



REVIEW ARTICLE

Decoding the impact of gut microbiota on heart failure

Shuhong Zhao^{a,b,1}, Lingxuan Dan^{a,b,1}, Rong Huang^{a,b},
Zhuoyu Shen^{a,b}, Dan Huang^{a,b}, Pan Wu^{c,d,**}, Zhenguo Ma^{a,b,*}

^a Department of Cardiology, Renmin Hospital of Wuhan University, Wuhan, Hubei 430060, China

^b Hubei Key Laboratory of Metabolic and Chronic Diseases, Wuhan, Hubei 430060, China

^c Department of Adult Internal Medicine, Maternal and Child Health Hospital of Hubei Province, Wuhan, Hubei 430070, China

^d Women and Children's Hospital of Hubei Province, Wuhan, Hubei 430070, China

Received 9 July 2024; received in revised form 24 December 2024; accepted 21 January 2025

Available online 6 March 2025

KEYWORDS

Gut microbiota;
Gut-heart axis;
Heart failure;
Substance
metabolism;
Therapeutic
modulation

Abstract Decreased cardiac output in heart failure leads to intestinal ischemia and increased permeability. Substantial changes occur in the gut microbiota, characterized by a decline in beneficial bacteria and an overgrowth of potentially harmful bacteria. The gut microbiota is intricately linked to prevalent risk factors for heart failure, including hypertension, diabetes, obesity, and renal insufficiency. Furthermore, imbalanced microbiota-derived metabolites enter the bloodstream and may contribute to the progression of heart failure. Ongoing research explores gut microbiota manipulation to alleviate heart failure with probiotics, targeted antibiotics, fecal microbiota transplantation, and dietary adjustments. This review summarizes how gut microbiota participates in heart failure and highlights the emerging promise of modulating gut dysbiosis as a therapeutic approach for managing heart failure.

© 2025 The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co., Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

* Corresponding author. Department of Cardiology, Renmin Hospital of Wuhan University, Wuhan, Hubei 430060, China.

** Corresponding author. Department of Adult Internal Medicine, Maternal and Child Health Hospital of Hubei Province, Wuhan, Hubei 430070, China.

E-mail addresses: pan_wu_vip@163.com (P. Wu), zhengma@whu.edu.cn (Z. Ma).

Peer review under the responsibility of the Genes & Diseases Editorial Office, in alliance with the Association of Chinese Americans in Cancer Research (ACACR, Baltimore, MD, USA).

¹ These authors contributed equally to this work.

Introduction

Cardiovascular diseases (CVDs) consistently rank as the leading cause of global mortality, significantly impacting both health and quality of life. However, the incidence of CVDs is currently increasing year by year.¹ Heart failure (HF) occurs in the end stage of various CVDs, such as hypertension, myocardial infarction, and myocarditis. It is characterized by cardiac remodeling, which involves various structural and functional changes in the myocardium that develop in response to chronic stress or injury to the heart.² These changes help maintain heart function to some extent. However, in the long term, they tend to accelerate the progression of CVDs and ultimately lead to HF.³ Therefore, cardiac remodeling has remained a focal point of our attention and investigation.^{4–11} Despite substantial advancements in the medication for HF, mortality rates from CVDs continue to be disturbingly high. Therefore, developing targeted therapies is of great importance for the prevention and treatment of CVDs.

Recently, the role of the gut microbiota in HF has received extensive attention. In HF patients, substantial changes occur in the gut microbiota, characterized by a decline in beneficial bacteria and an overgrowth of potentially harmful bacteria.¹² These changes indicate that gut dysbiosis plays an important role in the development of HF, and therapy targeting the gut microbiota may become a new treatment approach. Furthermore, increasing evidence suggests that microbiota-derived metabolites, such as trimethylamine N-oxide (TMAO), bile acids (BAs), short-chain fatty acids (SCFAs), and amino acids (AAs), may influence the development of myocardial remodeling.^{13–18} Modulating the composition of the gut microbiota appears to help ameliorate myocardial fibrosis and delay the development of HF.

In the past few years, there has been an explosion of reports regarding gut microbiota, and many excellent review articles have summarized the interactions between

the gut microbiota and various organs in the body.^{19–24} Here, we focus on the role of gut microbiota in HF and its significance in cardiac remodeling. Although emerging evidence suggests that gut dysbiosis significantly influences the progression of HF, the specific mechanisms remain unclear. Given that HF is a multifactorial chronic condition influenced by numerous risk factors, it is critical to understand how these factors interact with gut dysbiosis in the context of HF. This review creatively explores how gut microbiota-derived metabolites influence HF and associated risk factors via molecular and cellular pathways. Furthermore, we discuss the potential advantages and challenges of novel therapeutic approaches targeting the gut microbiota, aiming to bridge the knowledge gap between gut health and HF, thereby laying a foundation for future research and clinical advancements.

Composition and function of the gut microbiota

Composition of the gut microbiota

The gut microbiota is the collection of microorganisms that reside in the digestive system.²⁵ The human gut microbiota comprises 10^{13} to 10^{14} microorganisms, and their collective genome, known as the “microbiome”, contains at least 100 times more genes than the human genome.²⁶ The intestinal tract harbors several major bacterial groups classified into the *phyla* *Bacteroidetes*, *Firmicutes*, *Actinobacteria*, and *Proteobacteria*.²⁷

Function of the gut microbiota

Currently, extensive research has elucidated the roles of specific gut microbiota species in a broad array of physiological activities (Table 1).

Table 1 The function of the main gut microbiota in the body.

	Host site	Functions	Reference
Gut microbiota			
<i>Lactobacillus</i>	Gastrointestinal tract, oral cavity, vagina, skin, breast	Increases barrier function Alleviates colitis Upregulates hepatic antioxidant enzymes and ruminal barrier function Ameliorates colorectal tumorigenesis Improves bone loss Improves insulin resistance	28 29 30 31 32 33
<i>Bifidobacterium</i>	Colons of newborns and infants, large intestines of adults, oral cavity	Improves cognitive function and mood Relieves gut dysmotility-related disorders Prevents colorectal cancer Enhances host antiviral response	34 35 36 37
<i>Faecalibacterium prausnitzii</i>	Terminal ileum, ileocecal region	Alleviates colitis Suppresses colonic inflammation Improves the epithelial barrier integrity	38 39 40,41
<i>Clostridium butyricum</i>	Colon	Alleviates intestinal inflammation Protects intestinal barrier function Promotes bone development	42,43 44,45 46
<i>Akkermansia muciniphila</i>	The mucous layer of the colon	Alleviates intestinal inflammation Promotes epithelial development	47 48

In a nutshell, the gut microbiota has two main functions: participation in substance metabolism and regulation of intestinal barrier function. Gut microbiota can degrade carbohydrates, monosaccharides, fats, proteins, and peptides. Specific bacteria possess the capability to degrade cellulose, converting it into SCFAs such as acetic, propionic, and butyric acids. Notably, these compounds serve as the primary energy source for colonic epithelial cells. Furthermore, SCFAs play a critical role in regulating immune responses and modulating inflammation.^{49,50} In addition, the gut microbiota impacts the synthesis, metabolism, and transportation of fat-soluble vitamins at various levels. Moreover, the synthesis of BAs and AAs is closely associated with the gut microbiota.^{51,52}

Gut microbiota closely interacts with intestinal epithelial cells to reduce the infiltration of harmful substances by producing adhesion proteins and polysaccharides, which strengthen the integrity of the mucosal barrier. Additionally, by competing for nutrients and attachment sites, gut microbiota reduces the colonization of pathogenic bacteria.^{53,54} It is generally accepted that regulating gut microbiota can improve intestinal barrier function. For example, the probiotic strain of *Lactobacillus plantarum* HN082 (*Lp082*) enhanced the chemical barrier through decreased levels of intercellular cell adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1). In addition, *Lp082* improved the mechanical barrier by increasing the levels of zonula occludens-1 and zonula occludens-2 while decreasing the levels of claudin-1 and claudin-2.⁵⁵ Certain strains of enteric bacteria can also degrade and eliminate harmful substances, such as toxins and pathogenic factors.

Relationship between gut microbiota and host health

Based on the above functions, strong associations exist between gut microbiota and human health. Currently, gut microbiota is considered to have bidirectional communication with multiple organs of the body, thus affecting the progression of a variety of diseases (Fig. 1).

The gut dysbiosis can lead to a variety of diseases. Conversely, treatments targeting gut microbiota can reverse these conditions. For example, gut dysbiosis plays a pivotal role as a direct instigator of inflammatory bowel disease.⁵⁶ From this point of view, fecal microbiota transplantation (FMT) has become a common treatment for inflammatory bowel disease, benefiting many patients.⁵⁷ Similarly, the disruption of the intestinal barrier is observed in patients with liver cirrhosis, and this disruption forms a vicious cycle that exacerbates disease progression. Many studies have confirmed the role of regulating microbial structure in the treatment of liver cirrhosis. Currently, FMT is gradually being translated into clinical practice to improve the outcome of liver cirrhosis.^{58–60} Moreover, gut microbiota has been implicated in the pathogenesis of various autoimmune diseases, including rheumatoid arthritis, systemic lupus erythematosus, and type 1 diabetes mellitus, as well as neuropsychiatric disorders like depression and Alzheimer's disease. Furthermore, a lot of substances from the intestine, such as inflammatory mediators, hormones, and metabolites, can reach the brain and thereby influence brain function.⁶¹ Intriguingly, gut microbiota can also modulate the synthesis and metabolism of neurotransmitters, including γ -

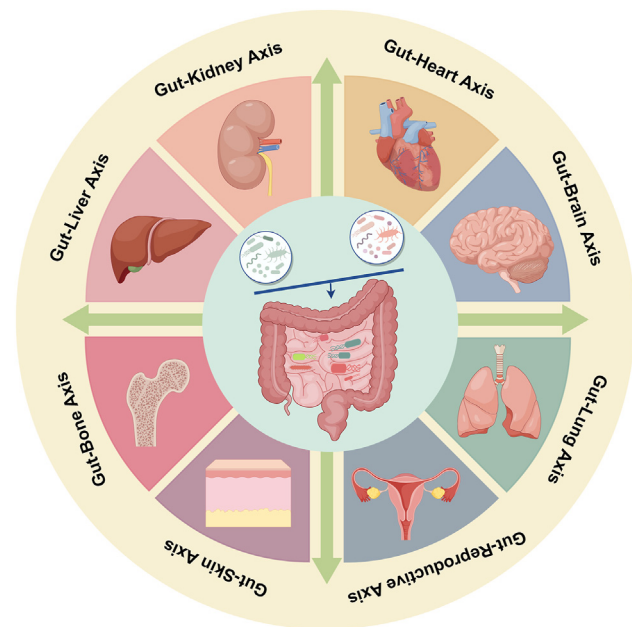


Figure 1 A bidirectional communication system exists between the gut and multiple organs throughout the body. The gut microbiota and their metabolic products engage in interactions with other host organs, creating what is known as cross-talk, which constitutes the host-microbial immune-metabolic axes. Currently, eight axes have been extensively researched and acknowledged: gut-heart axis, gut-brain axis, gut-lung axis, gut-liver axis, gut-kidney axis, gut-skin axis, gut-bone axis, and gut-reproductive axis.

aminobutyric acid, dopamine, and serotonin. These neurotransmitters play crucial regulatory roles in the brain, being intimately associated with emotional states, memory, and cognitive processes.^{62,63}

Last but not least, the connection between the gut microbiota and the heart is equally intimate. HF is associated with chronic low-grade inflammation, a condition that can be exacerbated or reversed by changes in the gut microbiota.^{33,64} Consistently, clinical evidence suggests that the use of certain probiotics can reduce inflammation in HF patients.⁶⁵ Moreover, gut dysbiosis shares common risk factors with HF. Multiple animal studies have demonstrated that modulating gut microbiota can improve cardiac function and survival rates in HF models.^{33,66,67}

Indeed, given its vast impact, the gut microbiota is often referred to as the second genome of the human body. Gut dysbiosis affects the overall state of the body beyond simple intestinal diseases. It can influence various physiological and pathological processes, including immune function, metabolic regulation, and even neurological function. Thus, investigating the multifaceted roles of the gut microbiota in both health and disease is of paramount importance.

The impact of gut microbiota on HF

Gut microbiota and risk factors of CVDs

With the increasing prevalence of poor lifestyle choices, there is a corresponding rise in the prevalence of individual

risk factors for HF, including hypertension, diabetes, and obesity. Gut microbiota can influence metabolism, immune system, and inflammatory response, all of which can contribute to the development of HF and high-risk factors for CVDs (Table 2). Furthermore, several additional factors are associated with CVDs, which may stem from or exacerbate gut dysbiosis. These factors include, but are not limited to, smoking, dietary patterns, environmental pollutants, and renal insufficiency.^{68–71} Each of these elements independently and cumulatively contributes to CVDs, highlighting the complexity of the interplay between gut microbiota and cardiovascular health.

Gut microbiota and HF

HF is the ultimate common process caused by various initial heart injuries. It is triggered by an imbalance in compensatory mechanisms and pathogenic processes. Numerous clinical and basic studies have consistently demonstrated the critical role that gut microbiota plays in myocardial fibrosis and HF. Research has shown that transverse aortic constriction surgery administered to germ-free mice often results in more severe cardiac hypertrophy and cardiac dysfunction compared with the control group. Conversely, supplementation of acetate and propionate can improve heart function, reduce fibrosis, and reverse extracellular matrix dysfunction.⁸⁷ In the transverse aortic constriction-induced cardiac remodeling model, 16S rRNA sequencing of fecal samples revealed a reduction in bacteria-producing tryptophan and SCFAs in wild-type mice.¹⁴ Mice treated with antibiotics demonstrated a notable dose-dependent increase in mortality compared with the control when subjected to myocardial infarction. High-performance liquid chromatography analysis showed a decrease in SCFAs after antibiotic treatment. Following fecal reconstruction and nutritional SCFA supplementation, the mice's survival rate and physiological state were significantly improved.¹⁸ Facts have shown that both reactive fibrosis and reparative fibrosis are affected by gut microbiota extensively. It was discovered that the gut microbiota of HF patients was primarily distinguished by a decline in both diversity and composition. In particular, there was an increase in *Proteobacteria* and *Candida* and a decrease in the number of taxa in the *Firmicutes*.^{88,89} *Lactobacillus* is one of the most extensively studied genera in the gut microbiota. Research has shown that *Lactiplantibacillus plantarum* P470 has the potential to improve the gut microbiota in the feces of patients with coronary heart disease.⁹⁰ On the contrary, *Proteobacteria* include many potential pathogens, such as *Escherichia coli* and *Salmonella*. An increase in pathogenic bacteria is associated with heightened inflammation and infection risk in HF patients.¹⁶

The gut microbiota acts directly on the failing heart

The structure and function of the gastrointestinal tract in patients with HF are altered.^{91,92} When HF occurs, the decrease in cardiac output and the activation of the neurohumoral system stimulate the redistribution of systemic blood, leading to insufficient perfusion of the gastrointestinal tract.⁹³ Structurally, intestinal wall thickening and

intestinal edema are often observed. Functionally, intestinal permeability increases and intestinal barrier function is impaired.⁹² Accompanied by the increase in intestinal inflammation and permeability, gut pathogenic microbiota, and their toxins can translocate from the intestinal tract to the bloodstream, causing systemic chronic inflammation and contributing to chronic HF.^{89,94} Elevated levels of inflammatory factors are frequently associated with worse prognosis and more severe clinical symptoms in HF patients. Evidence suggests that these inflammatory mediators are linked to fibrosis, apoptosis, and hypertrophy of cardiomyocytes.⁹⁵ Additionally, a significant increase in bacterial concentration and adhesion in the intestinal mucosal biofilm has been observed in patients with HF.^{91,93} Mamic et al conducted a nationwide study of hospitalized patients and found that those with HF had higher rates of sepsis, pneumonia, and urinary tract infections, indirectly confirming the high inflammatory state of patients with HF.⁹⁶ A higher infection rate of *Clostridium difficile* is observed in hospitalized HF patients. Therefore, the disorder of gut microbiota can directly affect the body, leading to the progression and deterioration of HF.

The gut microbiota acts indirectly on the failing heart through its metabolites

The metabolic substrates for gut microbiota primarily originate from food that the host cannot efficiently digest, as well as mucus secreted by intestinal epithelial cells. Following the activity of gut microbiota, a multitude of metabolites are produced, including both detrimental and beneficial compounds such as lipopolysaccharides, peptidoglycans, trimethylamine, BAs, and SCFAs.¹² Numerous studies have demonstrated that the gut microbiota-derived metabolites can significantly influence systemic health. (Fig. 2).

TMAO

TMAO is a metabolite derived from dietary choline and has been shown to promote the development of atherosclerosis through various mechanisms. On one hand, TMAO can stimulate inflammation and endothelial dysfunction by activating the reactive oxygen species/thioredoxin-interacting protein (TXNIP)/NOD-like receptor family pyrin domain-containing 3 (NLRP3) inflammasome pathway.⁹⁷ On the other hand, TMAO exhibits pro-thrombotic potential *in vivo*.⁹⁸ Supplementation with dietary choline and TMAO accelerates atherosclerotic plaque formation in mice.⁹⁹ In a prospective study conducted by Tang et al, fasting TMAO levels were significantly elevated in patients with HF compared with those without HF, thereby suggesting an increased risk of long-term mortality.¹⁰⁰ These findings have been corroborated in animal research as well. In a mouse model of doxorubicin-induced cardiac fibrosis, a TMAO-enriched diet exacerbated cardiac dysfunction and myocardial fibrosis. TMAO promoted collagen deposition, fibrosis, and inflammatory factor expression in a dose-dependent manner through the activation of the transforming growth factor beta1 (TGF- β 1)/Smad3 signaling pathway.¹⁰¹ 3,3-Dimethyl-1-butanol has been shown to

Table 2 The role of gut microbiota in the risk factors of CVDs.

Risk factors	Model	Observations	Interventions and effects	Reference
Hypertension	Chronic Ang II infusion	Decreased microbial richness Increased ratio of <i>Firmicutes</i> to <i>Bacteroides</i>	Minocycline: lowers blood pressure and restores the ecological balance of gut microbiota	72
	Angiotensin II infusion + apolipoprotein E-deficient	Myocardial hypertrophy, myocardial fibrosis, and vascular dysfunction	Propionate: attenuates hypertension	73
	Spontaneously hypertension	Increased ratio of <i>Firmicutes</i> to <i>Bacteroides</i> Lower acetate- and higher lactate-producing bacteria	Losartan: alleviates intestinal ecological imbalance and intestinal sympathetic drive	74
	High-salt diet	Increased intestinal inflammation Increased <i>Firmicutes</i> , <i>Proteobacteria</i> , and <i>Prevotella</i>	Fecal microbiota transplantation from high-salt diet mouse to germ-free mouse: make intestinal inflammation and hypertension prone to occur	75
	Angiotensin II infusion	Vascular dysfunction and hypertension	Germ-free mice: relieves cardiac inflammation, fibrosis and systolic dysfunction	76
Diabetes	HFD + streptozotocin-induced dilated cardiomyopathy	Increased <i>Clostridiales</i> and <i>Lachnospiraceae</i> increased intestinal permeability	Pyridostigmine: improves the intestinal barrier, increases ATP production, and attenuates cardiac dysfunction, hypertrophy, and fibrosis	77
	HFD-induced obesity and diabetes	Decreased abundance of <i>Blautia</i> Increased abundance of <i>Faecalibacterium</i> , <i>Butyricicoccus</i> , and <i>Megasphaera</i>	<i>Blautia wexlerae</i> : alleviates T2DM via altering metabolism and exhibiting anti-inflammatory effects	78
	HFD-induced obesity	Increased concentrations of secondary BAs and SCFAs	Vitamin K2: improves impaired glycemic homeostasis and insulin sensitivity in T2DM through the gut microbiota and fecal metabolites	79
	Streptozotocin-induced T2DM	Lower community richness and evenness	Papeiflorin: ameliorates cardiac dysfunction and myocardial injury via altering the structure and composition of the gut microbiota community	80
	HFD-induced obesity and diabetes	Increased <i>Bacteroides fragilis</i> and decreased GUDCA Glucose intolerance and insulin resistance	Metformin: improves metabolic dysfunction part through a <i>B. fragilis</i> –GUDCA–intestinal FXR axis	81
	db/db mice	High blood glucose levels and insulin resistance Gut microbiota perturbations and disruption of gut barrier function	Cholinergic drugs: mitigates diabetic cardiac injury by augmenting the quantity of anti-inflammatory bacteria while diminishing the quantity of pro-inflammatory bacteria through the LPS-1-mediated ERK/Egr-1 pathway	82
	Obesity	HF + intermittent hypoxia-induced Obstructive sleep apnea	<i>Lactobacillus rhamnosus</i> GG: improves cardiac dysfunction and reduces cardiac remodeling and inflammation in obstructive sleep apnea mice via the activation of the Nrf2 pathway	83

(continued on next page)

Table 2 (continued)

Risk factors	Model	Observations	Interventions and effects	Reference
	HFD-induced obesity	Decreased capacity to metabolize ethanolamine	Probiotic: restores the activity of ethanolamine- β -lactamase and reduces intestinal permeability and inflammation	84
	HFD-induced obesity	Glucose intolerance, insulin resistance, and hyperinsulinemia Impaired gut barrier function	Spermidine: decreases metabolic endotoxemia and improves intestinal barrier integrity through autophagy pathway and TLR4-mediated microbial signal transduction	85
	HFD-induced obesity	Decreased fermentation of SCFAs and transformation of BAs Increased metabolism of BCAAs	Co-housing Ob and Ln mice: prevents weight gain and obesity-related metabolic phenotypes in obese mice	86

Note: HFD, high-fat diet; GUDCA, glycochenodeoxycholic acid; BCAAs, branched-chain amino acids; BAs, bile acids; SCFAs, short-chain fatty acids; T2DM, type 2 diabetes mellitus.

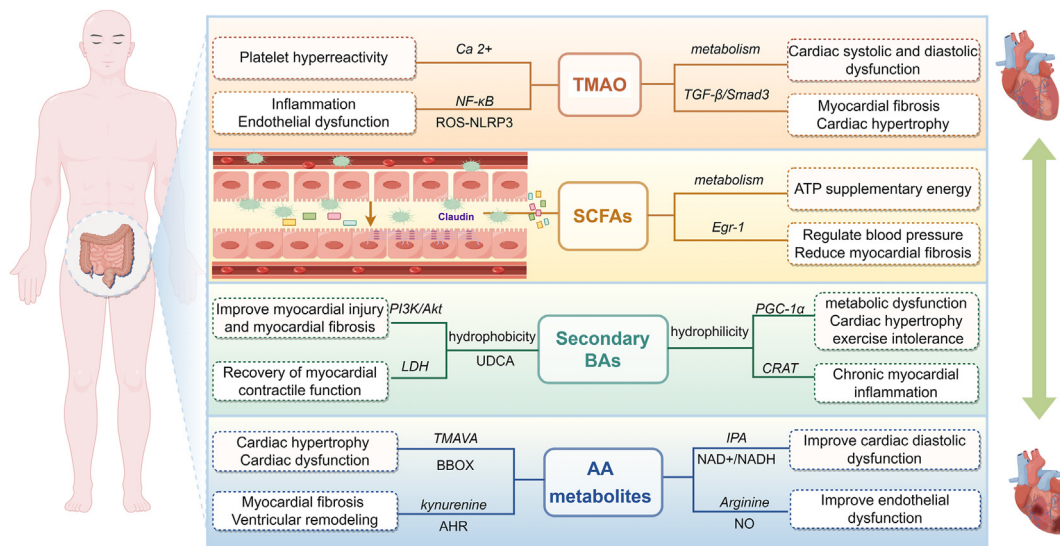


Figure 2 Mechanisms of dietary metabolites generated by the gut microbiota in heart failure. Through the processes of intestinal motility and the metabolic activities of the gut microbiota, substances such as TMAO, SCFAs, BAs, and AAs are synthesized in the intestine. In patients with heart failure, an imbalance in the metabolism of these substances serves as a critical indirect driver that either triggers or worsens the condition. TMAO, trimethylamine N-oxide; BAs, bile acids; SCFAs, short-chain fatty acids; AAs, amino acids.

mitigate TMAO levels in HF mice by inhibiting the p65 nuclear factor kappa B (NF- κ B) signaling pathway and the TGF- β 1/Smad3 signaling cascade, thereby ameliorating the adverse cardiac remodeling driven by TMAO.¹⁰² The research by Organ et al directly substantiated the detrimental impact of TMAO on the heart. In comparison with a standard diet, a TMAO-enriched diet was found to notably exacerbate myocardial hypertrophy, myocardial fibrosis, and the severity of HF in mice following transverse aortic constriction.¹⁰³ Similarly, neonatal rat cardiomyocytes exposed to TMAO *in vitro* exhibited increased cell size and up-regulated the expression of hypertrophic markers such

as atrial natriuretic peptide (ANP) and β -myosin heavy chain (β -MHC).¹⁰⁴ The increased concentration of TMAO in plasma and heart tissue can directly inhibit pyruvate and fatty acid oxidation. The disorder in cardiac energy metabolism leads to both systolic and diastolic dysfunction in cardiomyocytes.¹⁰⁵ Moreover, TMAO has been shown to stimulate the elevation of inflammatory cytokines and the accumulation of reactive oxygen species within the body, leading to an inflammatory response and direct injury to cardiomyocytes.¹⁰⁶ Consequently, TMAO has been substantiated as a promising cardiovascular risk biomarker and holds potential as a therapeutic target in HF management.

SCFAs

SCFAs are produced through the anaerobic fermentation of dietary fiber by colonic bacteria or yeast.¹⁰⁷ The most prevalent SCFAs in the body include acetic acid, propionic acid, and butyric acid.¹⁰⁸ The primary bacterial genera responsible for producing SCFAs include *Bacteroides*, *Bifidobacterium*, *Firmicutes*, *Ruminococcus*, *Lactobacillus*, *Clostridium*, and *Streptococcus*.¹⁰⁷ Currently, many studies have demonstrated that SCFAs can enhance intestinal barrier integrity, modulate the immune system, regulate inflammatory responses, improve glucose and lipid metabolism, and influence blood pressure.^{109–112} As previously discussed, the levels of SCFAs are significantly reduced in patients with HF compared with healthy controls, while exogenous supplementation has been shown to mitigate the progression of HF. Specifically, butyrate potentiates the integrity of intestinal epithelial tight junctions by up-regulating tight junction components and stimulating protein recombination. It preserves intestinal barrier function and reduces the ingress of detrimental substances, such as endotoxins, into the systemic circulation. In essence, SCFAs can reduce the risk of systemic inflammation.¹¹³ Moreover, transcriptome analysis showed that dietary supplements high in fiber and acetate reduced the proportion of *Bacteroides* and *Prevotella*, accompanied by a down-regulation of early growth response 1 (Egr-1) in the heart and kidney, which significantly reduced systolic and diastolic blood pressure, cardiac fibrosis, and left ventricular hypertrophy.¹¹⁴ SCFAs also compensate for the impaired oxidation capacity of long-chain fatty acids in the hypertrophic heart, serving as an alternative substrate for ATP production through oxidation.¹¹⁵ Therefore, it can be identified that SCFAs hold considerable significance in the prevention and management of HF.

BAs

BAs are recognized as emulsifiers of fats and fat-soluble vitamins. Primary BAs are synthesized from cholesterol in the liver, while secondary BAs are produced by bacteria residing in the intestinal tract.

Several studies have demonstrated that the total concentration and composition ratio of BAs significantly altered in patients with HF. A prospective clinical investigation found that plasma BA levels in HF patients were notably lower than those in healthy controls. This decline was accompanied by an increase in the ratio of secondary to primary BAs. Through univariate analysis, it has been demonstrated that the altered pattern of BAs correlates with a reduced overall survival rate in patients with HF.¹¹⁶ In a BA overload model, pronounced cardiac hypertrophy and exercise intolerance were observed. Further research has revealed that elevated serum BA levels can suppress the expression of peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 α), thereby inhibiting fatty acid oxidation both *in vivo* and *in vitro*, which ultimately leads to biliary and cardiac metabolic dysfunction.¹¹⁷ In dysfunctional cardiomyocytes, the depletion of carnitine acetyltransferase promotes

cholesterol utilization for BA synthesis. The intracellular accumulation of BAs or their intermediate metabolites results in the release of mitochondrial DNA into the cytosol, which subsequently initiates a type I interferon (IFN- λ) response and activates absent in melanoma 2 (AIM2) inflammasomes. This cascade ultimately contributes to the progression of chronic myocardial inflammation and HF.¹¹⁸

BAs can be classified into hydrophobic BAs and hydrophilic BAs based on their distinct structures. The former category is known for inducing cytotoxicity and apoptosis, and the latter exhibits a reduced cytotoxic effect. Ursodeoxycholic acid stands out as the most hydrophilic of the cholic acids. Initially used in the treatment of chronic liver diseases, ursodeoxycholic acid can also decrease lactate dehydrogenase release and enhance the recovery of cardiac systolic function in the ischemia/reperfusion model.¹¹⁹ Moreover, ursodeoxycholic acid ameliorates myocardial infarction and inhibits mitochondrial permeability transition pore opening in a phosphoinositide 3-kinase (PI3K)/protein kinase B (PKB/Akt)-dependent pathway.¹²⁰ Ursodeoxycholic acid can also exert inhibitory effects on apoptosis and reduce myocardial hypertrophy and fibrosis.¹²¹ Therefore, BAs significantly influence the progression and management of HF, positioning them as a potential therapeutic target for intervention.

AAs

Alterations in plasma AAs and their derivatives are detectable in patients with HF and have been deemed as early indicators of CVDs.¹²² Zhao et al identified a strong association between increased levels of N-trimethyl-5-aminopentanoic acid and the risks of heart-related death and transplantation. This acid can interfere with cardiac energy metabolism via the γ -butyrobetaine hydroxylase (BBOX) pathway. It inhibits fatty acid oxidation and leads to the accumulation of lipids, thus exacerbating cardiac hypertrophy and dysfunction.¹²³ Likewise, through evidence supported by single-nucleus RNA sequencing, it has been established that certain molecules can activate aromatic hydrocarbon receptors and contribute to ventricular remodeling.¹²⁴ During the progression of HF, impairments in AA metabolism can result in inadequate energy supply to the myocardium, exacerbating damage to cardiomyocytes. However, the role of AAs and their metabolites in HF has yielded varying outcomes across different studies. In the pressure overload model, replacing dietary AAs has been shown to improve systemic glucose metabolism and rebalance metabolic substrate utilization in HF by enhancing mitochondrial fuel oxidation efficiency.¹²⁵ Furthermore, Wang et al have demonstrated that FMT can restore cardiac levels of nicotinamide and sirtuin 3 (SIRT3). This restoration subsequently reduces metabolic remodeling and inflammation, thereby mediating a protective effect against diastolic dysfunction in the heart.¹²⁶

The diverse roles played by AAs and their metabolites in CVDs can be attributed to the body's energy intake status. In instances of surplus energy, excessive consumption of AAs might elevate the risk of HF, whereas in conditions characterized by energy insufficiency, dietary supplementation with AAs could promote health.

Immune regulation induced by gut microbiota

Gut microbiota can produce a variety of immunomodulatory factors that directly or indirectly regulate the activity of immune cells, such as dendritic cells and CD4⁺ T cells.^{127,128} Gut microbiota can modulate the inflammatory response of the immune system by activating signaling pathways such as Toll-like receptors (TLRs), NF- κ B, and mitogen-activated protein kinases (MAPKs).¹²⁷ SCFAs also have important effects on immune cell activity and function. The effects of butyrate on immune cells are mainly mediated in three ways: direct activation of surface G-protein-coupled receptors (GPCRs), activation of peroxisome proliferator-activated receptor gamma (PPAR γ), and inhibition of butyrate-sensitive histone deacetylase (HDAC) isoforms.

An imbalance in the gut microbiota can lead to excessive growth of pathogenic microorganisms, resulting in the release of large amounts of endotoxins and inflammatory agents. This fosters chronic low-grade systemic inflammation, which increases susceptibility to various illnesses, including CVDs, autoimmune disorders, and certain metabolic conditions, such as atherosclerosis. Concurrently, a reduction in beneficial microbiota weakens resistance against infections, increasing the likelihood of pathogen invasion. For example, rheumatic heart disease, which is caused by an autoimmune response to group A streptococcal infection, has been shown to be closely related to changes in gut microbiota.¹²⁹ Experiments in myocardial infarction models showed improvements after FMT, monocyte transfer, or SCFA dietary supplementation and highlighted the relevance of immune modulation in treating CVDs.¹⁸

Novel therapeutic interventions in HF

Probiotic bacteria

The decline in the number and relative abundance of beneficial gut microbiota in HF patients can exacerbate systemic inflammation and accelerate the progression of HF. Probiotics can stimulate the proliferation of specific microbial populations and enhance the balance of the gut microbiota. Components intrinsic to probiotics, such as peptidoglycan and lipoteichoic acid, can serve as antigens and exert both direct and indirect immune-stimulatory effects, thereby enhancing overall immunity.¹³⁰ Previous investigations have shown that administering *Lactobacillus* and probiotics before myocardial infarction in mice shifted the balance of SCFAs towards propionate and conferred cardioprotective benefits.¹⁸ Another study conducted by Gan et al also observed a protective effect of *Lactobacillus rhamnosus* on the heart following myocardial infarction.¹³¹ In a randomized controlled clinical trial, consuming probiotic yogurt over 10 weeks effectively improved the inflammatory status in patients with chronic HF.^{132,133} Furthermore, probiotics can facilitate nutrient digestion and absorption, indirectly influencing cardiovascular metabolism. Notably, despite probiotics generally having an excellent safety profile, a lack of regulation may raise the risk of unintended translocation of probiotics into the bloodstream, potentially leading to sepsis. Therefore,

careful consideration and appropriate regulation are essential when administering probiotics.

Diet

CVDs are intrinsically linked to metabolism. Hence, the importance of adopting healthy dietary habits is quite evident. Both in humans and mice, high salt intake has been associated with an increase in certain bacteria, such as *Streptococcus*, *Proteus*, and *Prevotella*, which predisposes mice to vascular inflammation and hypertension.⁷⁵ The dietary approach to stop hypertension (DASH) diet, recommended by the American Heart Association guidelines, emphasizes a rich intake of fruits, vegetables, whole grains, and lean products while limiting the consumption of sugary foods, beverages, red meat, and saturated fats.¹³⁴ Currently, accumulating evidence supports the benefits of the DASH diet in HF patients. A high-fiber diet has notably been shown to decrease systolic and diastolic blood pressure, cardiac fibrosis, and left ventricular hypertrophy, partly through the down-regulation of Egr-1 in the heart and kidneys.¹¹⁴ Diets enriched with fermented foods consistently enhance microbiota diversity and diminish inflammatory biomarkers.¹³⁵ Moreover, the CORDIOPREV study suggests that a Mediterranean diet intervention may effectively reduce the risk of myocardial infarction, revascularization, and ischemic stroke, outperforming a low-fat diet approach.¹³⁶ Beyond monitoring food choices, paying attention to eating patterns is equally significant. Research indicates that intermittent fasting practices, such as alternate-day fasting or time-restricted feeding, may assist in lowering the risk of CVDs.¹³⁷

FMT

As a novel therapeutic approach, FMT has garnered substantial interest in the realm of HF research in recent years. FMT involves the transfer of a donor's fecal microbial community to a patient, aiming to restore or reconstruct the balance of gut microbiota and thereby enhance the overall health status of the recipient. When applied to HF model organisms, introducing healthy microbiota has been shown to improve cardiac function, attenuate myocardial fibrosis, suppress inflammation, and facilitate the restoration of metabolic homeostasis. FMT has gained widespread acceptance and utilization in patients suffering from recurrent *Clostridioides difficile* infection.¹³⁸ Nevertheless, the application of FMT in HF remains at an exploratory stage. Further clinical studies are essential to rigorously validate its safety and efficacy specifically for HF patients. With the ever-deepening comprehension of the intricate relationship between the gut microbiota and CVDs, FMT holds promise as a potential new strategy for the management and treatment of HF.

Rational use of antibiotics

Infection, often a principal trigger for acute exacerbations of HF, ranks among the primary reasons for hospitalizations and fatalities in HF patients. Hence, numerous studies have concentrated on the application of antibiotics to manage

infections instigated by intestinal pathogens. Doxycycline, for instance, has been shown to mitigate left ventricular remodeling in patients with myocardial infarction.¹³⁹ However, antibiotics pose a dualistic challenge to the gut microbiota. Improper usage of antibiotics, particularly misuse or deviation from recommended indications, doses, and treatment durations, can engender gut microbiota imbalance and deplete the body's normal symbiotic microbial community. Experimental models demonstrated that antibiotic-treated mice exhibited a severe, dose-dependent increase in mortality post-myocardial infarction.¹⁸ Therefore, the judicious use of antibiotics transcends merely preventing the emergence of drug resistance. It is also crucial for preserving the balance of the gut microbiota, thereby maintaining gut health and overall well-being. Indeed, nonspecific antibiotic interventions might inflict more harm than benefit.

Conclusion and prospect

The results of clinical and basic studies have revealed that the changes in gut microbiota significantly influence the progression of HF. There is a decrease in beneficial species such as *Lactobacilli* and *Bifidobacteria*, concurrent with an increase in potentially harmful bacteria like *Clostridia* and *Bacteroides*. A compromised intestinal barrier enables bacterial translocation and abnormal production of metabolites, amplifying inflammation and promoting cardiac remodeling. We have discussed the effects of gut dysbiosis on the host's health, particularly the occurrence of HF. However, given that the gut microbiota plays a role in the metabolism of various substances, the relationship between different types of microbiota-derived metabolites and HF remains to be further elucidated. Additionally, further research should focus on identifying specific molecules and their pathways.

In addition to descriptive investigations, we have also attempted to reveal the cellular and molecular mechanisms of gut microbiota causing HF. As a result, in terms of therapeutic strategy development, HF can be alleviated by regulating gut dysbiosis, and new therapeutic drugs can be developed based on microbiota-derived metabolites. Probiotics, prebiotics, and FMT have demonstrated potential in both experimental studies and clinical scenarios to enhance the prognosis of HF. We have summarized the therapeutic effects and accessibility of the above interventions. However, many issues still need to be further addressed in current clinical treatment strategies. On one hand, longitudinal studies are needed to track the long-term effects of microbiota changes on cardiovascular health due to the gut's dynamism. On the other hand, more extensive and rigorous randomized controlled trials are required to validate their therapeutic benefits and long-term safety. Moreover, the gut microbiota plays an active role in drug metabolism, affecting HF patients' response to standard treatments. Therefore, studying the interaction between the microbiota and drug metabolism is the key to optimizing regimens, enhancing drug effectiveness, and reducing side effects.

In summary, the relationship between gut microbiota and HF opens a promising research field, shedding new light on disease etiology and progression and potentially

introducing novel therapeutic approaches. To incorporate gut microbiota regulation into HF management, ongoing basic research and prospective clinical trials are essential to deepening our foundational knowledge.

Outstanding questions

To thoroughly examine the pivotal role of gut microbiota in CVDs and effectively translate these findings into therapies, several key areas require focused research: i) Determine causality: Is gut microbiota disruption a direct cause or a secondary effect in HF? Longitudinal studies and mechanistic experiments are essential to establish causality. ii) Identify metabolites: Are there unknown critical metabolites produced by gut microbiota, and what are their precise impacts on cardiovascular health? iii) Assess therapeutic potential: Can manipulating gut microbiota serve as an effective adjunct therapy for managing HF? iv) Develop personalized interventions: How can individualized gut microbiota intervention strategies be designed for HF patients with different disease etiologies and stages of progression?

CRedit authorship contribution statement

Shuhong Zhao: Writing – original draft, Visualization, Conceptualization. **Lingxuan Dan:** Writing – original draft, Conceptualization. **Rong Huang:** Writing – review & editing. **Zhuoyu Shen:** Writing – review & editing. **Dan Huang:** Writing – review & editing. **Pan Wu:** Writing – review & editing, Conceptualization. **Zhenguo Ma:** Writing – review & editing, Supervision, Conceptualization.

Conflict of interests

The authors declared no competing interests.

Funding

This work is supported in part by grants from the National Natural Science Foundation of China (No. 82070410, 82270248), the Young Top-notch Talent Cultivation Program of Hubei Province (China), the Knowledge Innovation Program of Wuhan-Basic Research (China), and the Fundamental Research Funds for the Central Universities (China) (No. 2042021kf0205). The funders did not have any role in the paper design, data collection, data analysis, interpretation, and writing of the paper.

Acknowledgements

All figures in this article are drawn by Figdraw (No. ITAWTcfc9f, AOPWU7bbb6, UOUAW4c4db).

References

1. Wang Z, Ma L, Liu M, Fan J, Hu S. Writing committee of the report on cardiovascular health and diseases in China.

- Summary of the 2022 report on cardiovascular health and diseases in China. *Chin Med J (Engl)*. 2023;136(24):2899–2908.
2. Hashimoto H, Olson EN, Bassel-Duby R. Therapeutic approaches for cardiac regeneration and repair. *Nat Rev Cardiol*. 2018;15(10):585–600.
 3. Ma ZG, Yuan YP, Wu HM, Zhang X, Tang QZ. Cardiac fibrosis: new insights into the pathogenesis. *Int J Biol Sci*. 2018;14(12):1645–1657.
 4. Ma ZG, Yuan YP, Zhang X, et al. C1q-tumour necrosis factor-related protein-3 exacerbates cardiac hypertrophy in mice. *Cardiovasc Res*. 2019;115(6):1067–1077.
 5. Che Y, Liu YT, Wang ZP, et al. Cardiac tumour necrosis factor receptor-associated factor 7 mediates the ubiquitination of apoptosis signal-regulating kinase 1 and aggravates cardiac hypertrophy. *Cardiovasc Res*. 2024;120(16):2031–2046.
 6. Wu Q, Yao Q, Hu T, et al. Dapagliflozin protects against chronic heart failure in mice by inhibiting macrophage-mediated inflammation, independent of SGLT2. *Cell Rep Med*. 2023;4(12):101334.
 7. Zhang X, Hu C, Kong CY, et al. FNDC5 alleviates oxidative stress and cardiomyocyte apoptosis in doxorubicin-induced cardiotoxicity via activating AKT. *Cell Death Differ*. 2020;27(2):540–555.
 8. Ma ZG, Yuan YP, Fan D, et al. IIRX2 regulates angiotensin II-induced cardiac fibrosis by transcriptionally activating EGR1 in male mice. *Nat Commun*. 2023;14(1):4967.
 9. Hu C, Zhang X, Song P, et al. Meteorin-like protein attenuates doxorubicin-induced cardiotoxicity via activating cAMP/PKA/SIRT1 pathway. *Redox Biol*. 2020;37:101747.
 10. Ma ZG, Kong CY, Wu HM, et al. Toll-like receptor 5 deficiency diminishes doxorubicin-induced acute cardiotoxicity in mice. *Theranostics*. 2020;10(24):11013–11025.
 11. Xie SY, Liu SQ, Zhang T, et al. USP28 serves as a key suppressor of mitochondrial morphofunctional defects and cardiac dysfunction in the diabetic heart. *Circulation*. 2024;149(9):684–706.
 12. Wilson Tang WH, Li DY, Hazen SL. Dietary metabolism, the gut microbiome, and heart failure. *Nat Rev Cardiol*. 2019;16:137–154.
 13. Li X, Li R, You N, Zhao X, Li J, Jiang W. Butyric acid ameliorates myocardial fibrosis by regulating M1/M2 polarization of macrophages and promoting recovery of mitochondrial function. *Front Nutr*. 2022;9:875473.
 14. Carrillo-Salinas FJ, Anastasiou M, Ngwenyama N, et al. Gut dysbiosis induced by cardiac pressure overload enhances adverse cardiac remodeling in a T cell-dependent manner. *Gut Microbes*. 2020;12(1):1–20.
 15. Yang W, Zhang S, Zhu J, et al. Gut microbe-derived metabolite trimethylamine N-oxide accelerates fibroblast-myofibroblast differentiation and induces cardiac fibrosis. *J Mol Cell Cardiol*. 2019;134:119–130.
 16. Zhao J, Zhang Q, Cheng W, et al. Heart–gut microbiota communication determines the severity of cardiac injury after myocardial ischaemia/reperfusion. *Cardiovasc Res*. 2023;119(6):1390–1402.
 17. Palm CL, Nijholt KT, Bakker BM, Daan Westenbrink B. Short-chain fatty acids in the metabolism of heart failure - Rethinking the fat stigma. *Front Cardiovasc Med*. 2022;9:915102.
 18. Tang TWH, Chen HC, Chen CY, et al. Loss of gut microbiota alters immune system composition and cripples postinfarction cardiac repair. *Circulation*. 2019;139(5):647–659.
 19. Budden KF, Gellatly SL, Wood DLA, et al. Emerging pathogenic links between microbiota and the gut-lung axis. *Nat Rev Microbiol*. 2017;15(1):55–63.
 20. Morais LH, Schreiber 4th HL, Mazmanian SK. The gut microbiota-brain axis in behaviour and brain disorders. *Nat Rev Microbiol*. 2021;19(4):241–255.
 21. Witkowski M, Weeks TL, Hazen SL. Gut microbiota and cardiovascular disease. *Circ Res*. 2020;127(4):553–570.
 22. Yang T, Richards EM, Pepine CJ, Raizada MK. The gut microbiota and the brain-gut-kidney axis in hypertension and chronic kidney disease. *Nat Rev Nephrol*. 2018;14(7):442–456.
 23. Zhang YW, Song PR, Wang SC, Liu H, Shi ZM, Su JC. Diets intervene osteoporosis via gut-bone axis. *Gut Microbes*. 2024;16(1):2295432.
 24. Huang F, Cao Y, Liang J, et al. The influence of the gut microbiome on ovarian aging. *Gut Microbes*. 2024;16(1):2295394.
 25. Cani PD. Human gut microbiome: hopes, threats and promises. *Gut*. 2018;67(9):1716–1725.
 26. Gill SR, Pop M, Deboy RT, et al. Metagenomic analysis of the human distal gut microbiome. *Science*. 2006;312(5778):1355–1359.
 27. Arumugam M, Raes J, Pelletier E, et al. Enterotypes of the human gut microbiome. *Nature*. 2011;473(7346):174–180.
 28. Zheng J, Ali Ahmad A, Yang Y, et al. *Lactobacillus rhamnosus* CY12 enhances intestinal barrier function by regulating tight junction protein expression, oxidative stress, and inflammation response in lipopolysaccharide-induced caco-2 cells. *Int J Mol Sci*. 2022;23(19):11162.
 29. Jia DJC, Wang QW, Hu YY, et al. *Lactobacillus johnsonii* alleviates colitis by TLR1/2-STAT3 mediated CD206⁺ macrophages^{IL-10} activation. *Gut Microbes*. 2022;14(1):2145843.
 30. Izuddin WI, Humam AM, Loh TC, Foo HL, Samsudin AA. Dietary postbiotic *Lactobacillus plantarum* improves serum and ruminal antioxidant activity and upregulates hepatic antioxidant enzymes and ruminal barrier function in post-weaning lambs. *Antioxidants*. 2020;9(3):250.
 31. Zhang Q, Zhao Q, Li T, et al. *Lactobacillus plantarum*-derived indole-3-lactic acid ameliorates colorectal tumorigenesis via epigenetic regulation of CD8⁺ T cell immunity. *Cell Metab*. 2023;35(6):943–960.e9.
 32. Yuan Y, Yang J, Zhuge A, Li L, Ni S. Gut microbiota modulates osteoclast glutathione synthesis and mitochondrial biogenesis in mice subjected to ovariectomy. *Cell Prolif*. 2022;55(3):e13194.
 33. Kang Y, Kang X, Yang H, et al. *Lactobacillus acidophilus* ameliorates obesity in mice through modulation of gut microbiota dysbiosis and intestinal permeability. *Pharmacol Res*. 2022;175:106020.
 34. Kim CS, Cha L, Sim M, et al. Probiotic supplementation improves cognitive function and mood with changes in gut microbiota in community-dwelling older adults: a randomized, double-blind, placebo-controlled, multicenter trial. *J Gerontol A Biol Sci Med Sci*. 2021;76(1):32–40.
 35. Cheng J, Laitila A, Ouwehand AC. *Bifidobacterium animalis* subsp. *lactis* HN019 effects on gut health: a review. *Front Nutr*. 2021;8:790561.
 36. Chen H, Tong T, Lu SY, et al. Urea cycle activation triggered by host-microbiota maladaptation driving colorectal tumorigenesis. *Cell Metab*. 2023;35(4):651–666.e7.
 37. Niu J, Cui M, Yang X, et al. Microbiota-derived acetate enhances host antiviral response via NLRP3. *Nat Commun*. 2023;14(1):642.
 38. Wang Y, Tao H, Huang H, et al. The dietary supplement *Rhodiola crenulata* extract alleviates dextran sulfate sodium-induced colitis in mice through anti-inflammation, mediating gut barrier integrity and reshaping the gut microbiome. *Food Funct*. 2021;12(7):3142–3158.
 39. Fritsch J, Garces L, Quintero MA, et al. Low-fat, high-fiber diet reduces markers of inflammation and dysbiosis and improves quality of life in patients with ulcerative colitis. *Clin Gastroenterol Hepatol*. 2021;19(6):1189–1199.e30.
 40. Mohebbi N, Ekat K, Kreikemeyer B, Breitrück A. Barrier protection and recovery effects of gut commensal bacteria on

- differentiated intestinal epithelial cells *in vitro*. *Nutrients*. 2020;12(8):2251.
41. Xu J, Liang R, Zhang W, et al. *Faecalibacterium prausnitzii*-derived microbial anti-inflammatory molecule regulates intestinal integrity in diabetes mellitus mice via modulating tight junction protein expression. *J Diabetes*. 2020;12(3):224–236.
 42. Ma L, Shen Q, Lyu W, et al. *Clostridium butyricum* and its derived extracellular vesicles modulate gut homeostasis and ameliorate acute experimental colitis. *Microbiol Spectr*. 2022;10(4):e0136822.
 43. Zhou M, Yuan W, Yang B, Pei W, Ma J, Feng Q. *Clostridium butyricum* inhibits the progression of colorectal cancer and alleviates intestinal inflammation via the myeloid differentiation factor 88 (MyD88)-nuclear factor-kappa B (NF- κ B) signaling pathway. *Ann Transl Med*. 2022;10(8):478.
 44. Liu M, Xie W, Wan X, Deng T. *Clostridium butyricum* protects intestinal barrier function via upregulation of tight junction proteins and activation of the Akt/mTOR signaling pathway in a mouse model of dextran sodium sulfate-induced colitis. *Exp Ther Med*. 2020;20(5):10.
 45. Li W, Xu B, Wang L, et al. Effects of *Clostridium butyricum* on growth performance, gut microbiota and intestinal barrier function of broilers. *Front Microbiol*. 2021;12:777456.
 46. Song M, Zhang X, Hao G, Lin H, Sun S. *Clostridium butyricum* can promote bone development by regulating lymphocyte function in layer pullets. *Int J Mol Sci*. 2023;24(2):1457.
 47. Zheng M, Han R, Yuan Y, et al. The role of *Akkermansia muciniphila* in inflammatory bowel disease: current knowledge and perspectives. *Front Immunol*. 2023;13:1089600.
 48. Kim S, Shin YC, Kim TY, et al. Mucin degrader *Akkermansia muciniphila* accelerates intestinal stem cell-mediated epithelial development. *Gut Microbes*. 2021;13(1):1–20.
 49. Salvi PS, Cowles RA. Butyrate and the intestinal epithelium: modulation of proliferation and inflammation in homeostasis and disease. *Cells*. 2021;10(7):1775.
 50. Singh V, Lee G, Son H, et al. Butyrate producers, “the sentinel of gut”: their intestinal significance with and beyond butyrate, and prospective use as microbial therapeutics. *Front Microbiol*. 2023;13:1103836.
 51. Fogelson KA, Dorrestein PC, Zarrinpar A, Knight R. The gut microbial bile acid modulation and its relevance to digestive health and diseases. *Gastroenterology*. 2023;164(7):1069–1085.
 52. Mayneris-Perxachs J, Castells-Nobau A, Arnoriaga-Rodríguez M, et al. Microbiota alterations in proline metabolism impact depression. *Cell Metab*. 2022;34(5):681–701.e10.
 53. Wang X, Zhang P, Zhang X. Probiotics regulate gut microbiota: an effective method to improve immunity. *Molecules*. 2021;26(19):6076.
 54. Seekatz AM, Safdar N, Khanna S. The role of the gut microbiome in colonization resistance and recurrent *Clostridioides difficile* infection. *Therap Adv Gastroenterol*. 2022;15:17562848221134396.
 55. Wu Y, Jha R, Li A, et al. Probiotics (*Lactobacillus plantarum* HNU082) supplementation relieves ulcerative colitis by affecting intestinal barrier functions, immunity-related gene expression, gut microbiota, and metabolic pathways in mice. *Microbiol Spectr*. 2022;10(6):e0165122.
 56. Haneishi Y, Furuya Y, Hasegawa M, Picarelli A, Rossi M, Miyamoto J. Inflammatory bowel diseases and gut microbiota. *Int J Mol Sci*. 2023;24(4):3817.
 57. Lopetuso LR, Deleu S, Godny L, et al. The first international Rome consensus conference on gut microbiota and faecal microbiota transplantation in inflammatory bowel disease. *Gut*. 2023;72(9):1642–1650.
 58. Li M, Li K, Tang S, et al. Restoration of the gut microbiota is associated with a decreased risk of hepatic encephalopathy after TIPS. *JHEP Rep*. 2022;4(5):100448.
 59. Cheng YW, Alhaffar D, Saha S, et al. Fecal microbiota transplantation is safe and effective in patients with *Clostridioides difficile* infection and cirrhosis. *Clin Gastroenterol Hepatol*. 2021;19(8):1627–1634.
 60. Bajaj JS, Shamsaddini A, Acharya C, et al. Multiple bacterial virulence factors focused on adherence and biofilm formation associate with outcomes in cirrhosis. *Gut Microbes*. 2021;13(1):1993584.
 61. Logsdon AF, Erickson MA, Rhea EM, Salameh TS, Banks WA. Gut reactions: how the blood-brain barrier connects the microbiome and the brain. *Exp Biol Med (Maywood)*. 2018;243(2):159–165.
 62. Quillin SJ, Tran P, Prindle A. Potential roles for gamma-aminobutyric acid signaling in bacterial communities. *Bioelectricity*. 2021;3(2):120–125.
 63. Paiva IHR, Maciel LM, Silva RSD, Mendonça IP, Souza JRB, Peixoto CA. Prebiotics modulate the microbiota-gut-brain axis and ameliorate anxiety and depression-like behavior in HFD-fed mice. *Food Res Int*. 2024;182:114153.
 64. Wang P, Ma Y, Wang D, et al. Protective effects of dietary resveratrol against chronic low-grade inflammation mediated through the gut microbiota in high-fat diet mice. *Nutrients*. 2022;14(10):1994.
 65. Awoyemi A, Mayerhofer C, Felix AS, et al. Rifaximin or *Saccharomyces boulardii* in heart failure with reduced ejection fraction: results from the randomized GutHeart trial. *EBioMedicine*. 2021;70:103511.
 66. Zheng H, Zhang X, Li C, et al. BCAA mediated microbiota-liver-heart crosstalk regulates diabetic cardiomyopathy via FGF21. *Microbiome*. 2024;12(1):157.
 67. Petrick HL, Ogilvie LM, Brunetta HS, et al. Dietary nitrate and corresponding gut microbiota prevent cardiac dysfunction in obese mice. *Diabetes*. 2023;72(7):844–856.
 68. Jardon KM, Canfora EE, Goossens GH, Blaak EE. Dietary macronutrients and the gut microbiome: a precision nutrition approach to improve cardiometabolic health. *Gut*. 2022;71(6):1214–1226.
 69. Fan J, Zhou Y, Meng R, et al. Cross-talks between gut microbiota and tobacco smoking: a two-sample Mendelian randomization study. *BMC Med*. 2023;21(1):163.
 70. Liu CX, Liu YB, Peng Y, Peng J, Ma QL. Causal effect of air pollution on the risk of cardiovascular and metabolic diseases and potential mediation by gut microbiota. *Sci Total Environ*. 2024;912:169418.
 71. Wang X, Yang S, Li S, et al. Aberrant gut microbiota alters host metabolome and impacts renal failure in humans and rodents. *Gut*. 2020;69(12):2131–2142.
 72. Yang T, Santisteban MM, Rodriguez V, et al. Gut dysbiosis is linked to hypertension. *Hypertension*. 2015;65(6):1331–1340.
 73. Bartolomeaus H, Balogh A, Yakoub M, et al. Short-chain fatty acid propionate protects from hypertensive cardiovascular damage. *Circulation*. 2019;139(11):1407–1421.
 74. Robles-Vera I, Toral M, de la Visitación N, et al. Changes to the gut microbiota induced by losartan contributes to its antihypertensive effects. *Br J Pharmacol*. 2020;177(9):2006–2023.
 75. Ferguson JF, Aden LA, Barbaro NR, et al. High dietary salt-induced dendritic cell activation underlies microbial dysbiosis-associated hypertension. *JCI Insight*. 2019;5(13):e126241.
 76. Karbach SH, Schönfelder T, Brandão I, et al. Gut microbiota promote angiotensin II-induced arterial hypertension and vascular dysfunction. *J Am Heart Assoc*. 2016;5(9):e003698.
 77. Yang Y, Zhao M, He X, Wu Q, Li DL, Zang WJ. Pyridostigmine protects against diabetic cardiomyopathy by regulating vagal activity, gut microbiota, and branched-chain amino acid catabolism in diabetic mice. *Front Pharmacol*. 2021;12:647481.
 78. Hosomi K, Saito M, Park J, et al. Oral administration of *Blautia wexlerae* ameliorates obesity and type 2 diabetes via

- metabolic remodeling of the gut microbiota. *Nat Commun.* 2022;13(1):4477.
79. Zhang Y, Liu L, Wei C, et al. Vitamin K2 supplementation improves impaired glycemic homeostasis and insulin sensitivity for type 2 diabetes through gut microbiome and fecal metabolites. *BMC Med.* 2023;21(1):174.
 80. Wu H, Zhang P, Zhou J, et al. Paeoniflorin confers ferroptosis resistance by regulating the gut microbiota and its metabolites in diabetic cardiomyopathy. *Am J Physiol Cell Physiol.* 2024;326(3):C724–C741.
 81. Sun L, Xie C, Wang G, et al. Gut microbiota and intestinal FXR mediate the clinical benefits of metformin. *Nat Med.* 2018;24(12):1919–1929.
 82. Wu Q, Zhao M, Li D, He X, Zang W. Cholinergic drugs reduce metabolic inflammation and diabetic myocardial injury by regulating the gut bacterial component lipopolysaccharide-induced ERK/Egr-1 pathway. *FASEB J.* 2023;37(5):e22917.
 83. Xu H, Wang J, Cai J, et al. Protective effect of *Lactobacillus rhamnosus* GG and its supernatant against myocardial dysfunction in obese mice exposed to intermittent hypoxia is associated with the activation of Nrf2 pathway. *Int J Biol Sci.* 2019;15(11):2471–2483.
 84. Mishra SP, Wang B, Jain S, et al. A mechanism by which gut microbiota elevates permeability and inflammation in obese-diabetic mice and human gut. *Gut.* 2023;72(10):1848–1865.
 85. Ma L, Ni Y, Wang Z, et al. Spermidine improves gut barrier integrity and gut microbiota function in diet-induced obese mice. *Gut Microbes.* 2020;12(1):1–19.
 86. Ridaura VK, Faith JJ, Rey FE, et al. Gut microbiota from twins discordant for obesity modulate metabolism in mice. *Science.* 2013;341(6150):1241214.
 87. Lin CJ, Cheng YC, Chen HC, et al. Commensal gut microbiota-derived acetate and propionate enhance heart adaptation in response to cardiac pressure overload in mice. *Theranostics.* 2022;12(17):7319–7334.
 88. Krummen M, Mayerhofer CCK, Vestad B, et al. Gut microbiota signature in heart failure defined from profiling of 2 independent cohorts. *J Am Coll Cardiol.* 2018;71(10):1184–1186.
 89. Pasini E, Aquilani R, Testa C, et al. Pathogenic gut flora in patients with chronic heart failure. *JACC Heart Fail.* 2016;4(3):220–227.
 90. Yang L, Wu Y, Yang J, et al. *Lactiplantibacillus plantarum* P470 isolated from fermented Chinese chives has the potential to improve *in vitro* the intestinal microbiota and biological activity in feces of coronary heart disease (CHD) patients. *Nutrients.* 2024;16(17):2945.
 91. Sandek A, Bauditz J, Swidsinski A, et al. Altered intestinal function in patients with chronic heart failure. *J Am Coll Cardiol.* 2007;50(16):1561–1569.
 92. Krack A, Sharma R, Figulla HR, Anker SD. The importance of the gastrointestinal system in the pathogenesis of heart failure. *Eur Heart J.* 2005;26(22):2368–2374.
 93. Sandek A, Swidsinski A, Schroedl W, et al. Intestinal blood flow in patients with chronic heart failure: a link with bacterial growth, gastrointestinal symptoms, and Cachexia. *J Am Coll Cardiol.* 2014;64(11):1092–1102.
 94. Niebauer J, Volk HD, Kemp M, et al. Endotoxin and immune activation in chronic heart failure: a prospective cohort study. *Lancet.* 1999;353(9167):1838–1842.
 95. Paulus WJ. How are cytokines activated in heart failure? *Eur J Heart Fail.* 1999;1(4):309–312.
 96. Mamic P, Heidenreich PA, Hedlin H, Tennakoon L, Staudenmayer KL. Hospitalized patients with heart failure and common bacterial infections: a nationwide analysis of concomitant *Clostridium difficile* infection rates and in-hospital mortality. *J Card Fail.* 2016;22(11):891–900.
 97. Sun X, Jiao X, Ma Y, et al. Trimethylamine N-oxide induces inflammation and endothelial dysfunction in human umbilical vein endothelial cells via activating ROS-TXNIP-NLRP3 inflammasome. *Biochem Biophys Res Commun.* 2016;481(1–2):63–70.
 98. Zhu W, Gregory JC, Org E, et al. Gut microbial metabolite TMAO enhances platelet hyperreactivity and thrombosis risk. *Cell.* 2016;165(1):111–124.
 99. Wang Z, Klipfell E, Bennett BJ, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature.* 2011;472(7341):57–63.
 100. Wilson Tang WH, Wang Z, Fan Y, et al. Prognostic value of elevated levels of intestinal microbe-generated metabolite trimethylamine-N-oxide in patients with heart failure: refining the gut hypothesis. *J Am Coll Cardiol.* 2014;64(18):1908–1914.
 101. Li X, Geng J, Zhao J, et al. Trimethylamine N-oxide exacerbates cardiac fibrosis via activating the NLRP3 inflammasome. *Front Physiol.* 2019;10:866.
 102. Wang G, Kong B, Shuai W, Fu H, Jiang X, Huang H. 3, 3-Dimethyl-1-butanol attenuates cardiac remodeling in pressure-overload-induced heart failure mice. *J Nutr Biochem.* 2020;78:108341.
 103. Organ CL, Otsuka H, Bhushan S, et al. Choline diet and its gut microbe-derived metabolite, trimethylamine N-oxide, exacerbate pressure overload-induced heart failure. *Circ Heart Fail.* 2016;9(1):e002314.
 104. Li Z, Wu Z, Yan J, et al. Gut microbe-derived metabolite trimethylamine N-oxide induces cardiac hypertrophy and fibrosis. *Lab Invest.* 2019;99(3):346–357.
 105. Makrecka-Kuka M, Volska K, Antone U, et al. Trimethylamine N-oxide impairs pyruvate and fatty acid oxidation in cardiac mitochondria. *Toxicol Lett.* 2017;267:32–38.
 106. Zhang H, Meng J, Yu H. Trimethylamine N-oxide supplementation abolishes the cardioprotective effects of voluntary exercise in mice fed a western diet. *Front Physiol.* 2017;8:944.
 107. Martin-Gallausiaux C, Marinelli L, Blottière HM, Larraufie P, Lapaque N. SCFA: mechanisms and functional importance in the gut. *Proc Nutr Soc.* 2021;80(1):37–49.
 108. Nogal A, Valdes AM, Menni C. The role of short-chain fatty acids in the interplay between gut microbiota and diet in cardio-metabolic health. *Gut Microbes.* 2021;13(1):1–24.
 109. Portincasa P, Bonfrate L, Vacca M, et al. Gut microbiota and short chain fatty acids: implications in glucose homeostasis. *Int J Mol Sci.* 2022;23(3):1105.
 110. Yang W, Yu T, Huang X, et al. Intestinal microbiota-derived short-chain fatty acids regulation of immune cell IL-22 production and gut immunity. *Nat Commun.* 2020;11(1):4457.
 111. Liu P, Wang Y, Yang G, et al. The role of short-chain fatty acids in intestinal barrier function, inflammation, oxidative stress, and colonic carcinogenesis. *Pharmacol Res.* 2021;165:105420.
 112. Hu T, Wu Q, Yao Q, Jiang K, Yu J, Tang Q. Short-chain fatty acid metabolism and multiple effects on cardiovascular diseases. *Ageing Res Rev.* 2022;81:101706.
 113. Parada Venegas D, De la Fuente MK, Landskron G, et al. Short chain fatty acids (SCFAs)-mediated gut epithelial and immune regulation and its relevance for inflammatory bowel diseases. *Front Immunol.* 2019;10:277.
 114. Marques FZ, Nelson E, Chu PY, et al. High-fiber diet and acetate supplementation change the gut microbiota and prevent the development of hypertension and heart failure in hypertensive mice. *Circulation.* 2017;135(10):964–977.
 115. Carley AN, Maurya SK, Fasano M, et al. Short-chain fatty acids outpace ketone oxidation in the failing heart. *Circulation.* 2021;143(18):1797–1808.
 116. Mayerhofer CCK, Ueland T, Broch K, et al. Increased secondary/primary bile acid ratio in chronic heart failure. *J Card Fail.* 2017;23(9):666–671.

117. Desai MS, Mathur B, Eblimit Z, et al. Bile acid excess induces cardiomyopathy and metabolic dysfunctions in the heart. *Hepatology*. 2017;65(1):189–201.
118. Mao H, Angelini A, Li S, et al. CRAT links cholesterol metabolism to innate immune responses in the heart. *Nat Metab*. 2023;5(8):1382–1394.
119. Lee WY, Han SH, Cho TS, Yoo YH, Lee SM. Effect of ursodeoxycholic acid on ischemia/reperfusion injury in isolated rat heart. *Arch Pharm Res (Seoul)*. 1999;22(5):479–484.
120. Rajesh KG, Suzuki R, Maeda H, Yamamoto M, Xing Y, Sasaguri S. Hydrophilic bile salt ursodeoxycholic acid protects myocardium against reperfusion injury in a PI3K/Akt dependent pathway. *J Mol Cell Cardiol*. 2005;39(5):766–776.
121. Rani S, Sreenivasiah PK, Kim JO, et al. Tauroursodeoxycholic acid (TUDCA) attenuates pressure overload-induced cardiac remodeling by reducing endoplasmic reticulum stress. *PLoS One*. 2017;12(4):e0176071.
122. Lopaschuk GD, Karwi QG, Tian R, Wende AR, Dale Abel E. Cardiac energy metabolism in heart failure. *Circ Res*. 2021;128(10):1487–1513.
123. Zhao M, Wei H, Li C, et al. Gut microbiota production of trimethyl-5-aminovaleric acid reduces fatty acid oxidation and accelerates cardiac hypertrophy. *Nat Commun*. 2022;13(1):1757.
124. Shi B, Zhang X, Song Z, et al. Targeting gut microbiota-derived kynurenine to predict and protect the remodeling of the pressure-overloaded young heart. *Sci Adv*. 2023;9(28):eadg7417.
125. Ragni M, Greco CM, Felicetta A, et al. Dietary essential amino acids for the treatment of heart failure with reduced ejection fraction. *Cardiovasc Res*. 2023;119(4):982–997.
126. Wang YC, Koay YC, Pan C, et al. Indole-3-propionic acid protects against heart failure with preserved ejection fraction. *Circ Res*. 2024;134(4):371–389.
127. Cristofori F, Dargenio VN, Dargenio C, Miniello VL, Barone M, Francavilla R. Anti-inflammatory and immunomodulatory effects of probiotics in gut inflammation: a door to the body. *Front Immunol*. 2021;12:578386.
128. Watzenboeck ML, Drobets B, Zahalka S, et al. Lipocalin 2 modulates dendritic cell activity and shapes immunity to influenza in a microbiome dependent manner. *PLoS Pathog*. 2021;17(4):e1009487.
129. Shi XR, Chen BY, Lin WZ, et al. Microbiota in gut, oral cavity, and mitral valves are associated with rheumatic heart disease. *Front Cell Infect Microbiol*. 2021;11:643092.
130. Yan F, Brent Polk D. Probiotics and probiotic-derived functional factors—mechanistic insights into applications for intestinal homeostasis. *Front Immunol*. 2020;11:1428.
131. Gan XT, Ettinger G, Huang CX, et al. Probiotic administration attenuates myocardial hypertrophy and heart failure after myocardial infarction in the rat. *Circ Heart Fail*. 2014;7(3):491–499.
132. Pourrajab B, Naderi N, Janani L, et al. Comparison of probiotic yogurt and ordinary yogurt consumption on serum Pentraxin3, NT-proBNP, oxLDL, and ApoB100 in patients with chronic heart failure: a randomized, triple-blind, controlled trial. *Food Funct*. 2020;11(11):10000–10010.
133. Pourrajab B, Naderi N, Janani L, et al. The impact of probiotic yogurt versus ordinary yogurt on serum sTWEAK, sCD163, ADMA, LCAT and BUN in patients with chronic heart failure: a randomized, triple-blind, controlled trial. *J Sci Food Agric*. 2022;102(13):6024–6035.
134. Appel LJ, Moore TJ, Obarzanek E, et al. A clinical trial of the effects of dietary patterns on blood pressure. DASH Collaborative Research Group. *N Engl J Med*. 1997;336(16):1117–1124.
135. Wastyk HC, Fragiadakis GK, Perelman D, et al. Gut-microbiota-targeted diets modulate human immune status. *Cell*. 2021;184(16):4137–4153.e14.
136. Delgado-Lista J, Alcala-Diaz JF, Torres-Peña JD, et al. Long-term secondary prevention of cardiovascular disease with a Mediterranean diet and a low-fat diet (CORDIOPREV): a randomised controlled trial. *Lancet*. 2022;399(10338):1876–1885.
137. Dong TA, Sandesara PB, Dhindsa DS, et al. Intermittent fasting: a heart healthy dietary pattern? *Am J Med*. 2020;133(8):901–907.
138. DeFilipp Z, Bloom PP, Torres Soto M, et al. Drug-resistant *E. coli* bacteremia transmitted by fecal microbiota transplant. *N Engl J Med*. 2019;381(21):2043–2050.
139. Cerisano G, Buonamici P, Valenti R, et al. Early short-term doxycycline therapy in patients with acute myocardial infarction and left ventricular dysfunction to prevent the ominous progression to adverse remodelling: the TIPTOP trial. *Eur Heart J*. 2014;35(3):184–191.