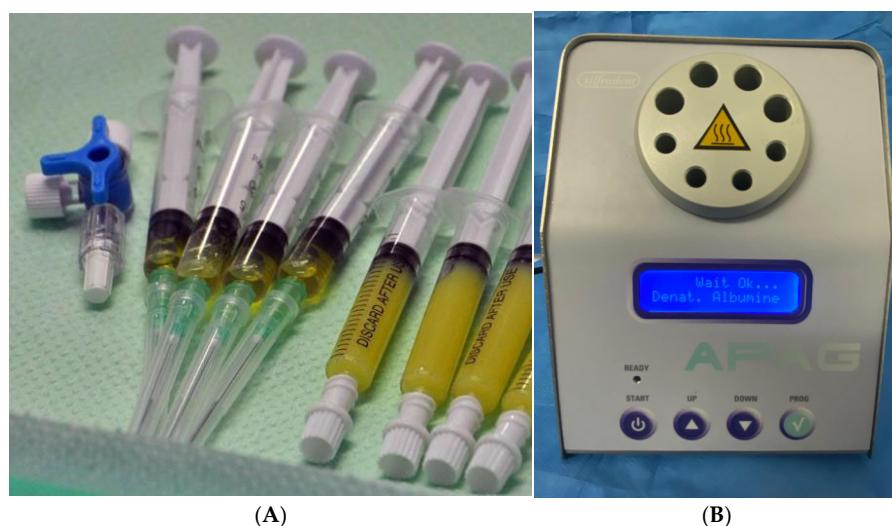


Figure 2. APAG on the right and PRP with CD34+ on the left for autologous filler preparation (A); APAG Silfradent® (B).

Unfortunately, more clinical studies are needed, as scientific evidence on the use of PCs AND AGFs is currently very limited [98–104].

The study addresses skin aging caused by the imbalance in ECM protein turnover, leading to collagen degradation, loss of elasticity, and wrinkles. It highlights the potential of platelet concentrates (PC) in regenerative esthetics to improve skin quality and counteract aging effects. The study explores how PCs stimulate collagen production, enhance elasticity, and support dermal regeneration. It also emphasizes standardizing preparation protocols to improve clinical outcomes and address gaps in current anti-aging treatments.

2. Materials and Methods

2.1. Protocol and Registration

This systematic review was conducted by the standards of the Preferred Reporting Items for Systematic Reviews and Meta-analysis (PRISMA) 2020 statement [105–108]. The protocol of the review was registered at PROSPERO under the unique number 601404.

2.2. Search Processing

The search period started on 12 August 2024, and the last search was carried out on 25 September 2024.

“PRP”, “Facial Aesthetic”, “Photoaging”, and “Rejuvenation” were the search terms utilized on the databases (PubMed, Web of Science, and Scopus) to select the papers under evaluation, with the Boolean operators “AND” and “OR”. Only content published in English over the previous five years (February 2019 to February 2024) was included in the search (Table 1).

Table 1. Database search indicators.

Article screening Strategy	Database: Scopus, Web of Science, and PubMed
	Keywords: A “PRP”; B “Facial Aesthetic”; C “Photoaging”; D “Rejuvenation”;
	Boolean variable: “AND” and “OR”
	Timespan: 2014–2024
	Language: English

2.3. Eligibility Criteria

Working in pairs, the reviewers selected pieces that met the following requirements to be included: (1) research involving just human beings; (2) clinical studies; and (3) our most relevant case series about skin aging and scars.

One of the exclusion criteria was in vitro research. The following were also included: (1) research on animals; (2) case reports; and (3) narrative reviews, meta-analyses, and systematic reviews.

Duplicate studies were removed manually.

2.4. Data Processing

Working separately, two reviewers (I.T. and A.D.) used the predetermined inclusion and exclusion criteria to filter the data that was taken from each database. Individual choices were hidden from the researchers. Both reviewers' results were converged upon in the final meeting. The complete text was obtained and examined when a reviewer thought an article might be accepted. Both independently and twice, this occurred.

Each qualifying main study's authors and publication date, study type, purpose, materials and methods, and findings are among the data that were taken from it.

Reviewers' disagreements on which article to choose were resolved through discussion.

2.5. Quality Assessment

Two reviewers, G.L. and F.I., evaluated the included papers' quality using the reliable Cochrane risk-of-bias assessment for randomized trials (RoB 2). This test assesses six potential areas of bias: inadequate outcome data, selective reporting, blinding of participants and staff, random sequence generation, allocation concealment, and outcome assessment blinding. A third reviewer (L.F.) was consulted if there was a disagreement and continued until a consensus was reached.

3. Results

Keyword searches of the Web of Science (229), Scopus (118), and PubMed (228). A total of 575 articles were found in the databases. After the duplicates were eliminated (240), 335 articles were included. In addition, 322 of these 335 studies were disqualified for violating the inclusion criteria that had been previously established. After screening, thirteen papers were chosen for this work (Figure 3). Each study's findings were listed in Table 2.

In total, seven studies were randomized, and nine studies were non-randomized. This includes randomized clinical trials, randomized controlled trials, and non-randomized observational studies.

The minimum group size was 10 participants, as seen in studies like the one by Tsai et al. (2024) [109] with 10 participants. The maximum group size was 94 participants, as reported in studies such as Ulusal B. (2017) [110], which involved 94 female patients.

The most common reasons for therapy across the studies include facial skin rejuvenation, photodamage, and aging-related skin conditions. PRP (Platelet-Rich Plasma) was most frequently used to treat issues like wrinkles, skin texture, and overall skin vitality, with the goal of improving facial appearance and reversing signs of photoaging.

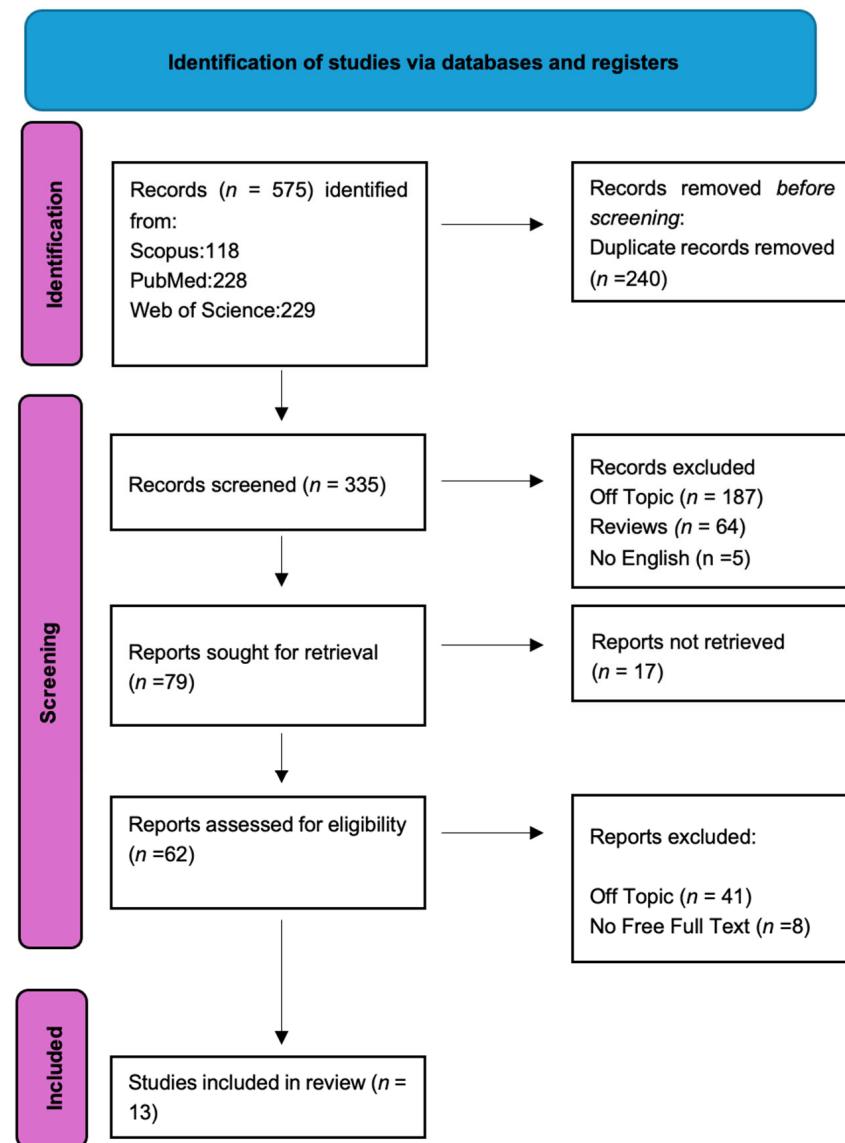


Figure 3. PRISMA flowchart diagram of the inclusion process. The literature search's Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) flow diagram.

PRP (Platelet-Rich Plasma) was the most frequently used therapeutic agent across the studies, often in combination with other treatments like hyaluronic acid (HA) or saline solution. In some studies, other treatments such as ADSC (Adipose-Derived Stem Cells) and PPP (Platelet-Poor Plasma) were also evaluated.

Some common limitations identified in the studies include small sample sizes, lack of standardization in PRP preparation and application, variability in treatment protocols, and limited follow-up periods. There were also issues with inconsistent results across different studies, making it difficult to generalize findings. Errors related to subjective assessments, such as patient satisfaction or physician evaluations, and the absence of long-term efficacy data were also noted as limitations.

Table 2. Qualitative analysis of the studies included.

Authors	Type of the Study	Patients	Material and Methods	Aim of the Study	Conclusions
Hassan et al. (2020) [111]	Prospective, uncontrolled	11 females	Monthly intradermal injections of injectable i-PRF for 3 months in specified facial areas.	To evaluate the efficacy of i-PRF for facial skin rejuvenation using objective skin analysis and patient-reported outcomes.	Significant improvements in skin spots and pores; increased patient satisfaction.
Draelos et al. (2020) [112]	Pilot study	20 subjects (30–60 years, both genders)	PRP in a preservative serum, applied twice daily for 8 weeks after electroporation.	Evaluate PRP serum effects on facial photoaging.	PRP showed 90-day stability; improved rete peg architecture and collagen I expression.
Rina Du et al. (2020) [113]	Clinical study	30 females (30 to 50 years)	Autologous PRP injected three times at 15-day intervals, analyzed with VISIA® and organotypic skin models.	Investigate the molecular mechanisms of PRP in rejuvenating aged skin.	PRP improved skin quality, reducing wrinkles and photoaging markers.
Gawdat et al. (2024) [114]	Split-face randomized study	20 females (35 to 55 years)	PRP on one side, GF on the other; assessed with GAIS and OCT.	Compare PRP and GF for skin rejuvenation.	Both improved skin vitality; PRP had better long-term results and satisfaction.
Lee et al. (2023) [115]	Pilot study	31 participants (27 females, 4 males) aged 27 to 71 years (median age: 38)	PRP treatment, evaluated using the WSRS and GAIS.	To evaluate the effectiveness and patient satisfaction of PRP treatment for photodamaged skin.	Modest benefits in skin aging treatment; adverse effects mild (e.g., tenderness, swelling).
Charles-de-Sá et al. (2020) [116]	Comparative experimental study	20 human subjects with aged or photoaged skin.	Two different therapies: PRP injections and expanded ADSC therapy.	To compare the effects of PRP and ADSC therapy on aged human skin.	PRP did not produce significant tissue regeneration. ADSC treatment was linked to ECM remodeling, new elastic fiber production, and the degradation of elastotic material.
Murad Alam et al. (2018) [117]	Randomized clinical trial	27 participants aged 18–70, of which only 19 completed the study	Each participant received 3 mL of PRP in one cheek and saline in the other. Follow-up included digital photos at baseline, 2 weeks, 3 months, and 6 months.	To assess PRP's effect on photoaging and compare it with normal saline (as a placebo).	PRP did not result in a significant improvement in photoaging scores when compared with normal saline based on independent dermatologist assessments of standardized photographs.
Paweł Surowiak et al. (2023) [118]	Prospective non-randomized controlled clinical study	10 volunteers (5 women and 5 men) between the ages of 29 and 49	PRP versus placebo (NaCl). After 21 days, skin biopsies were performed.	To test whether PRP stimulates procollagen type I synthesis in human skin compared with placebo.	PRP increased the expression of procollagen type I compared with placebo.

Table 2. Cont.

Authors	Type of the Study	Patients	Material and Methods	Aim of the Study	Conclusions
Betul Gozel Ulusal (2016) [110]	Observational clinical study	94 female patients, average age 53.0 ± 5.6 years.	PRP kit (Dr B PRP™) and centrifuge. Hyaluronic Acid (HA) gel (3.5%) and procaine.	To evaluate the effectiveness of a combined treatment of PRP and HA for facial rejuvenation.	A combination of PRP and HA injections, with additional needling, resulted in significant facial rejuvenation.
Tsai Y. et al. (2024) [109]	Double-blind randomized controlled splitting face study	10 participants	The participants were randomly assigned to receive 2.5 mL injections of PRP and PPP on different sides of the face in three sessions with 1-month intervals.	Compare the efficacy of PRP and PPP for facial rejuvenation.	PRP and PPP are effective in treating facial photoaging. PRP exhibited slightly superior efficacy in enhancing overall skin condition, while PPP was slightly more effective in improving shallow wrinkles.
Everts P. et al. (2019) [119]		Eleven healthy female volunteers between 45 and 65 years old	All women signed an informed consent form before treatment with 3 facial PRP injections.	the efficacy of autologous PRP injections for facial skin rejuvenation.	A series of 3 Pure PRP injections at 6-month follow-up resulted in significant skin rejuvenation as demonstrated by biometric parameters and confirmed by patient self-assessment score.
De Silva L. et al. (2021) [120]	Non-randomized, controlled, pilot trial	Nineteen women ($54 \text{ years} \pm 7$ years) with Glogau photoaging II and III types	All patients received monthly intradermal injections of lyophilized PRP and saline solution (as a control) into the facial skin during a period of 2 months.	Evaluate the effect of lyophilized PRP in the treatment of skin aging.	In a well-controlled pilot split-face study using VISIA® and SHG methods, we did not demonstrate an effect of lyophilized PRP by mesotherapy on skin aging.
Hui Q. et al. (2017) [121]		13 female patients suffering from facial aging	13 facial aging females were treated with ultra-pulsed fractional CO ₂ laser. One side of the face was injected with PRP as the experimental group, while the other, as the control group, received the same dose of saline.	Evaluate the potential synergistic effects of combining PRP with ultra-pulsed fractional CO ₂ laser for facial rejuvenation therapy.	PRP and ultra-pulsed fractional CO ₂ laser had a synergistic effect on the therapy for facial rejuvenation.

Note: Adipose-Derived Stem Cell (ADSC), hyaluronic acid (HA), sodium chloride (NaCl), Platelet-Rich Plasma (PRP), Platelet-Rich Fibrin (i-PRF).

Case Series

In the Case Study section, images of the study participants are presented. Informed consent for the publication of these images was obtained from all participants. Additionally, the study was approved by the relevant ethics committee, which included consent for the use and publication of participant images.

Clinical case 1. A 26-year-old female patient, in good health, with a request for volume augmentation of the upper and lower lips. The treatment was performed with lip filler with APAG (Figure 4).

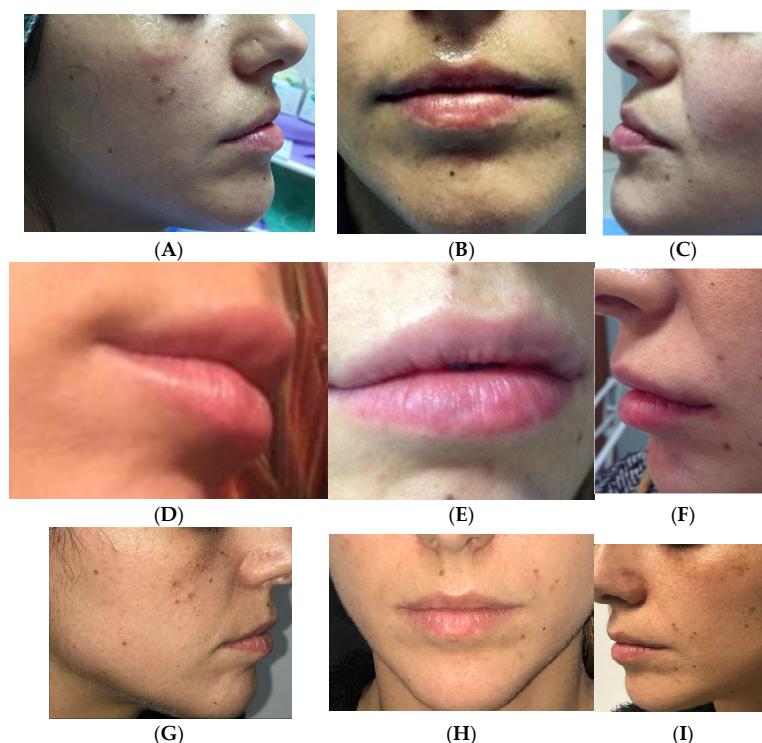


Figure 4. Before treatment (A–C), immediately after the first treatment (D–F), and after two years without other treatments (G–I).

Clinical case 2. Healthy female patient aged 45 years with a request for antiaging treatment, with repair of wrinkles and full-face skin texture. The treatment was filler with APAG and LP CGF for the lips and zygomatic area and mesotherapy with LP CGF full-face (Figure 5).

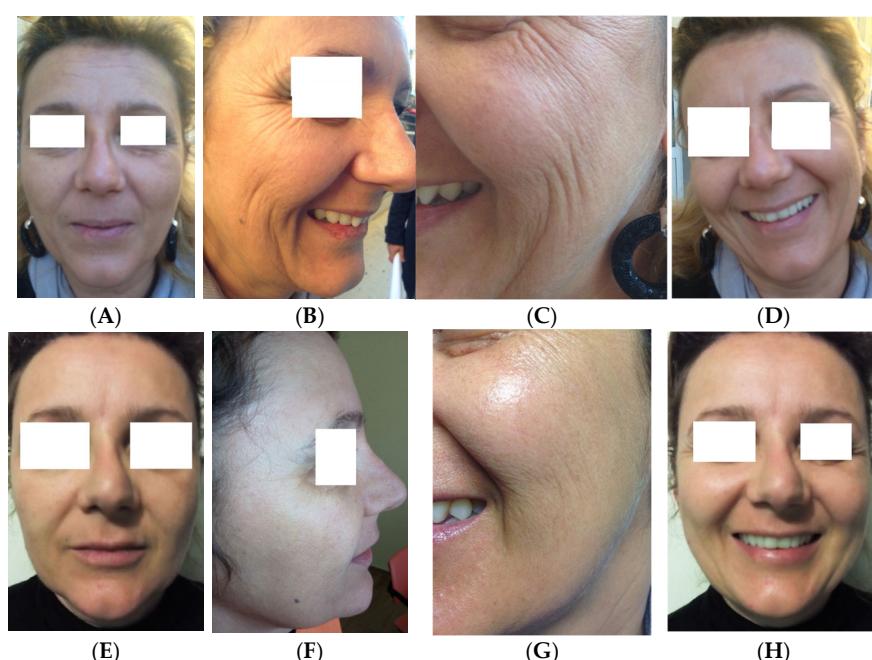


Figure 5. Before treatment, 13 December 2014 (A–D), and after three treatments, 24 March 2015 (E–H).

Clinical case 3. A 72-year-old female patient in good health with a request for antiaging treatment to resolve skin relaxation in the face and neck. The treatment was filler with APAG and LP CGF for the lips and zygomatic area and mesotherapy with LP CGF full-face (Figure 6).

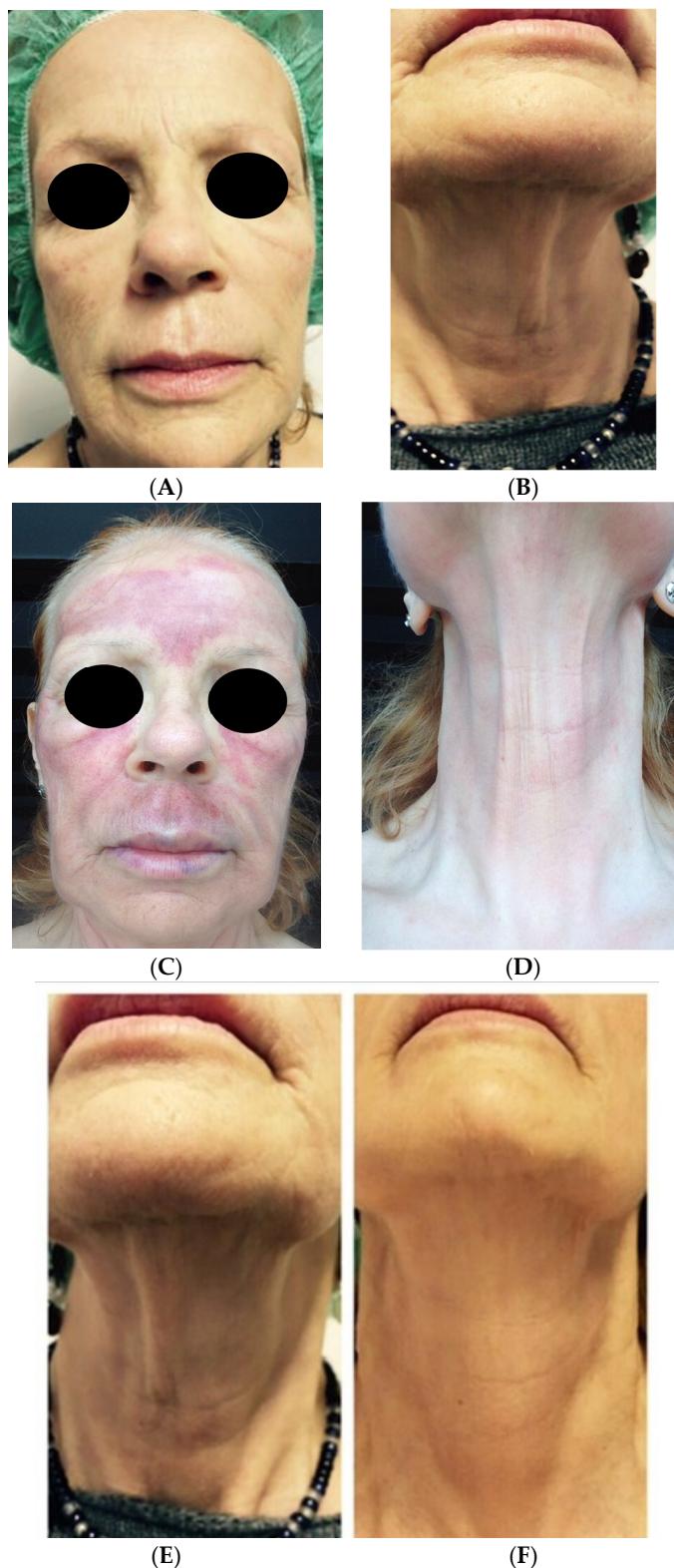


Figure 6. Cont.



Figure 6. Before (A,B) and immediately after the treatment (C,D) of LPCGF with microneedling. The desired esthetic effect was best achieved through a noticeable reduction in signs of aging and a general improvement in skin texture. In comparison: signs of skin aging in the neck, cheek, and cheekbone area before treatment (E,G,I) and the clear esthetic improvements in the same areas after treatment (F,H–J). Desired final esthetic result achieved of face and neck (K): improvement of skin texture and reduction in wrinkle depth.

Clinical case 4. A 16-year-old female patient in good health, requiring treatment for acne and acne scars. The treatment was CGF and microneedling (Figure 7).

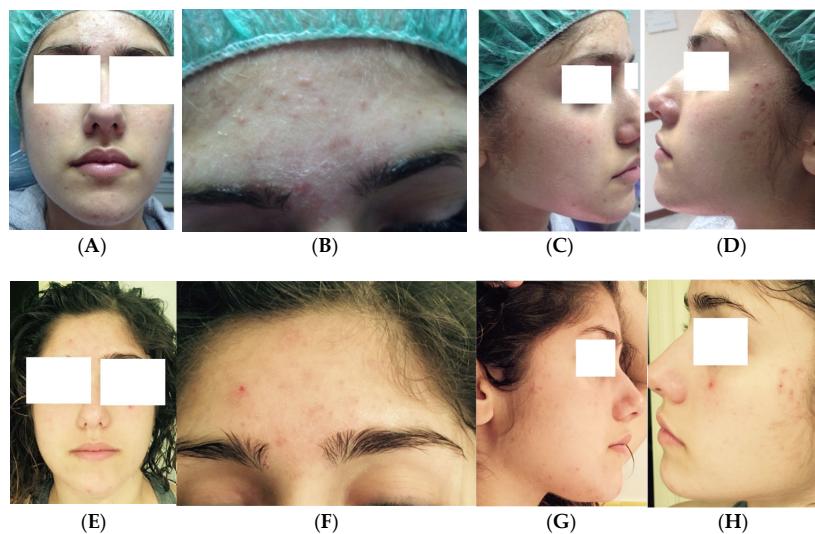


Figure 7. Before treatment (A–D) and 5 days after the first treatment (E–H).

Clinical case 5. Large post-surgical scar of thyroid carcinoma (40 cm) treated with LP CGF and microneedling in a 32-year-old female patient in good health (Figure 8).



Figure 8. Cont.

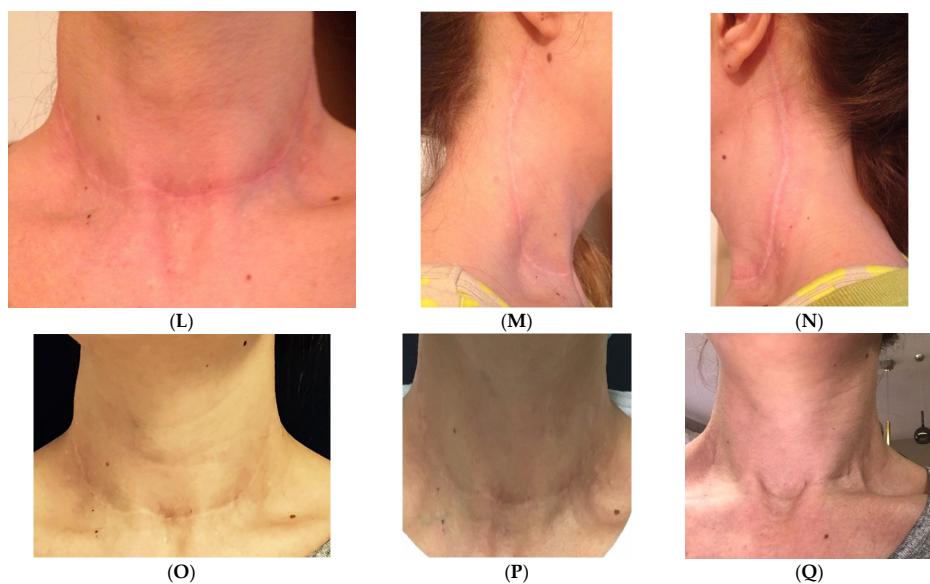


Figure 8. Before (A–C), during the first treatment (D,E), immediately after the first treatment (F–H), after three days (I–K), and after 10 days from the first treatment (L–N). Hyperemia and swelling of the scar area are expected consequences of surgery. Hyperemia, in particular, is due to the use of needling to a depth of 2/3 mm to treat scar fibrosis. These symptoms are temporary, normal, and part of the natural healing process. Over time, the hyperemia and swelling will gradually subside as the tissue heals (D–H). The same patient before (O), after three treatments (P), and after three years, without further treatment (Q).

Clinical case 6. A 42-year-old female patient, suffering from hypercholesterolemia treated with statins, with melasma, dark or hyperpigmented spots on the forehead and face, treated with LP CGF and microneedling full-face in two sessions 3 weeks apart (Figure 9).

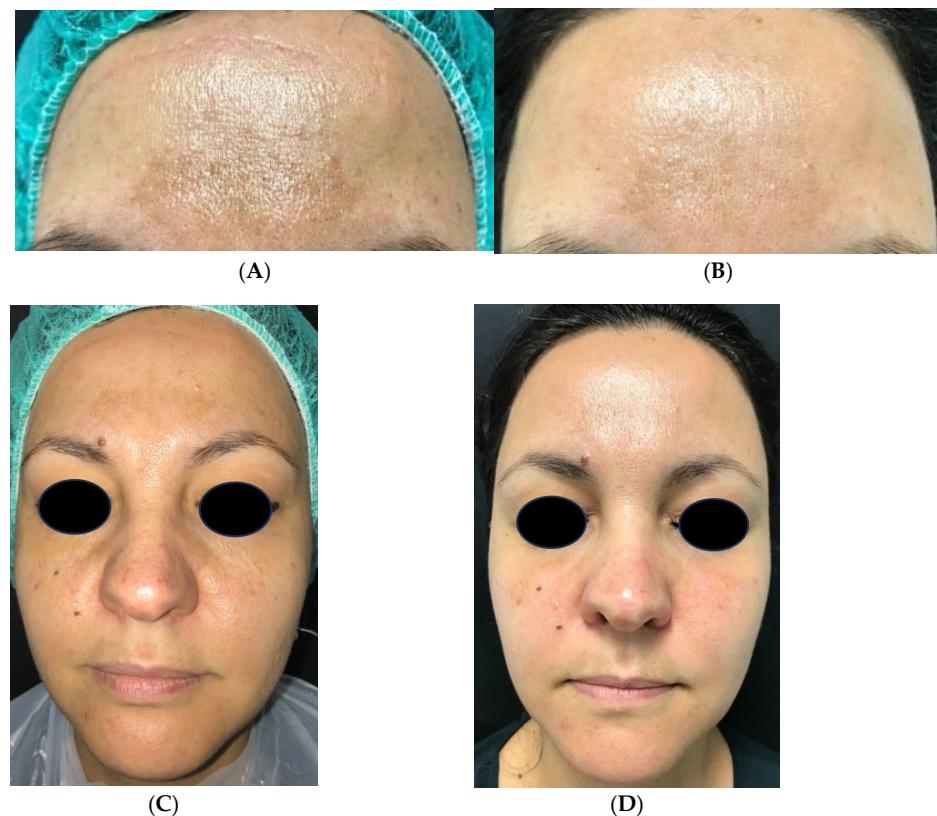


Figure 9. Before treatment (A,C) and after the second treatment, after 3 weeks (B,D).

4. Discussion

In recent years, PRP (Platelet-Rich Plasma) has gained popularity for its applications in the regenerative field, particularly in esthetic medicine and the treatment of skin aging. PRP is prepared using a patient's autologous blood and contains platelet GF that stimulate tissue repair and collagen production [122–126]. However, recent research indicates that another formulation, PRF (Platelet-Rich Fibrin), may have greater regenerative potential due to its three-dimensional fibrin matrix that allows prolonged release of GF, promoting more effective tissue regeneration. Whereas PRP is prepared by rapid centrifugation that separates platelets without clots, PRF is formed using slow centrifugation without anticoagulants, which leads to the formation of a fibrin lattice [127–129]. This lattice traps GF and allows gradual and steady release, which is ideal for the long-term regenerative process. The characteristics of PRF have been well documented, for example, in the Hassan et al. study, where PRF showed improvements in skin texture and patient satisfaction in a more stable manner than PRP. PRF, because of its structure, not only promotes tissue healing but also the formation of new blood vessels and controlled inflammation, key elements for complete skin regeneration. In addition, recent studies have proposed a further evolution of PRF, CGF (Concentrated Growth Factors), which optimizes the concentration of GF and may represent one of the most promising alternatives in the regenerative field. Although PRP is a viable option for skin regeneration, PRF may offer additional benefits due to its fibrin matrix and sustained release of GF. This suggests that PRF, or even CGF, may become a preferred choice for long-term treatments requiring more robust and durable regeneration [130–134].

Other regenerative techniques with GF like LPCGF, APGF (Autologous Platelet and Growth Factors), and microneedling are innovative tools that naturally stimulate tissue repair and enhance esthetic and functional results [135–137]. These methods use the patient's own biological materials to promote tissue regeneration, offering a synergistic approach that visibly improves appearance while supporting tissue health. LPCGF adds antimicrobial and anti-inflammatory benefits, making it ideal for complex healing scenarios. APGF further stimulates tissue renewal and improves skin texture, enhancing both recovery speed and quality in esthetic and post-surgical treatments. Microneedling, on the other hand, creates micro-perforations in the skin that boost collagen and elastin production, refining skin texture and complementing GF treatments for a maximized regenerative effect. Together, these techniques provide a natural, effective approach to comprehensive tissue rejuvenation [81,134,138].

4.1. PRP for Facial Rejuvenation

PRP has gained wide acceptance in the last few years as an instrument of treatment because of its non-invasive action for skin rejuvenation. It enhances the quality of the skin, minimizes wrinkles, and thus triggers regeneration of tissues. PRP, being prepared from the patient's autologous blood, is rich in GF, which has a stimulatory effect on tissue repair and collagen production. Although there is already a fair amount of literature on its support, more research is necessary for the elucidation of standards of treatment and long-term results as well as comparison with other options of regenerative therapy [139–141].

One of the recent studies relating to this approach toward PRP was conducted by Hassan et al. They investigated the PRP variant (Platelet-Rich Fibrin) for skin spot improvement, skin texture, and patient satisfaction; they found that while changes in wrinkle reduction were not statistically significant, the skin spot improvement was excellent. Such outcomes suggest that other blood-derived treatments, like PRF, do in fact show similar yet not identical benefits when compared to PRP and reinforce a need for systematic comparisons among these methods [142–144].

For facial rejuvenation, the research made by Draelos et al. examined the innovative use of autologous PRP in a topical cream over an eight-week period with 20 healthy participants aged 30 to 60 years assessing the effects of a PRP-containing serum compared to a serum without PRP [145,146]. The study showed that PRP remained stable for 90 days when refrigerated, highlighting its potential for topical use, and histological analysis revealed enhanced skin structure with increased collagen type I levels after applying the PRP serum. Despite these findings, both the PRP serum and a non-PRP serum demonstrated significant improvements in skin texture, raising questions about the unique benefits of PRP in this context. The treatments were well tolerated, and electroporation was used to improve absorption, addressing a challenge in topical PRP application. Future studies should aim to extend treatment duration and explore long-term efficacy, as the results suggest potential for PRP in skin rejuvenation but need further validation [147–149].

At the molecular level, Du et al. investigated the mechanisms of the effects of PRP. The authors demonstrated that injections of PRP may minimize the injury from aging and photoexposure by enhancing the synthesis of such important structural proteins as fibrillin and tropoelastin. This *in vitro* model study further supported previous concepts about the activity of PRP in the protection of skin against the detrimental influence of UV rays and maintenance of epidermal structure [113,150,151].

An interesting comparison of PRP to ready-to-use GF was given by Gawdat et al. In this split-face study, where each side of a patient's face received different treatments, it emerged that PRP yielded more long-lasting results than ready-to-use GF, with a higher level of satisfaction in the PRP-treated patients [152,153]. This finding points out the fact that even though PRP might take more time to prepare than GF, more benefits might be yielded by PRP [154,155].

In the pilot study by Lee et al., the effectiveness of PRP for treating photodamaged skin was evaluated with 31 participants aged 27 to 71 over a follow-up period of 5.7 weeks. The study found only one patient showing improvement based on the Wrinkle Severity Rating Scale, while the Global Aesthetic Improvement Scale indicated esthetic enhancement in 14 patients, although 17 patients saw no change. Despite this, patient satisfaction was high, with 74.2% pleased with the results, and 80% believing the treatment was worth it, feeling an average of 19.2 months younger. Mild adverse effects like tenderness and swelling were noted but did not significantly impact overall satisfaction. The study concluded that while PRP shows promise in rejuvenating photodamaged skin, further research is necessary to improve preparation methods and optimize results for broader clinical application [156–158].

Luiz Charles-de-Sá et al.'s comparative study, in 2020, evaluated the effects of Platelet-Rich Plasma (PRP) and expanded Adipose-Derived Stem Cell (ADSC) therapy on aged human skin. PRP injections induced an inflammatory response that increased collagen fiber deposits and thickened the dermis but did not contribute significantly to tissue regeneration [159–161]. In contrast, ADSC therapy led to extracellular matrix (ECM) remodeling, increased production of new elastic fibers, and degradation of elastosis, resulting in significant skin regeneration. The findings suggest that while PRP has limited rejuvenative properties, ADSC therapy shows considerable potential as an effective treatment for skin rejuvenation [162,163].

Randomized trials by Murad Alam et al. highlighted the disparities between the perception of patients and dermatologist assessment. While in 2018 participants perceived areas treated with PRP had significantly improved compared to areas treated with saline, dermatologists saw no apparent differences. These findings are supported by a further experiment, carried out by the same authors, which demonstrated how patient perceptions

of PRP's effects may be stronger than those of objective clinical evaluation. This is a crucial finding for any evaluation of PRP's effectiveness.

The aim of the clinical study by Paweł Surowiak et al. (2023) [115] was to analyze the effects of PRP on collagen production in human skin, specifically on type I procollagen synthesis. The methodology involved injecting 1 mL of autologous PRP into the skin of each participant's left forearm, while a placebo (0.9 percent sodium chloride) was injected into the right forearm. The results showed that PRP significantly stimulated the expression of type I procollagen. In contrast, placebo showed minimal collagen production. Interestingly, no correlation was found between patients' age, sex, or menopausal status and PRP response, although the small sample size may limit these results. No adverse effects were observed.

Finally, in this observational study, Ulusal et al. present the efficacy of Platelet-Rich Plasma treatment combined with hyaluronic acid, demonstrating that Platelet-Rich Plasma treatment significantly improves skin texture and firmness, with quite a high grade of patient satisfaction, especially after three or more treatments. Possible synergistic effects after PRP combination with other regenerative substances should be underscored [121].

In summary, the reviewed articles eloquently point out that PRP is a promising modality for skin rejuvenation since it improves texture, reduces wrinkling, and protects against photoaging.

Anyway, there are some major obstacles that need to be overcome: variability in results, non-uniformity in treatment protocols, and further research that needs to be conducted to define the molecular mechanisms underlying the effects of PRP. This is further put into comparison with other regenerative therapies, such as stem cells and GF; it suggests that PRP, even though it may not be the final answer for skin regeneration, can nevertheless become a useful component in combined treatments. In conclusion, PRP provides some hope in the field of skin rejuvenation but needs to be further researched in order to maximize benefits and improve treatment protocols [164,165].

4.2. PRP for Photoaging

Facial rejuvenation is a central theme in esthetic plastic surgery, as is the study of the various mechanisms underlying skin aging. Skin aging is characterized by the degradation of the extracellular matrix (ECM), with loss of elasticity due to the breakdown of fibers such as collagen and elastin and the decrease in fibroblasts. Research of Everts P and Coll. on the treatment of skin aging has explored the use of PRP (Platelet-Rich Plasma), particularly for its GF that support tissue repair. However, the lack of standardized protocols to produce PRP has led to conflicting data [166]. PRP is used in medicine to improve wound healing, with mechanisms parallel to photoaging and skin regeneration; however, the formulation used seems to be decisive on the therapeutic efficacy, as well as its use in combination with other preparations [167–172].

Formulations such as PRP buffy coat show superior results thanks to the high concentration of platelets and leukocytes. PRP works via PGF (platelet growth factors) that regulate cell proliferation and ECM remodeling. The use of Pure PRP has shown significant improvements in facial skin after three injections, reducing wrinkles and brown spots, with high patient satisfaction [173]. However, further studies with detailed platelet and white blood cell analyses would be needed to confirm these results [119,173].

A randomized trial by Tsai Y. and Coll. examined the effects of PRP (Platelet-Rich Plasma) and PPP (Platelet-Poor Plasma) on facial rejuvenation, showing significant improvement in periocular wrinkles from 3 to 6 months, with a 20% increase in MFWS (modified Fitzpatrick wrinkle score) in three months [174,175]. PRP did not improve nasolabial folds, while PPP showed a slight, non-significant improvement. PRP improved skin smoothness and texture, while PPP had immediate and long-lasting effects on fine

lines, with greater efficacy on dark circles and infraorbital folds [176,177]. PRP-stimulated fibroblasts and modulated inflammation, while PPP acted as a scaffold for the sustained release of GF. Both treatments were safe and well tolerated, with mild and transient adverse events. Limitations included the small sample size and a six-month observation period, indicating the need for further research [109,178].

A study by Ulusal B. has been conducted combining PRP with hyaluronic acid (HA), which showed that PRP can improve skin elasticity by removing photodamaged components and stimulating collagen synthesis. HA, due to its ability to retain water, is used in cosmetic treatments and supports dermal regeneration [179]. Combining PRP and HA, the esthetic results were more satisfactory than the single treatments. Combined treatments reduce pain and improve efficacy due to the viscous environment created by HA [180]. Repeated injections improve facial rejuvenation without surgery [181]. The combination with acupuncture promotes the formation of collagen and elastin, supported by the intake of vitamin C. The injection of PRP and HA after Botox reduces the duration of Botox itself, an effect still to be explained [182]. This confirms the effectiveness of biostimulation with PRP and HA for facial rejuvenation [183–185].

The combination of PRP with fractional, ultrapulsed CO₂ laser treatments seems to reduce laser-related side effects, especially in Asian populations with darker skin, as shown in the study of Hui Q and Coll. PRP can reduce these effects, accelerating healing and improving angiogenesis thanks to its GF, such as VEGF, PDGF, and TGF- β . The synergistic effect between laser and PRP promotes dermal remodeling and epidermal regrowth. Studies have shown that combined treatment is more effective and leads to a faster recovery than laser alone [186,187]. However, high expectations can affect patient satisfaction, especially in more severe cases [188]. PRP is a promising option to improve the efficacy of laser treatment, although further research is needed to optimize clinical protocols [121,189,190].

Although clinical evidence, like in the studies of da Silva L and Coll., seems to show the efficacy of PRP, the lack of standardized protocols for PRP production has led to conflicting data [191,192]. A non-randomized controlled trial compares the use of lyophilized PRP with saline, using objective techniques such as VISIA® imaging to evaluate the effect on photoaging. No significant improvements in collagen levels or skin aging were observed. The results differ from previous studies on fresh PRP, indicating the need for more methodological and well-designed studies [193,194]. The importance of the number of applications and the evaluation period is critical to evaluate the efficacy of lyophilized PRP [195]. Additionally, lyophilization may affect cellular interactions essential for its functionality, which requires further research to fully understand the clinical benefits of lyophilized PRP [196–203].

Our study identified several limitations in using PRP treatments. One key challenge is the variability in PRP preparation, as differences in centrifugation protocols can affect the concentration of platelets and growth factors, leading to inconsistent results. Additionally, many studies had small sample sizes, limiting the generalizability of findings. Patient factors such as age and skin type also influence outcomes but were not always controlled. Although PRP is generally safe, mild side effects like swelling and bruising are common, and rare complications such as infection can occur, especially with improper technique. Furthermore, the lack of standardized protocols for PRP preparation and injection methods makes it difficult to achieve consistent results across studies.

In conclusion, while PRP shows promise, further research is needed to optimize protocols, ensure safety, and address variability in treatment outcomes.

5. Conclusions

Autologous platelet concentrates (PCs), such as PRP and PRF, are promising emerging therapies for skin rejuvenation and anti-aging treatments due to their ability to stimulate collagen production, improve skin texture, and promote tissue repair. Among these, PRF stands out for its sustained release of growth factors, potentially offering more stable and lasting results compared to PRP. Additionally, the combination of PRP with complementary treatments, such as hyaluronic acid or laser therapies, enhances its rejuvenation effects. However, significant challenges remain, including variability in outcomes, lack of standardized protocols, and the need for further research to optimize clinical applications. Overall, PCs present significant potential in skin regeneration, but further studies are crucial to refine protocols, establish efficacy, and integrate these therapies into broader regenerative treatment approaches.

Author Contributions: Conceptualization, F.I., G.M., A.D.I., A.M.I., G.L., A.P., I.T., L.F. and P.N.; methodology, A.M.I. and G.M.; software, M.L., F.I. and A.D.I.; validation, F.I., G.D. and G.M.; formal analysis, M.L. and A.P.; investigation, A.P., I.T. and L.F.; resources, G.L., A.M.I., G.M. and A.M.I.; data curation, G.L., A.D.I., A.M.I. and F.I.; writing—original draft preparation, L.F., I.T. and P.N.; writing—review and editing, M.L., P.N. and G.D.; visualization, G.M.; supervision, L.F., G.L., I.T., P.N. and F.I.; project administration, F.I. All authors have read and agreed to the published version of the manuscript.

Funding: Prot. Number: 00152571, 15/02/2023 JAOUCPG23ICOMETIP; Study number: 7593, approved date: 25 January 2023; U.O. di Odontostomatologia; CODE: EAP AS; Principal investigator: Prof. F. Inchingolo, U.O. di Odontostomatologia, University Polyclinic of Bari.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing is not applicable to this article as no new data were created or analyzed in this study.

Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviations

ADSC	Adipose-Derived Stem Cell
AGFs	Autologous Growth Factors
APAG	Activated Plasma Albumin Gel
APGF	Autologous Platelet and Growth Factors
BC	Buffy Coat
bFGF	Basic Fibroblast Growth Factor
CGF	Concentrated Growth Factors
ECM	Extracellular Matrix
EGF	Epithelial Growth Factor
GF	Growth Factor
HA	Hyaluronic Acid
I-PRF	Injectable Platelet-Rich Fibrin
LPCGF	Leukocyte and Platelet-Concentrated Growth Factors
MMP	Matrix Metalloproteinases
NaCl	Sodium Chloride
PC	Platelet Concentrates
PDGF	Platelet-derived Growth Factors

PPP	Platelet Poor Plasma
PRF	Platelet-Rich Fibrin
PRGF	Plasma Rich in Growth Factor
PRP	Platelet-Rich Plasma
RBCs	Red Blood Cells
RCF	Relative Centrifugal Force
RP	Red Part
TGF	Transforming Growth Factor
UV	Ultraviolet
VEGF	Vascular Endothelial Growth Factor
WP	White Part

References

- Chung, J.H.; Eun, H.C. Angiogenesis in Skin Aging and Photoaging. *J. Dermatol.* **2007**, *34*, 593–600. [CrossRef] [PubMed]
- El-Domyati, M.; Abdel-Wahab, H.; Hossam, A. Combining Microneedling with Other Minimally Invasive Procedures for Facial Rejuvenation: A Split-Face Comparative Study. *Int. J. Dermatol.* **2018**, *57*, 1324–1334. [CrossRef] [PubMed]
- Kakudo, N.; Morimoto, N.; Kushida, S.; Ogawa, T.; Kusumoto, K. Platelet-Rich Plasma Releasate Promotes Angiogenesis in Vitro and in Vivo. *Med. Mol. Morphol.* **2014**, *47*, 83–89. [CrossRef] [PubMed]
- Li, Z.J.; Choi, H.-I.; Choi, D.-K.; Sohn, K.-C.; Im, M.; Seo, Y.-J.; Lee, Y.-H.; Lee, J.-H.; Lee, Y. Autologous Platelet-Rich Plasma: A Potential Therapeutic Tool for Promoting Hair Growth. *Dermatol. Surg.* **2012**, *38*, 1040–1046. [CrossRef]
- Uitto, J. The Role of Elastin and Collagen in Cutaneous Aging: Intrinsic Aging versus Photoexposure. *J. Drugs Dermatol.* **2008**, *7*, s12–s16.
- Wan, D.; Amirlak, B.; Giessler, P.; Rasko, Y.; Rohrich, R.J.; Yuan, C.; Lysikowski, J.; Delgado, I.; Davis, K. The Differing Adipocyte Morphologies of Deep versus Superficial Midfacial Fat Compartments: A Cadaveric Study. *Plast. Reconstr. Surg.* **2014**, *133*, 615e–622e. [CrossRef]
- Fujimura, T.; Hotta, M. The Preliminary Study of the Relationship between Facial Movements and Wrinkle Formation. *Skin Res. Technol.* **2012**, *18*, 219–224. [CrossRef]
- Chung, H.; Goo, B.; Lee, H.; Roh, M.; Chung, K. Enlarged Pores Treated with a Combination of Q-Switched and Micropulsed 1064 Nm Nd:YAG Laser with and without Topical Carbon Suspension: A Simultaneous Split-Face Trial. *Laser Ther.* **2011**, *20*, 181–188. [CrossRef]
- Varani, J.; Warner, R.L.; Gharaee-Kermani, M.; Phan, S.H.; Kang, S.; Chung, J.; Wang, Z.; Datta, S.C.; Fisher, G.J.; Voorhees, J.J. Vitamin A Antagonizes Decreased Cell Growth and Elevated Collagen-Degrading Matrix Metalloproteinases and Stimulates Collagen Accumulation in Naturally Aged Human Skin1. *J. Investigig. Dermatol.* **2000**, *114*, 480–486. [CrossRef]
- Kakudo, N.; Minakata, T.; Mitsui, T.; Kushida, S.; Notodihardjo, F.Z.; Kusumoto, K. Proliferation-Promoting Effect of Platelet-Rich Plasma on Human Adipose-Derived Stem Cells and Human Dermal Fibroblasts. *Plast. Reconstr. Surg.* **2008**, *122*, 1352–1360. [CrossRef]
- Kim, D.H.; Je, Y.J.; Kim, C.D.; Lee, Y.H.; Seo, Y.J.; Lee, J.H.; Lee, Y. Can Platelet-Rich Plasma Be Used for Skin Rejuvenation? Evaluation of Effects of Platelet-Rich Plasma on Human Dermal Fibroblast. *Ann. Dermatol.* **2011**, *23*, 424–431. [CrossRef] [PubMed]
- Cho, S.B.; Lee, J.H.; Choi, M.J.; Lee, K.-Y.; Oh, S.H. Efficacy of the Fractional Photothermolysis System with Dynamic Operating Mode on Acne Scars and Enlarged Facial Pores. *Dermatol. Surg.* **2009**, *35*, 108–114. [CrossRef] [PubMed]
- Chung, J.H.; Seo, J.Y.; Choi, H.R.; Lee, M.K.; Youn, C.S.; Rhie, G.; Cho, K.H.; Kim, K.H.; Park, K.C.; Eun, H.C. Modulation of Skin Collagen Metabolism in Aged and Photoaged Human Skin in Vivo. *J. Investigig. Dermatol.* **2001**, *117*, 1218–1224. [CrossRef]
- Chung, J.H.; Yano, K.; Lee, M.K.; Youn, C.S.; Seo, J.Y.; Kim, K.H.; Cho, K.H.; Eun, H.C.; Detmar, M. Differential Effects of Photoaging vs Intrinsic Aging on the Vascularization of Human Skin. *Arch. Dermatol.* **2002**, *138*, 1437–1442. [CrossRef]
- Sadoghi, P.; Lohberger, B.; Aigner, B.; Kaltenegger, H.; Friesenbichler, J.; Wolf, M.; Sununu, T.; Leithner, A.; Vavken, P. Effect of Platelet-Rich Plasma on the Biologic Activity of the Human Rotator-Cuff Fibroblasts: A Controlled in Vitro Study. *J. Orthop. Res.* **2013**, *31*, 1249–1253. [CrossRef]
- Xian, L.J.; Chowdhury, S.R.; Bin Saim, A.; Idrus, R.B.H. Concentration-Dependent Effect of Platelet-Rich Plasma on Keratinocyte and Fibroblast Wound Healing. *Cytotherapy* **2015**, *17*, 293–300. [CrossRef]
- Mehryan, P.; Zartab, H.; Rajabi, A.; Pazhoohi, N.; Firooz, A. Assessment of Efficacy of Platelet-Rich Plasma (PRP) on Infraorbital Dark Circles and Crow’s Feet Wrinkles. *J. Cosmet. Dermatol.* **2014**, *13*, 72–78. [CrossRef]
- Neiva-Sousa, M.; Carracha, C.; da Silva, L.N.; Valejo Coelho, P. Does Platelet-Rich Plasma Promote Facial Rejuvenation? Revising the Latest Evidence in a Narrative Review. *J. Cutan. Aesthet. Surg.* **2023**, *16*, 263–269. [CrossRef]

19. Marinelli, G.; Inchingo, A.D.; Inchingo, A.M.; Malcangi, G.; Limongelli, L.; Montenegro, V.; Coloccia, G.; Laudadio, C.; Patano, A.; Inchingo, F.; et al. White Spot Lesions in Orthodontics: Prevention and Treatment. A Descriptive Review. *J. Biol. Regul. Homeost. Agents* **2021**, *35*, 227–240. [[CrossRef](#)]
20. Inchingo, A.D.; Cazzolla, A.P.; Di Cosola, M.; Greco Lucchina, A.; Santacroce, L.; Charitos, I.A.; Topi, S.; Malcangi, G.; Hazballa, D.; Scarano, A.; et al. The Integumentary System and Its Microbiota between Health and Disease. *J. Biol. Regul. Homeost. Agents* **2021**, *35*, 303–321. [[CrossRef](#)]
21. Scandurra, C.; Gasparro, R.; Dolce, P.; Bochicchio, V.; Muzii, B.; Sammartino, G.; Marenzi, G.; Maldonato, N.M. The Role of Cognitive and Non-Cognitive Factors in Dental Anxiety: A Mediation Model. *Eur. J. Oral Sci.* **2021**, *129*, e12793. [[CrossRef](#)] [[PubMed](#)]
22. Sammartino, G.; Marenzi, G.; Tammaro, L.; Bolognese, A.; Calignano, A.; Costantino, U.; Califano, L.; Mastrangelo, F.; Tetè, S.; Vittoria, V. Anti-Inflammatory Drug Incorporation into Polymeric Nano-Hybrids for Local Controlled Release. *Int. J. Immunopathol. Pharmacol.* **2005**, *18*, 55–62. [[PubMed](#)]
23. Lubkowska, A.; Dolegowska, B.; Banfi, G. Growth Factor Content in PRP and Their Applicability in Medicine. *J. Biol. Regul. Homeost. Agents* **2012**, *26*, 3S–22S. [[PubMed](#)]
24. Mazzocca, A.D.; McCarthy, M.B.R.; Chowaniec, D.M.; Dugdale, E.M.; Hansen, D.; Cote, M.P.; Bradley, J.P.; Romeo, A.A.; Arciero, R.A.; Beitzel, K. The Positive Effects of Different Platelet-Rich Plasma Methods on Human Muscle, Bone, and Tendon Cells. *Am. J. Sports Med.* **2012**, *40*, 1742–1749. [[CrossRef](#)]
25. Browning, S.R.; Weiser, A.M.; Woolf, N.; Golish, S.R.; SanGiovanni, T.P.; Scuderi, G.J.; Carballo, C.; Hanna, L.S. Platelet-Rich Plasma Increases Matrix Metalloproteinases in Cultures of Human Synovial Fibroblasts. *J. Bone Jt. Surg. Am.* **2012**, *94*, e172. [[CrossRef](#)]
26. Cho, H.S.; Song, I.H.; Park, S.-Y.; Sung, M.C.; Ahn, M.-W.; Song, K.E. Individual Variation in Growth Factor Concentrations in Platelet-Rich Plasma and Its Influence on Human Mesenchymal Stem Cells. *Korean J. Lab. Med.* **2011**, *31*, 212–218. [[CrossRef](#)]
27. Alsousou, J.; Thompson, M.; Hulley, P.; Noble, A.; Willett, K. The Biology of Platelet-Rich Plasma and Its Application in Trauma and Orthopaedic Surgery: A Review of the Literature. *J. Bone Jt. Surg. Br.* **2009**, *91*, 987–996. [[CrossRef](#)]
28. Edelblute, C.M.; Donate, A.L.; Hargrave, B.Y.; Heller, L.C. Human Platelet Gel Supernatant Inactivates Opportunistic Wound Pathogens on Skin. *Platelets* **2015**, *26*, 13–16. [[CrossRef](#)]
29. Nicoli, F.; Balzani, A.; Lazzeri, D.; Gentile, P.; Chilgar, R.M.; Di Pasquali, C.; Nicoli, M.; Bocchini, I.; Agovino, A.; Cervelli, V. Severe Hidradenitis Suppurativa Treatment Using Platelet-Rich Plasma Gel and Hyalomatrix. *Int. Wound J.* **2015**, *12*, 338–343. [[CrossRef](#)]
30. Hou, X.; Yuan, J.; Aisaiti, A.; Liu, Y.; Zhao, J. The Effect of Platelet-Rich Plasma on Clinical Outcomes of the Surgical Treatment of Periodontal Intrabony Defects: A Systematic Review and Meta-Analysis. *BMC Oral Health* **2016**, *16*, 71. [[CrossRef](#)]
31. Lavker, R.M.; Zheng, P.S.; Dong, G. Aged Skin: A Study by Light, Transmission Electron, and Scanning Electron Microscopy. *J. Investig. Dermatol.* **1987**, *88*, 44s–51s. [[CrossRef](#)] [[PubMed](#)]
32. Montagna, W.; Carlisle, K. Structural Changes in Aging Human Skin. *J. Investig. Dermatol.* **1979**, *73*, 47–53. [[CrossRef](#)] [[PubMed](#)]
33. West, M.D.; Pereira-Smith, O.M.; Smith, J.R. Replicative Senescence of Human Skin Fibroblasts Correlates with a Loss of Regulation and Overexpression of Collagenase Activity. *Exp. Cell Res.* **1989**, *184*, 138–147. [[CrossRef](#)] [[PubMed](#)]
34. Draelos, Z.D.; Rheins, L.A.; Wootten, S.; Kellar, R.S.; Diller, R.B. Pilot Study: Autologous Platelet-Rich Plasma Used in a Topical Cream for Facial Rejuvenation. *J. Cosmet. Dermatol.* **2019**, *18*, 1348–1352. [[CrossRef](#)]
35. Eppley, B.L.; Pietrzak, W.S.; Blanton, M. Platelet-Rich Plasma: A Review of Biology and Applications in Plastic Surgery. *Plast. Reconstr. Surg.* **2006**, *118*, 147e–159e. [[CrossRef](#)]
36. Fabi, S.; Sundaram, H. The Potential of Topical and Injectable Growth Factors and Cytokines for Skin Rejuvenation. *Facial Plast. Surg.* **2014**, *30*, 157–171. [[CrossRef](#)]
37. Panda, S.; Doraiswamy, J.; Malaiappan, S.; Varghese, S.S.; Del Fabbro, M. Additive Effect of Autologous Platelet Concentrates in Treatment of Intrabony Defects: A Systematic Review and Meta-Analysis. *J. Investig. Clin. Dent.* **2016**, *7*, 13–26. [[CrossRef](#)]
38. Ilgenli, T.; Dündar, N.; Kal, B.I. Demineralized Freeze-Dried Bone Allograft and Platelet-Rich Plasma vs Platelet-Rich Plasma Alone in Infrabony Defects: A Clinical and Radiographic Evaluation. *Clin. Oral Investig.* **2007**, *11*, 51–59. [[CrossRef](#)]
39. Yamamiya, K.; Okuda, K.; Kawase, T.; Hata, K.-I.; Wolff, L.F.; Yoshie, H. Tissue-Engineered Cultured Periosteum Used with Platelet-Rich Plasma and Hydroxyapatite in Treating Human Osseous Defects. *J. Periodontol.* **2008**, *79*, 811–818. [[CrossRef](#)]
40. Del Corso, M.; Verville, A.; Simonpieri, A.; Jimbo, R.; Inchingo, F.; Sammartino, G.; Dohan Ehrenfest, D.M. Current Knowledge and Perspectives for the Use of Platelet-Rich Plasma (PRP) and Platelet-Rich Fibrin (PRF) in Oral and Maxillofacial Surgery Part 1: Periodontal and Dentoalveolar Surgery. *Curr. Pharm. Biotechnol.* **2012**, *13*, 1207–1230. [[CrossRef](#)]
41. Hsu, W.K.; Mishra, A.; Rodeo, S.R.; Fu, F.; Terry, M.A.; Randelli, P.; Canale, S.T.; Kelly, F.B. Platelet-Rich Plasma in Orthopaedic Applications: Evidence-Based Recommendations for Treatment. *J. Am. Acad. Orthop. Surg.* **2013**, *21*, 739–748. [[CrossRef](#)] [[PubMed](#)]

42. Mehta, V.; Fiorillo, L.; Langaliya, A.; Obulareddy, V.T.; Cicciu, M. The Effect of Xenograft and Platelet-Rich Plasma in the Surgical Management of Intrabony Defects in Periodontitis Patients: A Systematic Review. *J. Craniofacial Surg.* **2023**, *34*, 2222–2227. [[CrossRef](#)]
43. Dang, Q.T.; Huynh, T.D.; Inchlingolo, F.; Dipalma, G.; Inchlingolo, A.D.; Cantore, S.; Paduanelli, G.; Nguyen, K.C.D.; Ballini, A.; Isacco, C.G.; et al. Human Chondrocytes from Human Adipose Tissue-Derived Mesenchymal Stem Cells Seeded on a Dermal-Derived Collagen Matrix Sheet: Our Preliminary Results for a Ready to Go Biotechnological Cartilage Graft in Clinical Practice. *Stem Cells Int.* **2021**, *2021*, 6664697. [[CrossRef](#)] [[PubMed](#)]
44. Anitua, E. Plasma Rich in Growth Factors: Preliminary Results of Use in the Preparation of Future Sites for Implants. *Int. J. Oral Maxillofac. Implants* **1999**, *14*, 529–535. [[PubMed](#)]
45. Anitua, E.; Pelacho, B.; Prado, R.; Aguirre, J.J.; Sánchez, M.; Padilla, S.; Aranguren, X.L.; Abizanda, G.; Collantes, M.; Hernandez, M.; et al. Infiltration of Plasma Rich in Growth Factors Enhances in Vivo Angiogenesis and Improves Reperfusion and Tissue Remodeling after Severe Hind Limb Ischemia. *J. Control Release* **2015**, *202*, 31–39. [[CrossRef](#)]
46. Dohan, D.M.; Choukroun, J.; Diss, A.; Dohan, S.L.; Dohan, A.J.J.; Mouhyi, J.; Gogly, B. Platelet-Rich Fibrin (PRF): A Second-Generation Platelet Concentrate. Part III: Leucocyte Activation: A New Feature for Platelet Concentrates? *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endodontol.* **2006**, *101*, e51–e55. [[CrossRef](#)]
47. Dohan Ehrenfest, D.M.; Andia, I.; Zumstein, M.A.; Zhang, C.-Q.; Pinto, N.R.; Bielecki, T. Classification of Platelet Concentrates (Platelet-Rich Plasma-PRP, Platelet-Rich Fibrin-PRF) for Topical and Infiltrative Use in Orthopedic and Sports Medicine: Current Consensus, Clinical Implications and Perspectives. *Muscles Ligaments Tendons J.* **2014**, *4*, 3–9. [[CrossRef](#)]
48. Hassan, H.; Quinlan, D.J.; Ghanem, A. Injectable Platelet-Rich Fibrin for Facial Rejuvenation: A Prospective, Single-Center Study. *J. Cosmet. Dermatol.* **2020**, *19*, 3213–3221. [[CrossRef](#)]
49. Agarwal, K.; Agarwal, K.; Kumar, N. Platelet Concentrates: Regenerating the Lost Tissues. *J. Pharm. Bioallied Sci.* **2013**, *5*, 329–330. [[CrossRef](#)]
50. Kobayashi, M.; Kawase, T.; Horimizu, M.; Okuda, K.; Wolff, L.F.; Yoshie, H. A Proposed Protocol for the Standardized Preparation of PRF Membranes for Clinical Use. *Biologicals* **2012**, *40*, 323–329. [[CrossRef](#)]
51. Isobe, K.; Watanabe, T.; Kawabata, H.; Kitamura, Y.; Okudera, T.; Okudera, H.; Uematsu, K.; Okuda, K.; Nakata, K.; Tanaka, T.; et al. Mechanical and Degradation Properties of Advanced Platelet-Rich Fibrin (A-PRF), Concentrated Growth Factors (CGF), and Platelet-Poor Plasma-Derived Fibrin (PPTF). *Int. J. Implant. Dent.* **2017**, *3*, 17. [[CrossRef](#)] [[PubMed](#)]
52. Laforgia, A.; Inchlingolo, A.D.; Piras, F.; Colonna, V.; Giorgio, R.V.; Carone, C.; Rapone, B.; Malcangi, G.; Inchlingolo, A.M.; Inchlingolo, F.; et al. Therapeutic Strategies and Genetic Implications for Periodontal Disease Management: A Systematic Review. *Int. J. Mol. Sci.* **2024**, *25*, 7217. [[CrossRef](#)] [[PubMed](#)]
53. Rodella, L.F.; Favero, G.; Boninsegna, R.; Buffoli, B.; Labanca, M.; Scari, G.; Sacco, L.; Batani, T.; Rezzani, R. Growth Factors, CD34 Positive Cells, and Fibrin Network Analysis in Concentrated Growth Factors Fraction. *Microsc. Res. Tech.* **2011**, *74*, 772–777. [[CrossRef](#)] [[PubMed](#)]
54. Sahin, İ.O.; Gokmenoglu, C.; Kara, C. Effect of Concentrated Growth Factor on Osteoblast Cell Response. *J. Stomatol. Oral Maxillofac. Surg.* **2018**, *119*, 477–481. [[CrossRef](#)] [[PubMed](#)]
55. Contaldo, M.; Itro, A.; Lajolo, C.; Gioco, G.; Inchlingolo, F.; Serpico, R. Overview on Osteoporosis, Periodontitis and Oral Dysbiosis: The Emerging Role of Oral Microbiota. *Appl. Sci.* **2020**, *10*, 6000. [[CrossRef](#)]
56. Coloccia, G.; Inchlingolo, A.D.; Inchlingolo, A.M.; Malcangi, G.; Montenegro, V.; Patano, A.; Marinelli, G.; Laudadio, C.; Limongelli, L.; Di Venere, D.; et al. Effectiveness of Dental and Maxillary Transverse Changes in Tooth-Borne, Bone-Borne, and Hybrid Palatal Expansion through Cone-Beam Tomography: A Systematic Review of the Literature. *Medicina* **2021**, *57*, 288. [[CrossRef](#)]
57. Kuznetsova, I.P.; Rakhmanir, I.A.; Zvezotkina, L.S.; Lipovich, M.M. [Clinico-Roentgenologic Dynamics of Ewing's Sarcoma during Radiotherapy]. *Med. Radiol.* **1971**, *16*, 22–27.
58. Anitua, E.; Tierno, R.; Azkargorta, M.; Elortza, F.; Alkhraisat, M.H. Effect of Health Status and Heat-Induced Inactivation on the Proteomic Profile of Plasma Rich in Growth Factors Obtained from Donors with Chronic Inflammatory Skin Conditions. *Biomolecules* **2024**, *14*, 763. [[CrossRef](#)]
59. Jeyaraman, M.; Pai, S.N.; Filippo, M.; Jeyaraman, N.; Venkatasalam, R.; Nallakumarasamy, A.; Khanna, M.; Patro, B.P.; Sharma, S.; Rangarajan, R.V. Informed Consent Form for Platelet Rich Plasma Injections: Evidence-Based and Legally Guide for Orthopaedic Surgeons. *Eur. J. Med. Res.* **2024**, *29*, 422. [[CrossRef](#)]
60. Mercader-Ruiz, J.; Beitia, M.; Delgado, D.; Sánchez, P.; Porras, B.; Gimeno, I.; González, S.; Benito-Lopez, F.; Basabe-Desmonts, L.; Sánchez, M. Current Challenges in the Development of Platelet-Rich Plasma-Based Therapies. *Biomed. Res. Int.* **2024**, *2024*, 6444120. [[CrossRef](#)]
61. Abdel-Maguid, E.M.; Awad, S.M.; Hassan, Y.S.; El-Mokhtar, M.A.; EL-Deek, H.E.; Mekkawy, M.M. Efficacy of Stem Cell-Conditioned Medium vs. Platelet-Rich Plasma as an Adjuvant to Ablative Fractional CO₂ Laser Resurfacing for Atrophic Post-Acne Scars: A Split-Face Clinical Trial. *J. Dermatol. Treat.* **2021**, *32*, 242–249. [[CrossRef](#)] [[PubMed](#)]

62. Beeson, W.; Woods, E.; Agha, R. Tissue Engineering, Regenerative Medicine, and Rejuvenation in 2010: The Role of Adipose-Derived Stem Cells. *Facial Plast. Surg.* **2011**, *27*, 378–387. [CrossRef] [PubMed]
63. Harrison, P.; Cramer, E.M. Platelet Alpha-Granules. *Blood Rev.* **1993**, *7*, 52–62. [CrossRef] [PubMed]
64. Garcia, B.A.; Smalley, D.M.; Cho, H.; Shabanowitz, J.; Ley, K.; Hunt, D.F. The Platelet Microparticle Proteome. *J. Proteome Res.* **2005**, *4*, 1516–1521. [CrossRef]
65. Yu, P.; Zhai, Z.; Jin, X.; Yang, X.; Qi, Z. Clinical Application of Platelet-Rich Fibrin in Plastic and Reconstructive Surgery: A Systematic Review. *Aesthetic Plast. Surg.* **2018**, *42*, 511–519. [CrossRef]
66. Wang, X.; Yang, Y.; Zhang, Y.; Miron, R.J. Fluid Platelet-Rich Fibrin Stimulates Greater Dermal Skin Fibroblast Cell Migration, Proliferation, and Collagen Synthesis When Compared to Platelet-Rich Plasma. *J. Cosmet. Dermatol.* **2019**, *18*, 2004–2010. [CrossRef]
67. Wend, S.; Kubesch, A.; Orlowska, A.; Al-Maawi, S.; Zender, N.; Dias, A.; Miron, R.J.; Sader, R.; Booms, P.; Kirkpatrick, C.J.; et al. Reduction of the Relative Centrifugal Force Influences Cell Number and Growth Factor Release within Injectable PRF-Based Matrices. *J. Mater. Sci. Mater. Med.* **2017**, *28*, 188. [CrossRef]
68. Dohan Ehrenfest, D.M.; Bielecki, T.; Jimbo, R.; Barbe, G.; Del Corso, M.; Inchingolo, F.; Sammartino, G. Do the Fibrin Architecture and Leukocyte Content Influence the Growth Factor Release of Platelet Concentrates? An Evidence-Based Answer Comparing a Pure Platelet-Rich Plasma (P-PRP) Gel and a Leukocyte- and Platelet-Rich Fibrin (L-PRF). *Curr. Pharm. Biotechnol.* **2012**, *13*, 1145–1152. [CrossRef]
69. Masuki, H.; Okudera, T.; Watanebe, T.; Suzuki, M.; Nishiyama, K.; Okudera, H.; Nakata, K.; Uematsu, K.; Su, C.-Y.; Kawase, T. Growth Factor and Pro-Inflammatory Cytokine Contents in Platelet-Rich Plasma (PRP), Plasma Rich in Growth Factors (PRGF), Advanced Platelet-Rich Fibrin (A-PRF), and Concentrated Growth Factors (CGF). *Int. J. Implant. Dent.* **2016**, *2*, 19. [CrossRef]
70. Everts, P.A.; Pinto, P.C.; Girão, L. Autologous Pure Platelet-Rich Plasma Injections for Facial Skin Rejuvenation: Biometric Instrumental Evaluations and Patient-Reported Outcomes to Support Antiaging Effects. *J. Cosmet. Dermatol.* **2019**, *18*, 985–995. [CrossRef]
71. Camargo, P.M.; Lekovic, V.; Weinlaender, M.; Vasilic, N.; Madzarevic, M.; Kenney, E.B. Platelet-Rich Plasma and Bovine Porous Bone Mineral Combined with Guided Tissue Regeneration in the Treatment of Intrabony Defects in Humans. *J. Periodontal Res.* **2002**, *37*, 300–306. [CrossRef] [PubMed]
72. Pradeep, A.R.; Rao, N.S.; Agarwal, E.; Bajaj, P.; Kumari, M.; Naik, S.B. Comparative Evaluation of Autologous Platelet-Rich Fibrin and Platelet-Rich Plasma in the Treatment of 3-Wall Intrabony Defects in Chronic Periodontitis: A Randomized Controlled Clinical Trial. *J. Periodontol.* **2012**, *83*, 1499–1507. [CrossRef] [PubMed]
73. Yilmaz, S.; Cakar, G.; Kuru, B.; Dirikan, S.; Yildirim, B. Platelet-Rich Plasma in Combination with Bovine Derived Xenograft in the Treatment of Deep Intrabony Periodontal Defects: A Report of 20 Consecutively Treated Patients. *Platelets* **2009**, *20*, 432–440. [CrossRef]
74. Camargo, P.M.; Lekovic, V.; Weinlaender, M.; Divnic-Resnik, T.; Pavlovic, M.; Kenney, E.B. A Surgical Reentry Study on the Influence of Platelet-Rich Plasma in Enhancing the Regenerative Effects of Bovine Porous Bone Mineral and Guided Tissue Regeneration in the Treatment of Intrabony Defects in Humans. *J. Periodontol.* **2009**, *80*, 915–923. [CrossRef]
75. Czuryszkiewicz-Cyrana, J.; Banach, J. Autogenous Bone and Platelet-Rich Plasma (PRP) in the Treatment of Intrabony Defects. *Adv. Med. Sci.* **2006**, *51* (Suppl. S1), 26–30.
76. Döri, F.; Nikolidakis, D.; Húszár, T.; Arweiler, N.B.; Gera, I.; Sculean, A. Effect of Platelet-Rich Plasma on the Healing of Intrabony Defects Treated with an Enamel Matrix Protein Derivative and a Natural Bone Mineral. *J. Clin. Periodontol.* **2008**, *35*, 44–50. [CrossRef]
77. Bonazza, V.; Borsani, E.; Buffoli, B.; Castrezzati, S.; Rezzani, R.; Rodella, L.F. How the Different Material and Shape of the Blood Collection Tube Influences the Concentrated Growth Factors Production. *Microsc. Res. Tech.* **2016**, *79*, 1173–1178. [CrossRef]
78. Miron, R.J.; Chai, J.; Zheng, S.; Feng, M.; Sculean, A.; Zhang, Y. A Novel Method for Evaluating and Quantifying Cell Types in Platelet Rich Fibrin and an Introduction to Horizontal Centrifugation. *J. Biomed. Mater. Res. A* **2019**, *107*, 2257–2271. [CrossRef]
79. Döri, F.; Arweiler, N.; Húszár, T.; Gera, I.; Miron, R.J.; Sculean, A. Five-Year Results Evaluating the Effects of Platelet-Rich Plasma on the Healing of Intrabony Defects Treated with Enamel Matrix Derivative and Natural Bone Mineral. *J. Periodontol.* **2013**, *84*, 1546–1555. [CrossRef]
80. Kassolis, J.D.; Reynolds, M.A. Evaluation of the Adjunctive Benefits of Platelet-Rich Plasma in Subantral Sinus Augmentation. *J. Craniofacial Surg.* **2005**, *16*, 280–287. [CrossRef]
81. Rullo, R.; Festa, V.M.; Rullo, F.; Trosino, O.; Cerone, V.; Gasparro, R.; Laino, L.; Sammartino, G. The Use of Piezosurgery in Genioplasty. *J. Craniofacial Surg.* **2016**, *27*, 414–415. [CrossRef] [PubMed]
82. Gasparro, R.; Sammartino, G.; Mariniello, M.; di Lauro, A.E.; Spagnuolo, G.; Marenzi, G. Treatment of Periodontal Pockets at the Distal Aspect of Mandibular Second Molar after Surgical Removal of Impacted Third Molar and Application of L-PRF: A Split-Mouth Randomized Clinical Trial. *Quintessence Int.* **2020**, *51*, 204–211. [CrossRef] [PubMed]

83. Del Amo, F.S.L.; Yu, S.-H.; Sammartino, G.; Sculean, A.; Zucchelli, G.; Rasperini, G.; Felice, P.; Pagni, G.; Iorio-Siciliano, V.; Grusovin, M.G.; et al. Peri-Implant Soft Tissue Management: Cairo Opinion Consensus Conference. *Int. J. Environ. Res. Public Health* **2020**, *17*, 2281. [CrossRef] [PubMed]
84. Canfora, F.; Calabria, E.; Cuocolo, R.; Ugga, L.; Buono, G.; Marenzi, G.; Gasparro, R.; Pecoraro, G.; Aria, M.; D’Aniello, L.; et al. Burning Fog: Cognitive Impairment in Burning Mouth Syndrome. *Front. Aging Neurosci.* **2021**, *13*, 727417. [CrossRef]
85. Tsai, Y.-W.; Cheng, C.-Y.; Hu, S.; Chang, S.-L.; Lin, T.-M.; Huang, Y.-L. Platelet-Rich Plasma Versus Platelet-Poor Plasma for Treating Facial Photoaging: A Double-Blind Randomized Controlled Splitting Face Study. *Aesthetic Plast. Surg.* **2024**, *48*, 2162–2170. [CrossRef]
86. Hesseler, M.J.; Shyam, N. Platelet-Rich Plasma and Its Utility in Medical Dermatology: A Systematic Review. *J. Am. Acad. Dermatol.* **2019**, *81*, 834–846. [CrossRef]
87. Innocenti, M.; Ramoni, S.; Doria, C.; Antropoli, C.; Garbagna, N.; Grossi, E.; Veraldi, S. Treatment of Periorcular Wrinkles with Topical Nifedipine. *J. Dermatol. Treat.* **2010**, *21*, 282–285. [CrossRef]
88. Gold, M.H.; Biron, J.A.; Sensing, W. Facial Skin Rejuvenation by Combination Treatment of IPL Followed by Continuous and Fractional Radiofrequency. *J. Cosmet. Laser Ther.* **2016**, *18*, 2–6. [CrossRef]
89. Camargo, P.M.; Lekovic, V.; Weinlaender, M.; Vasilic, N.; Madzarevic, M.; Kenney, E.B. A Reentry Study on the Use of Bovine Porous Bone Mineral, GTR, and Platelet-Rich Plasma in the Regenerative Treatment of Intrabony Defects in Humans. *Int. J. Periodontics Restor. Dent.* **2005**, *25*, 49–59.
90. Lekovic, V.; Camargo, P.M.; Weinlaender, M.; Vasilic, N.; Kenney, E.B. Comparison of Platelet-Rich Plasma, Bovine Porous Bone Mineral, and Guided Tissue Regeneration versus Platelet-Rich Plasma and Bovine Porous Bone Mineral in the Treatment of Intrabony Defects: A Reentry Study. *J. Periodontol.* **2002**, *73*, 198–205. [CrossRef]
91. Okuda, K.; Tai, H.; Tanabe, K.; Suzuki, H.; Sato, T.; Kawase, T.; Saito, Y.; Wolff, L.F.; Yoshiex, H. Platelet-Rich Plasma Combined with a Porous Hydroxyapatite Graft for the Treatment of Intrabony Periodontal Defects in Humans: A Comparative Controlled Clinical Study. *J. Periodontol.* **2005**, *76*, 890–898. [CrossRef] [PubMed]
92. Roselló-Camps, À.; Monje, A.; Lin, G.-H.; Khoshkam, V.; Chávez-Gatty, M.; Wang, H.-L.; Gargallo-Albiol, J.; Hernandez-Alfaro, F. Platelet-Rich Plasma for Periodontal Regeneration in the Treatment of Intrabony Defects: A Meta-Analysis on Prospective Clinical Trials. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol.* **2015**, *120*, 562–574. [CrossRef] [PubMed]
93. Malcangi, G.; Patano, A.; Palmieri, G.; Di Pede, C.; Latini, G.; Inchingolo, A.D.; Hazballa, D.; de Ruvo, E.; Garofoli, G.; Inchingolo, F.; et al. Maxillary Sinus Augmentation Using Autologous Platelet Concentrates (Platelet-Rich Plasma, Platelet-Rich Fibrin, and Concentrated Growth Factor) Combined with Bone Graft: A Systematic Review. *Cells* **2023**, *12*, 1797. [CrossRef] [PubMed]
94. Bonazza, V.; Borsani, E.; Buffoli, B.; Parolini, S.; Inchingolo, F.; Rezzani, R.; Rodella, L.F. In Vitro Treatment with Concentrated Growth Factors (CGF) and Sodium Orthosilicate Positively Affects Cell Renewal in Three Different Human Cell Lines. *Cell Biol. Int.* **2018**, *42*, 353–364. [CrossRef]
95. Borsani, E.; Buffoli, B.; Bonazza, V.; Brunelli, G.; Monini, L.; Inchingolo, F.; Ballini, A.; Rezzani, R.; Rodella, L.F. In Vitro Effects of Concentrated Growth Factors (CGF) on Human SH-SY5Y Neuronal Cells. *Eur. Rev. Med. Pharmacol. Sci.* **2020**, *24*, 304–314. [CrossRef]
96. Inchingolo, F.; Hazballa, D.; Inchingolo, A.D.; Malcangi, G.; Marinelli, G.; Mancini, A.; Maggiore, M.E.; Bordea, I.R.; Scarano, A.; Farronato, M.; et al. Innovative Concepts and Recent Breakthrough for Engineered Graft and Constructs for Bone Regeneration: A Literature Systematic Review. *Materials* **2022**, *15*, 1120. [CrossRef]
97. Dipalma, G.; Inchingolo, A.M.; Malcangi, G.; Ferrara, I.; Viapiano, F.; Netti, A.; Patano, A.; Isacco, C.G.; Inchingolo, A.D.; Inchingolo, F. Sixty-Month Follow Up of Clinical MRONJ Cases Treated with CGF and Piezosurgery. *Bioengineering* **2023**, *10*, 863. [CrossRef]
98. Inchingolo, F.; Inchingolo, A.M.; Malcangi, G.; De Leonardi, N.; Sardano, R.; Pezzolla, C.; de Ruvo, E.; Di Venere, D.; Palermo, A.; Inchingolo, A.D.; et al. The Benefits of Probiotics on Oral Health: Systematic Review of the Literature. *Pharmaceuticals* **2023**, *16*, 1313. [CrossRef]
99. Bonazza, V.; Hajistilly, C.; Patel, D.; Patel, J.; Woo, R.; Cocchi, M.A.; Buffoli, B.; Lancini, D.; Gheno, E.; Rezzani, R.; et al. Growth Factors Release from Concentrated Growth Factors: Effect of β -Tricalcium Phosphate Addition. *J. Craniofacial Surg.* **2018**, *29*, 2291–2295. [CrossRef]
100. Buffoli, B.; Rosi, S.; Borsani, E.; Rodella, L.F.; Mortellaro, C. Effect of Two Different Parts of CGF on Post-Extractive Alveolar Ridge Preservation: A Preliminary Histomorphometric Analysis in a Split-Mouth Design. *J. Biol. Regul. Homeost. Agents* **2021**, *35*, 155–161. [CrossRef]
101. Leonida, A.; Favero, G.; Caccianiga, P.; Ceraulo, S.; Rodella, L.F.; Rezzani, R.; Caccianiga, G. Concentrated Growth Factors (CGF) Combined with Melatonin in Guided Bone Regeneration (GBR): A Case Report. *Diagnostics* **2022**, *12*, 1257. [CrossRef] [PubMed]
102. D’Esposito, V.; Lecce, M.; Marenzi, G.; Cabaro, S.; Ambrosio, M.R.; Sammartino, G.; Misso, S.; Migliaccio, T.; Liguoro, P.; Oriente, F.; et al. Platelet-Rich Plasma Counteracts Detrimental Effect of High-Glucose Concentrations on Mesenchymal Stem Cells from Bichat Fat Pad. *J. Tissue Eng. Regen. Med.* **2020**, *14*, 701–713. [CrossRef] [PubMed]

103. Gasparro, R.; Qorri, E.; Valletta, A.; Masucci, M.; Sammartino, P.; Amato, A.; Marenzi, G. Non-Transfusional Hemocomponents: From Biology to the Clinic—A Literature Review. *Bioengineering* **2018**, *5*, 27. [[CrossRef](#)]
104. Vig, K.; Chaudhari, A.; Tripathi, S.; Dixit, S.; Sahu, R.; Pillai, S.; Dennis, V.; Singh, S. Advances in Skin Regeneration Using Tissue Engineering. *Int. J. Mol. Sci.* **2017**, *18*, 789. [[CrossRef](#)]
105. Liberati, A.; Altman, D.G.; Tetzlaff, J.; Mulrow, C.; Gøtzsche, P.C.; Ioannidis, J.P.A.; Clarke, M.; Devereaux, P.J.; Kleijnen, J.; Moher, D. The PRISMA Statement for Reporting Systematic Reviews and Meta-Analyses of Studies That Evaluate Health Care Interventions: Explanation and Elaboration. *J. Clin. Epidemiol.* **2009**, *62*, e1–e34. [[CrossRef](#)]
106. Malcangi, G.; Patano, A.; Ciocia, A.M.; Netti, A.; Viapiano, F.; Palumbo, I.; Trilli, I.; Guglielmo, M.; Inchingolo, A.D.; Dipalma, G.; et al. Benefits of Natural Antioxidants on Oral Health. *Antioxid. Basel Switz.* **2023**, *12*, 1309. [[CrossRef](#)]
107. Inchingolo, F.; Tatullo, M.; Pacifici, A.; Gargari, M.; Inchingolo, A.D.; Inchingolo, A.M.; Dipalma, G.; Marrelli, M.; Abenavoli, F.M.; Pacifici, L. Use of Dermal-Fat Grafts in the Post-Oncological Reconstructive Surgery of Atrophies in the Zygomatic Region: Clinical Evaluations in the Patients Undergone to Previous Radiation Therapy. *Head Face Med.* **2012**, *8*, 33. [[CrossRef](#)]
108. Inchingolo, F.; Inchingolo, A.M.; Piras, F.; Ferrante, L.; Mancini, A.; Palermo, A.; Inchingolo, A.D.; Dipalma, G. The Interaction between Gut Microbiome and Bone Health. *Curr. Opin. Endocrinol. Diabetes Obes.* **2024**, *31*, 122–130. [[CrossRef](#)]
109. Inchingolo, A.M.; Patano, A.; Piras, F.; Mancini, A.; Inchingolo, A.D.; Paduanelli, G.; Inchingolo, F.; Palermo, A.; Dipalma, G.; Malcangi, G. Interconnection between Microbiota-Gut-Brain Axis and Autism Spectrum Disorder Comparing Therapeutic Options: A Scoping Review. *Microorganisms* **2023**, *11*, 1477. [[CrossRef](#)]
110. Ulusal, B.G. Platelet-rich Plasma and Hyaluronic Acid—An Efficient Biostimulation Method for Face Rejuvenation. *J. Cosmet. Dermatol.* **2017**, *16*, 112–119. [[CrossRef](#)]
111. Inchingolo, F.; Dipalma, G.; Guglielmo, M.; Palumbo, I.; Campanelli, M.; Inchingolo, A.D.; de Ruvo, E.; Palermo, A.; Di Venere, D.; Inchingolo, A.M. Correlation between Vegetarian Diet and Oral Health: A Systematic Review. *Eur. Rev. Med. Pharmacol. Sci.* **2024**, *28*, 2127–2143. [[CrossRef](#)] [[PubMed](#)]
112. Inchingolo, F.; Inchingolo, A.D.; Latini, G.; Trilli, I.; Ferrante, L.; Nardelli, P.; Malcangi, G.; Inchingolo, A.M.; Mancini, A.; Palermo, A.; et al. The Role of Curcumin in Oral Health and Diseases: A Systematic Review. *Antioxidants* **2024**, *13*, 660. [[CrossRef](#)] [[PubMed](#)]
113. Du, R.; Lei, T. Effects of Autologous Platelet-rich Plasma Injections on Facial Skin Rejuvenation. *Exp. Ther. Med.* **2020**, *19*, 3024–3030. [[CrossRef](#)] [[PubMed](#)]
114. Gawdat, H.I.; Tawdy, A.M.; Hegazy, R.A.; Zakaria, M.M.; Allam, R.S. Autologous Platelet-rich Plasma versus Readymade Growth Factors in Skin Rejuvenation: A Split Face Study. *J. Cosmet. Dermatol.* **2017**, *16*, 258–264. [[CrossRef](#)]
115. Lee, Z.; Sinno, S.; Poudrier, G.; Motosko, C.C.; Chiodo, M.; Saia, W.; Gothard, D.; Thomson, J.E.; Hazen, A. Platelet Rich Plasma for Photodamaged Skin: A Pilot Study. *J. Cosmet. Dermatol.* **2019**, *18*, 77–83. [[CrossRef](#)]
116. Charles-de-Sá, L.; Gontijo-de-Amorim, N.; Sbarbati, A.; Benati, D.; Bernardi, P.; Borojevic, R.; Carias, R.B.V.; Rigotti, G. Photoaging Skin Therapy with PRP and ADSC: A Comparative Study. *Stem Cells Int.* **2020**, *2020*, 1–13. [[CrossRef](#)]
117. Alam, M.; Hughart, R.; Champlain, A.; Geisler, A.; Paghdal, K.; Whiting, D.; Hammel, J.A.; Maisel, A.; Rapcan, M.J.; West, D.P.; et al. Effect of Platelet-Rich Plasma Injection for Rejuvenation of Photoaged Facial Skin. *JAMA Dermatol.* **2018**, *154*, 1447. [[CrossRef](#)]
118. Surowiak, P.; Tsepkołenko, V.; Olszański, R. Platelet-Rich Plasma Stimulates Collagen Type I Synthesis in the Human Skin: A Placebo-Controlled in Vivo Study. *Acta Dermatovenerol. Croat.* **2023**, *31*, 171–177.
119. Corriero, A.; Giglio, M.; Inchingolo, F.; Moschetta, A.; Varrassi, G.; Puntillo, F. Gut Microbiota Modulation and Its Implications on Neuropathic Pain: A Comprehensive Literature Review. *Pain Ther.* **2024**, *13*, 33–51. [[CrossRef](#)]
120. da Silva, L.Q.; Cancela, R.B.B.; de Lima Montalvão, S.A.; Huber, S.C.; Vieira-Damiani, G.; Triglia, R.M.; Annichino-Bizzacchi, J.M. The Effect of Lyophilized Platelet Rich-Plasma on Skin Aging: A Non-Randomized, Controlled, Pilot Trial. *Arch. Dermatol. Res.* **2021**, *313*, 863–871. [[CrossRef](#)]
121. Hui, Q.; Chang, P.; Guo, B.; Zhang, Y.; Tao, K. The Clinical Efficacy of Autologous Platelet-Rich Plasma Combined with Ultra-Pulsed Fractional CO₂ Laser Therapy for Facial Rejuvenation. *Rejuvenation Res.* **2017**, *20*, 25–31. [[CrossRef](#)] [[PubMed](#)]
122. Li, Y.; Wang, X.; Li, Y.; Li, D.; Li, S.; Shen, C. Efficacy and Safety of Allogeneic Platelet-Rich Plasma in Chronic Wound Treatment: A Meta-Analysis of Randomized Controlled Trials. *Sci. Rep.* **2024**, *14*, 25209. [[CrossRef](#)] [[PubMed](#)]
123. Emer, J. Platelet-Rich Plasma (PRP): Current Applications in Dermatology. *Skin Ther. Lett.* **2019**, *24*, 1–6.
124. Kassir, M.; Kroumpouzos, G.; Puja, P.; Katsambas, A.; Galadari, H.; Lotti, T.; Abdelmaksoud, A.; Grabbe, S.; Juchems, E.; Goldust, M. Update in Minimally Invasive Periorbital Rejuvenation with a Focus on Platelet-Rich Plasma: A Narrative Review. *J. Cosmet. Dermatol.* **2020**, *19*, 1057–1062. [[CrossRef](#)]
125. Motosko, C.C.; Khouri, K.S.; Poudrier, G.; Sinno, S.; Hazen, A. Evaluating Platelet-Rich Therapy for Facial Aesthetics and Alopecia: A Critical Review of the Literature. *Plast. Reconstr. Surg.* **2018**, *141*, 1115–1123. [[CrossRef](#)]
126. Elghblawi, E. Platelet-Rich Plasma, the Ultimate Secret for Youthful Skin Elixir and Hair Growth Triggering. *J. Cosmet. Dermatol.* **2018**, *17*, 423–430. [[CrossRef](#)]

127. Chamata, E.S.; Bartlett, E.L.; Weir, D.; Rohrich, R.J. Platelet-Rich Plasma: Evolving Role in Plastic Surgery. *Plast. Reconstr. Surg.* **2021**, *147*, 219–230. [[CrossRef](#)]
128. Evans, A.G.; Ivanic, M.G.; Botros, M.A.; Pope, R.W.; Halle, B.R.; Glassman, G.E.; Genova, R.; Al Kassis, S. Rejuvenating the Periorbital Area Using Platelet-Rich Plasma: A Systematic Review and Meta-Analysis. *Arch. Dermatol. Res.* **2021**, *313*, 711–727. [[CrossRef](#)]
129. Phoebe, L.K.W.; Lee, K.W.A.; Chan, L.K.W.; Hung, L.C.; Wu, R.; Wong, S.; Wan, J.; Yi, K.-H. Use of Platelet Rich Plasma for Skin Rejuvenation. *Skin Res. Technol.* **2024**, *30*, e13714. [[CrossRef](#)]
130. White, C.; Brahs, A.; Dorton, D.; Witfill, K. Platelet-Rich Plasma: A Comprehensive Review of Emerging Applications in Medical and Aesthetic Dermatology. *J. Clin. Aesthetic Dermatol.* **2021**, *14*, 44–57.
131. Samadi, P.; Sheykhasan, M.; Khoshinani, H.M. The Use of Platelet-Rich Plasma in Aesthetic and Regenerative Medicine: A Comprehensive Review. *Aesthetic Plast. Surg.* **2019**, *43*, 803–814. [[CrossRef](#)] [[PubMed](#)]
132. Gorodilova, A.V.; Kharisova, C.B.; Osinnikova, M.N.; Kitaeva, K.V.; Filin, I.Y.; Mayasin, Y.P.; Solovyeva, V.V.; Rizvanov, A.A. The Well-Forgotten Old: Platelet-Rich Plasma in Modern Anti-Aging Therapy. *Cells* **2024**, *13*, 1755. [[CrossRef](#)] [[PubMed](#)]
133. Cianci, C.; Pappalettera, G.; Renna, G.; Casavola, C.; Laurenziello, M.; Battista, G.; Pappalettere, C.; Ciavarella, D. Mechanical Behavior of PET-G Tooth Aligners Under Cyclic Loading. *Front. Mater.* **2020**, *7*, 104. [[CrossRef](#)]
134. Compilato, D.; Cirillo, N.; Termine, N.; Kerr, A.R.; Paderni, C.; Ciavarella, D.; Campisi, G. Long-Standing Oral Ulcers: Proposal for a New “S-C-D Classification System”. *J. Oral Pathol. Med.* **2009**, *38*, 241–253. [[CrossRef](#)] [[PubMed](#)]
135. Inchincolo, F.; Tatullo, M.; Abenavoli, F.M.; Marrelli, M.; Inchincolo, A.D.; Villabruna, B.; Inchincolo, A.M.; Dipalma, G. Severe Anisocoria after Oral Surgery under General Anesthesia. *Int. J. Med. Sci.* **2010**, *7*, 314–318. [[CrossRef](#)]
136. Inchincolo, A.D.; Patano, A.; Coloccia, G.; Ceci, S.; Inchincolo, A.M.; Marinelli, G.; Malcangi, G.; Di Pede, C.; Garibaldi, M.; Ciocia, A.M.; et al. Treatment of Class III Malocclusion and Anterior Crossbite with Aligners: A Case Report. *Medicina* **2022**, *58*, 603. [[CrossRef](#)]
137. Inchincolo, A.D.; Inchincolo, A.M.; Malcangi, G.; Avantario, P.; Azzollini, D.; Buongiorno, S.; Viapiano, F.; Campanelli, M.; Ciocia, A.M.; De Leonardi, N.; et al. Effects of Resveratrol, Curcumin and Quercetin Supplementation on Bone Metabolism—A Systematic Review. *Nutrients* **2022**, *14*, 3519. [[CrossRef](#)]
138. Russo, L.L.; Ciavarella, D.; Salamini, A.; Guida, L. Alignment of Intraoral Scans and Registration of Maxillo-Mandibular Relationships for the Edentulous Maxillary Arch. *J. Prosthet. Dent.* **2019**, *121*, 737–740. [[CrossRef](#)]
139. Lin, J.; Sclafani, A.P. Platelet-Rich Plasma for Skin Rejuvenation and Tissue Fill. *Facial Plast. Surg. Clin. N. Am.* **2018**, *26*, 439–446. [[CrossRef](#)]
140. Abuaf, O.K.; Yildiz, H.; Baloglu, H.; Bilgili, M.E.; Simsek, H.A.; Dogan, B. Histologic Evidence of New Collagen Formulation Using Platelet Rich Plasma in Skin Rejuvenation: A Prospective Controlled Clinical Study. *Ann. Dermatol.* **2016**, *28*, 718–724. [[CrossRef](#)]
141. Cazzato, G.; Massaro, A.; Colagrande, A.; Lettini, T.; Cicco, S.; Parente, P.; Nacchiero, E.; Lospalluti, L.; Cascardi, E.; Giudice, G.; et al. Dermatopathology of Malignant Melanoma in the Era of Artificial Intelligence: A Single Institutional Experience. *Diagn. Basel Switz.* **2022**, *12*, 1972. [[CrossRef](#)] [[PubMed](#)]
142. Inchincolo, F.; Inchincolo, A.D.; Palumbo, I.; Trilli, I.; Guglielmo, M.; Mancini, A.; Palermo, A.; Inchincolo, A.M.; Dipalma, G. The Impact of Cesarean Section Delivery on Intestinal Microbiota: Mechanisms, Consequences, and Perspectives—A Systematic Review. *Int. J. Mol. Sci.* **2024**, *25*, 1055. [[CrossRef](#)] [[PubMed](#)]
143. Inchincolo, F.; Inchincolo, A.M.; Avantario, P.; Settanni, V.; Fatone, M.C.; Piras, F.; Di Venere, D.; Inchincolo, A.D.; Palermo, A.; Dipalma, G. The Effects of Periodontal Treatment on Rheumatoid Arthritis and of Anti-Rheumatic Drugs on Periodontitis: A Systematic Review. *Int. J. Mol. Sci.* **2023**, *24*, 17228. [[CrossRef](#)] [[PubMed](#)]
144. Lei, X.; Xu, P.; Cheng, B. Problems and Solutions for Platelet-Rich Plasma in Facial Rejuvenation: A Systematic Review. *Aesthetic Plast. Surg.* **2019**, *43*, 457–469. [[CrossRef](#)]
145. Minetti, E.; Dipalma, G.; Palermo, A.; Patano, A.; Inchincolo, A.D.; Inchincolo, A.M.; Inchincolo, F. Biomolecular Mechanisms and Case Series Study of Socket Preservation with Tooth Grafts. *J. Clin. Med.* **2023**, *12*, 5611. [[CrossRef](#)]
146. Lorusso, F.; Inchincolo, F.; Dipalma, G.; Postiglione, F.; Fulle, S.; Scarano, A. Synthetic Scaffold/Dental Pulp Stem Cell (DPSC) Tissue Engineering Constructs for Bone Defect Treatment: An Animal Studies Literature Review. *Int. J. Mol. Sci.* **2020**, *21*, 9765. [[CrossRef](#)]
147. Corriero, A.; Giglio, M.; Soloperto, R.; Inchincolo, F.; Varrassi, G.; Puntillo, F. Microbial Symphony: Exploring the Role of the Gut in Osteoarthritis-Related Pain. *A Narrative Review. Pain Ther.* **2024**, *13*, 409–433. [[CrossRef](#)]
148. Inchincolo, A.M.; Malcangi, G.; Piras, F.; Palmieri, G.; Settanni, V.; Riccaldo, L.; Morolla, R.; Buongiorno, S.; de Ruvo, E.; Inchincolo, A.D.; et al. Precision Medicine on the Effects of Microbiota on Head-Neck Diseases and Biomarkers Diagnosis. *J. Pers. Med.* **2023**, *13*, 933. [[CrossRef](#)]

149. Inchingo, F.; Inchingo, A.M.; Inchingo, A.D.; Fatone, M.C.; Ferrante, L.; Avantario, P.; Fiore, A.; Palermo, A.; Amenduni, T.; Galante, F.; et al. Bidirectional Association between Periodontitis and Thyroid Disease: A Scoping Review. *Int. J. Environ. Res. Public. Health* **2024**, *21*, 860. [[CrossRef](#)]
150. Cervelli, V.; Garcovich, S.; Bielli, A.; Cervelli, G.; Curcio, B.C.; Scioli, M.G.; Orlandi, A.; Gentile, P. The Effect of Autologous Activated Platelet Rich Plasma (AA-PRP) Injection on Pattern Hair Loss: Clinical and Histomorphometric Evaluation. *Biomed. Res. Int.* **2014**, *2014*, 60709. [[CrossRef](#)]
151. Nakamura, S.; Ishihara, M.; Takikawa, M.; Murakami, K.; Kishimoto, S.; Nakamura, S.; Yanagibayashi, S.; Kubo, S.; Yamamoto, N.; Kiyosawa, T. Platelet-Rich Plasma (PRP) Promotes Survival of Fat-Grafts in Rats. *Ann. Plast. Surg.* **2010**, *65*, 101–106. [[CrossRef](#)] [[PubMed](#)]
152. Redaelli, A.; Romano, D.; Marcianó, A. Face and Neck Revitalization with Platelet-Rich Plasma (PRP): Clinical Outcome in a Series of 23 Consecutively Treated Patients. *J. Drugs Dermatol.* **2010**, *9*, 466–472. [[PubMed](#)]
153. Stessuk, T.; Puzzi, M.B.; Chaim, E.A.; Alves, P.C.M.; de Paula, E.V.; Forte, A.; Izumizawa, J.M.; Oliveira, C.C.; Frei, F.; Ribeiro-Paes, J.T. Platelet-Rich Plasma (PRP) and Adipose-Derived Mesenchymal Stem Cells: Stimulatory Effects on Proliferation and Migration of Fibroblasts and Keratinocytes in Vitro. *Arch. Dermatol. Res.* **2016**, *308*, 511–520. [[CrossRef](#)] [[PubMed](#)]
154. Inchingo, A.M.; Gargiulo Isacco, C.; Inchingo, A.D.; Nguyen, K.C.D.; Cantore, S.; Santacroce, L.; Scacco, S.; Cirulli, N.; Corriero, A.; Puntillo, F.; et al. The Human Microbiota Key Role in the Bone Metabolism Activity. *Eur. Rev. Med. Pharmacol. Sci.* **2023**, *27*, 2659–2670. [[CrossRef](#)]
155. Peng, G.L. Platelet-Rich Plasma for Skin Rejuvenation. *Facial Plast. Surg. Clin. N. Am.* **2019**, *27*, 405–411. [[CrossRef](#)]
156. Gargiulo Isacco, C.; Balzanelli, M.G.; Garzone, S.; Lorusso, M.; Inchingo, F.; Nguyen, K.C.D.; Santacroce, L.; Mosca, A.; Del Prete, R. Alterations of Vaginal Microbiota and Chlamydia Trachomatis as Crucial Co-Causative Factors in Cervical Cancer Genesis Procured by HPV. *Microorganisms* **2023**, *11*, 662. [[CrossRef](#)]
157. Cantore, S.; Ballini, A.; De Vito, D.; Martelli, F.S.; Georgakopoulos, I.; Almasri, M.; Dibello, V.; Altini, V.; Farronato, G.; Dipalma, G.; et al. Characterization of Human Apical Papilla-Derived Stem Cells. *J. Biol. Regul. Homeost. Agents* **2017**, *31*, 901–910.
158. Huynh, T.D.; Nguyen, H.K.; Inchingo, A.M.; Bao Tran, H.L.; Dipalma, G.; Mancini, A.; Cao Diem Nguyen, K.; Balzanelli, M.G.; Distratis, P.; Lazzaro, R.; et al. Soft Tissue Regeneration in Animal Models Using Grafts from Adipose Mesenchymal Stem Cells and Peripheral Blood Fibrin Gel. *Eur. Rev. Med. Pharmacol. Sci.* **2023**, *27*, 3670–3680. [[CrossRef](#)]
159. de Almeida Malzoni, C.M.; Pichotano, E.C.; Freitas de Paula, L.G.; de Souza, R.V.; Okamoto, R.; Austin, R.S.; Marcantonio, E., Jr.; de Molon, R.S.; Zandim-Barcelos, D.L. Combination of Leukocyte and Platelet-Rich Fibrin and Demineralized Bovine Bone Graft Enhanced Bone Formation and Healing after Maxillary Sinus Augmentation: A Randomized Clinical Trial. *Clin. Oral Investig.* **2023**, *27*, 5485–5498. [[CrossRef](#)]
160. Knoll, B.; Hersant, B. Invited Discussion on: Concentrated Growth Factor (CGF): The Newest Platelet Concentrate and Its Application in Nasal Hyaluronic Acid Injection Complications. *Aesthetic Plast. Surg.* **2023**, *47*, 1794–1795. [[CrossRef](#)]
161. Huniadi, A.; Zaha, I.A.; Naghi, P.; Stefan, L.; Sachelarie, L.; Bodog, A.; Szuhai-Bimbo, E.; Macovei, C.; Sandor, M. Autologous Platelet-Rich Plasma (PRP) Efficacy on Endometrial Thickness and Infertility: A Single-Centre Experience from Romania. *Medicina* **2023**, *59*, 1532. [[CrossRef](#)] [[PubMed](#)]
162. Manole, C.G.; Soare, C.; Ceafalan, L.C.; Voiculescu, V.M. Platelet-Rich Plasma in Dermatology: New Insights on the Cellular Mechanism of Skin Repair and Regeneration. *Life* **2023**, *14*, 40. [[CrossRef](#)] [[PubMed](#)]
163. Zhang, Y.; Liang, J.; Lu, F.; Dong, Z. Survival Mechanisms and Retention Strategies in Large-Volume Fat Grafting: A Comprehensive Review and Future Perspectives. *Aesthetic Plast. Surg.* **2024**, *48*, 4178–4193. [[CrossRef](#)]
164. Lo Muzio, L.; Santarelli, A.; Panzarella, V.; Campisi, G.; Carella, M.; Ciavarella, D.; Di Cosola, M.; Giannone, N.; Bascones, A. Oral Squamous Cell Carcinoma and Biological Markers: An Update on the Molecules Mainly Involved in Oral Carcinogenesis. *Minerva Stomatol.* **2007**, *56*, 341–347.
165. Ciavarella, D.; Monsurrò, A.; Padricelli, G.; Battista, G.; Laino, L.; Perillo, L. Unilateral Posterior Crossbite in Adolescents: Surface Electromyographic Evaluation. *Eur. J. Paediatr. Dent.* **2012**, *13*, 25–28.
166. Cameli, N.; Mariano, M.; Cordone, I.; Abril, E.; Masi, S.; Foddai, M.L. Autologous Pure Platelet-Rich Plasma Dermal Injections for Facial Skin Rejuvenation: Clinical, Instrumental, and Flow Cytometry Assessment. *Dermatol. Surg.* **2017**, *43*, 826–835. [[CrossRef](#)]
167. Magalon, J.; Chateau, A.L.; Bertrand, B.; Louis, M.L.; Silvestre, A.; Giraudo, L.; Veran, J.; Sabatier, F. DEPA Classification: A Proposal for Standardising PRP Use and a Retrospective Application of Available Devices. *BMJ Open Sport Exerc. Med.* **2016**, *2*, e000060. [[CrossRef](#)]
168. Everts, P.A.M.; Hoogbergen, M.M.; Weber, T.A.; Devilee, R.J.J.; van Monftort, G.; de Hingh, I.H.J.T. Is the Use of Autologous Platelet-Rich Plasma Gels in Gynecologic, Cardiac, and General, Reconstructive Surgery Beneficial? *Curr. Pharm. Biotechnol.* **2012**, *13*, 1163–1172. [[CrossRef](#)]
169. Livden, J.K.; Bjerke, J.R.; Degre, M.; Matre, R. Effect of UV Radiation on Interferon, Immunoglobulins and Complement Components in Serum from Healthy Individuals. *Photodermatology* **1987**, *4*, 296–301.

170. Inchingo, F.; Tatullo, M.; Abenavoli, F.M.; Marrelli, M.; Inchingo, A.D.; Corelli, R.; Inchingo, A.M.; Dipalma, G. Surgical Treatment of Depressed Scar: A Simple Technique. *Int. J. Med. Sci.* **2011**, *8*, 377–379. [[CrossRef](#)]
171. Gasparro, R.; Adamo, D.; Masucci, M.; Sammartino, G.; Mignogna, M.D. Use of Injectable Platelet-Rich Fibrin in the Treatment of Plasma Cell Mucositis of the Oral Cavity Refractory to Corticosteroid Therapy: A Case Report. *Dermatol. Ther.* **2019**, *32*, e13062. [[CrossRef](#)] [[PubMed](#)]
172. Arezzo, F.; Loizzi, V.; La Forgia, D.; Moschetta, M.; Tagliafico, A.S.; Cataldo, V.; Kawosha, A.A.; Venerito, V.; Cazzato, G.; Ingravallo, G.; et al. Radiomics Analysis in Ovarian Cancer: A Narrative Review. *Appl. Sci.* **2021**, *11*, 7833. [[CrossRef](#)]
173. Díaz-Ley, B.; Cuevast, J.; Alonso-Castro, L.; Calvo, M.I.; Ríos-Buceta, L.; Orive, G.; Anitua, E.; Jaén, P. Benefits of Plasma Rich in Growth Factors (PRGF) in Skin Photodamage: Clinical Response and Histological Assessment. *Dermatol. Ther.* **2015**, *28*, 258–263. [[CrossRef](#)] [[PubMed](#)]
174. Neinaa, Y.M.E.-H.; Hodeib, A.A.E.-H.; Morquos, M.M.; Elgarhy, L.H. Platelet-Poor Plasma Gel vs Platelet-Rich Plasma for Infraorbital Rejuvenation: A Clinical and Dermoscopic Comparative Study. *Dermatol. Ther.* **2020**, *33*, e14255. [[CrossRef](#)]
175. Inchingo, A.M.; Patano, A.; Di Pede, C.; Inchingo, A.D.; Palmieri, G.; de Ruvo, E.; Campanelli, M.; Buongiorno, S.; Carpentiere, V.; Piras, F.; et al. Autologous Tooth Graft: Innovative Biomaterial for Bone Regeneration. *Tooth Transformer®* and the Role of Microbiota in Regenerative Dentistry. A Systematic Review. *J. Funct. Biomater.* **2023**, *14*, 132. [[CrossRef](#)]
176. Yang, H.S.; Shin, J.; Bhang, S.H.; Shin, J.-Y.; Park, J.; Im, G.-I.; Kim, C.-S.; Kim, B.-S. Enhanced Skin Wound Healing by a Sustained Release of Growth Factors Contained in Platelet-Rich Plasma. *Exp. Mol. Med.* **2011**, *43*, 622. [[CrossRef](#)]
177. Man, D.; Plosker, H.; Winland-Brown, J.E. The Use of Autologous Platelet-Rich Plasma (Platelet Gel) and Autologous Platelet-Poor Plasma (Fibrin Glue) in Cosmetic Surgery. *Plast. Reconstr. Surg.* **2001**, *107*, 229–236. [[CrossRef](#)]
178. Kim, D.-S.; Park, S.-H.; Park, K.-C. Transforming Growth Factor-B1 Decreases Melanin Synthesis via Delayed Extracellular Signal-Regulated Kinase Activation. *Int. J. Biochem. Cell Biol.* **2004**, *36*, 1482–1491. [[CrossRef](#)]
179. Har-Shai, Y.; Bodner, S.R.; Egozy-Golan, D.; Lindenbaum, E.S.; Ben-Izhak, O.; Mitz, V.; Hirshowitz, B. Viscoelastic Properties of the Superficial Musculoaponeurotic System (SMAS): A Microscopic and Mechanical Study. *Aesthetic Plast. Surg.* **1997**, *21*, 219–224. [[CrossRef](#)]
180. Cho, J.-W.; Kim, S.-A.; Lee, K.-S. Platelet-Rich Plasma Induces Increased Expression of G1 Cell Cycle Regulators, Type I Collagen, and Matrix Metalloproteinase-1 in Human Skin Fibroblasts. *Int. J. Mol. Med.* **2012**, *29*, 32–36. [[CrossRef](#)]
181. Anitua, E.; Sanchez, M.; De la Fuente, M.; Zalduendo, M.M.; Orive, G. Plasma Rich in Growth Factors (PRGF-Endoret) Stimulates Tendon and Synovial Fibroblasts Migration and Improves the Biological Properties of Hyaluronic Acid. *Knee Surg. Sports Traumatol. Arthrosc.* **2012**, *20*, 1657–1665. [[CrossRef](#)] [[PubMed](#)]
182. Bioulac, B.; Heppt, W.; Heppt, M. Transfer of Autologous Fat and Plasma: The Future of Anti-Aging Medicine? *HNO* **2015**, *63*, 497–503. [[CrossRef](#)] [[PubMed](#)]
183. Zeitter, S.; Sikora, Z.; Jahn, S.; Stahl, F.; Strauß, S.; Lazaridis, A.; Reimers, K.; Vogt, P.M.; Aust, M.C. Microneedling: Matching the Results of Medical Needling and Repetitive Treatments to Maximize Potential for Skin Regeneration. *Burns* **2014**, *40*, 966–973. [[CrossRef](#)] [[PubMed](#)]
184. Nusgens, B.V.; Humbert, P.; Rougier, A.; Colige, A.C.; Haftek, M.; Lambert, C.A.; Richard, A.; Creidi, P.; Lapière, C.M. Topically Applied Vitamin C Enhances the mRNA Level of Collagens I and III, Their Processing Enzymes and Tissue Inhibitor of Matrix Metalloproteinase 1 in the Human Dermis. *J. Investigig. Dermatol.* **2001**, *116*, 853–859. [[CrossRef](#)] [[PubMed](#)]
185. Inchingo, F.; Inchingo, A.M.; Latini, G.; Ferrante, L.; de Ruvo, E.; Campanelli, M.; Longo, M.; Palermo, A.; Inchingo, A.D.; Dipalma, G. Difference in the Intestinal Microbiota between Breastfeed Infants and Infants Fed with Artificial Milk: A Systematic Review. *Pathogens* **2024**, *13*, 533. [[CrossRef](#)]
186. Dutta, S.R.; Singh, P.; Passi, D.; Patter, P. Mandibular Third Molar Extraction Wound Healing with and Without Platelet Rich Plasma: A Comparative Prospective Study. *J. Maxillofac. Oral Surg.* **2015**, *14*, 808–815. [[CrossRef](#)]
187. Inchingo, F.; Tatullo, M.; Abenavoli, F.M.; Marrelli, M.; Inchingo, A.D.; Inchingo, A.M.; Dipalma, G. Comparison between Traditional Surgery, CO₂ and Nd:Yag Laser Treatment for Generalized Gingival Hyperplasia in Sturge-Weber Syndrome: A Retrospective Study. *J. Investigig. Clin. Dent.* **2010**, *1*, 85–89. [[CrossRef](#)]
188. Shin, M.-K.; Lee, J.-H.; Lee, S.-J.; Kim, N.-I. Platelet-Rich Plasma Combined with Fractional Laser Therapy for Skin Rejuvenation. *Dermatol. Surg.* **2012**, *38*, 623–630. [[CrossRef](#)]
189. Grassi, F.R.; Ciccolella, F.; D'Apolito, G.; Papa, F.; Iuso, A.; Salzo, A.E.; Trentadue, R.; Nardi, G.M.; Scivetti, M.; De Matteo, M.; et al. Effect of Low-Level Laser Irradiation on Osteoblast Proliferation and Bone Formation. *J. Biol. Regul. Homeost. Agents* **2011**, *25*, 603–614.
190. Mandriani, B.; Pellè, E.; Mannavola, F.; Palazzo, A.; Marsano, R.M.; Ingravallo, G.; Cazzato, G.; Ramello, M.C.; Porta, C.; Strosberg, J.; et al. Development of Anti-Somatostatin Receptors CAR T Cells for Treatment of Neuroendocrine Tumors. *J. Immunother. Cancer* **2022**, *10*, e004854. [[CrossRef](#)]
191. Shamsaldeen, O.; Peterson, J.D.; Goldman, M.P. The Adverse Events of Deep Fractional CO₂: A Retrospective Study of 490 Treatments in 374 Patients. *Lasers Surg. Med.* **2011**, *43*, 453–456. [[CrossRef](#)] [[PubMed](#)]

192. Kohl, E.; Meierhöfer, J.; Koller, M.; Zeman, F.; Groesser, L.; Karrer, S.; Hohenleutner, U.; Landthaler, M.; Hohenleutner, S. Fractional Carbon Dioxide Laser Resurfacing of Rhytides and Photoaged Skin—A Prospective Clinical Study on Patient Expectation and Satisfaction. *Lasers Surg. Med.* **2015**, *47*, 111–119. [CrossRef] [PubMed]
193. Longo, C.; Galimberti, M.; De Pace, B.; Pellacani, G.; Bencini, P.L. Laser Skin Rejuvenation: Epidermal Changes and Collagen Remodeling Evaluated by in Vivo Confocal Microscopy. *Lasers Med. Sci.* **2013**, *28*, 769–776. [CrossRef]
194. Martínez, C.E.; Smith, P.C.; Palma Alvarado, V.A. The Influence of Platelet-Derived Products on Angiogenesis and Tissue Repair: A Concise Update. *Front. Physiol.* **2015**, *6*, 290. [CrossRef]
195. Anitua, E.; Sánchez, M.; Nurden, A.T.; Zalduendo, M.M.; de la Fuente, M.; Azofra, J.; Andía, I. Platelet-Released Growth Factors Enhance the Secretion of Hyaluronic Acid and Induce Hepatocyte Growth Factor Production by Synovial Fibroblasts from Arthritic Patients. *Rheumatology* **2007**, *46*, 1769–1772. [CrossRef]
196. Weng, T.; Wu, P.; Zhang, W.; Zheng, Y.; Li, Q.; Jin, R.; Chen, H.; You, C.; Guo, S.; Han, C.; et al. Regeneration of Skin Appendages and Nerves: Current Status and Further Challenges. *J. Transl. Med.* **2020**, *18*, 53. [CrossRef]
197. Caggiano, M.; Gasparro, R.; D'Ambrosio, F.; Pisano, M.; Di Palo, M.P.; Contaldo, M. Smoking Cessation on Periodontal and Peri-Implant Health Status: A Systematic Review. *Dent. J.* **2022**, *10*, 162. [CrossRef]
198. Gong, H.; Li, K.; Xie, R.; Du, G.; Li, L.; Wang, S.; Yin, J.; Gu, J.; Wang, P.; Chen, M.; et al. Clinical Therapy of Platelet-Rich Plasma vs Hyaluronic Acid Injections in Patients with Knee Osteoarthritis. *Medicine* **2021**, *100*, e25168. [CrossRef]
199. Murali, A.; Khan, I.; Tiwari, S. Navigating the Treatment Landscape: Choosing between Platelet-Rich Plasma (PRP) and Hyaluronic Acid (HA) for Knee Osteoarthritis Management—A Narrative Review. *J. Orthop. Rep.* **2024**, *3*, 100248. [CrossRef]
200. Mai, Y.; Zhang, J.; Huang, G.; He, J.; Liu, X.; Guo, L.; Wei, Z.; Jiang, L. Efficacy of the Combination Therapy of Platelet-Rich Plasma and Hyaluronic Acid on Improving Knee Pain and Dysfunction in Patients with Moderate-to-Severe KOA: A Protocol for a Randomised Controlled Trial. *BMJ Open* **2023**, *13*, e068743. [CrossRef]
201. Xu, Y.; Wu, X.; Zhang, Y.; Yu, Y.; Gan, J.; Tan, Q. Engineered Artificial Skins: Current Construction Strategies and Applications. *Eng. Regen.* **2023**, *4*, 438–450. [CrossRef]
202. Wang, M.; Hong, Y.; Fu, X.; Sun, X. Advances and Applications of Biomimetic Biomaterials for Endogenous Skin Regeneration. *Bioact. Mater.* **2024**, *39*, 492–520. [CrossRef] [PubMed]
203. Li, Y.-Y.; Ji, S.-F.; Fu, X.-B.; Jiang, Y.-F.; Sun, X.-Y. Biomaterial-Based Mechanical Regulation Facilitates Scarless Wound Healing with Functional Skin Appendage Regeneration. *Mil. Med. Res.* **2024**, *11*, 13. [CrossRef] [PubMed]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.