

# EGFR-TKIs Combined with Allogeneic CD8+NKT Cell Immunotherapy to Treat Patients with Advanced EGFR-Mutated Lung Cancer

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## Abstract

**Background:** To evaluate the efficacy and safety of allogenic CD8 + natural killer T (CD8+ NKT) immunotherapy combined with gefitinib in the treatment of advanced or metastatic EGFR mutant non-small cell lung cancer (NSCLC). **Methods:** This study is prospective. The NSCLC patients with exon 19 (Ex19del) or exon 21 L858R point mutations, and response to gefitinib treatment were enrolled into the trial to be randomly assigned into the gefitinib arm and the gefitinib/NKT arm. Allogenic CD8+ NKT cells were cultured in vitro and adaptive transferred into the patients via vein in the gefitinib/NKT arm. The primary endpoint was progression-free survival (PFS). Secondary endpoint analysis included time to disease progression (TTP), overall survival (OS), levels of serum tumour markers for carcinoembryonic antigen (CEA) and alanine aminotransferase (ALT) in the blood, the response rate and safety. From July 2017 to June 2021, 19 patients were randomly assigned to the gefitinib arm (n = 8) and the gefitinib/NKT arm (n = 11). **Results:** The estimated median survival PFS in the gefitinib/NKT arm was significantly longer than that of the gefitinib arm (12 months vs 7 months). Similar results were also observed for the median TTP. Moreover, the gefitinib/NKT arm had better CEA control than the gefitinib arm. Clinical grade 3 adverse reactions occurred in 64% and 39% of patients in the gefitinib/NKT arm and the gefitinib arm, respectively. The most common grade 3 adverse events in the gefitinib/NKT arm included abnormal liver function in 8 cases (73%) and diarrhoea in 1 case (9%), both of which resolved after drug intervention. **Conclusion:** The PFS of EGFR-mutated advanced NSCLC treated with

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allogenic CD8+ NKT cells combined with gefitinib was longer than that of gefitinib alone. No obvious serious adverse reactions occurred, and the patients compliance and survival status were good.

## Keywords

NSCLC, EGFR, gefitinib, CD8, NKT cell, immunotherapy

## Abbreviations

EGFR, Epidermal growth factor receptor; NKT, Natural killer T; NSCLC, Non-small-cell lung cancer; PFS, Progression-free survival; TTP, Time to progression; OS, Overall survival; CEA, carcinoembryonic antigen; ALT, alanine aminotransferase; RECIST, Response Evaluation Criteria In Solid Tumors; SPIRIT, Standard Protocol Items: Recommendations for Interventional Trials; EGFR-TKIs, Epidermal growth factor receptor tyrosine kinase inhibitors; ICIs, immune checkpoint inhibitors; AE, Adverse event; Anti-PD-1, Anti-programmed death-1; Anti-PD-L1, Antiprogrammed death-1 ligand; TMB, tumor mutational burden; TILs, Tumour-infiltrating lymphocytes; CAR-T, Chimeric antigen receptor t; PBMCs, Peripheral blood mononuclear cells; CTCAE, Common Terminology Criteria for AEs; DCCIK, Dendritic cell-cytokine induced killer; CT, Computed tomography; CR, complete remission; SD, stable disease; PD, progressive disease; MDSCs, Myeloid-derived suppressor cells; GVHD, Graft-versus host disease; HSCs, Haematopoietic stem cells; HLAs, Human leukocyte antigens.

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## Introduction

According to recent cancer burden data, the most common cancer in China is lung cancer; which is also the leading cause of cancer death in China and in the USA.<sup>1,2</sup> Among nearly two million new cases of lung cancer diagnosed every year, more than 85% of these cases were non-small cell lung cancers (NSCLCs) (which were identified by histopathological grading), including adenocarcinoma, squamous cell carcinoma, and large cell carcinoma.<sup>3,4</sup> Regardless of clinical characteristics, various genetic screenings (such as for EGFR, ALK, BRAF) are currently recommended by ASCO in all patients with advanced lung adenocarcinoma. Cumulative data further showed that more than 40% of patients with NSCLC in the Asian population have epidermal growth factor receptor (EGFR) mutations, most of which are deletions in exon 19 (Ex19del) or exon 21 L858R point mutations.<sup>5-7</sup> Depending on the involvement of EGFR receptor tyrosine kinase inhibitors (EGFR-TKIs) on EGFR mutations in 2004, various generations of EGFR-TKIs have been developed for the sequential administration of targeted therapy and have replaced traditional platinum-based chemotherapies, such as gefitinib, afatinib and osimertinib.<sup>8</sup> Indeed, these small-molecule drugs have been proved with significant clinical responses and reduced treatment-related toxicities, and become the standard of care for the treatment of NSCLC with EGFR mutation.<sup>3</sup> Unfortunately, most of patients will acquired drug resistance to EGFR-TKIs treatment and show disease progression within approximately 10 months, including the third-generation EGFR-TKI osimertinib.<sup>9</sup> These mutations associated resistance to EGFR-TKIs generally arise due to the drug selective pressure on tumor cells in the treatment through EGFR-dependent or EGFR-indepent resistance mechanisms.<sup>9</sup> Thus, it is a significant challenge to treat advanced NSCLC with mutation to avoiding or obviously delay the emergence of acquired resistance, and maximize the benefits from EGFR-TKIs in clinic. So, the combination of EGFR-TKIs with

other target anticancer strategies, such as chemotherapy, radiotherapy, antiangiogenesis therapy and immunotherapy have been under consideration.

The cumulative data showed that EGFR-TKIs treatment has the regulation on the tumor immunological microenvironment and the EGFR activation up-regulates the expression of PD-L1 on tumor cells<sup>10-13</sup>. So, immunotherapy using immune checkpoint inhibitors (ICIs) for tumors has attracted more attention due to its benefits in the clinic and has been integrated into the treatment of NSCLC.<sup>14</sup> Unfortunately, published data also have revealed that patients with EGFR-mutant NSCLC have a poor response to ICI monotherapy, such as anti-PD-1/PD-L1 therapy.<sup>15</sup> The potential mechanisms for the poor efficacy of anti-PD-1/PD-L1 therapy in EGFR mutant NSCLC include lower expression of PD-L1 in cancer cells and low tumour mutational burden (TMB).<sup>16,17</sup> Moreover, multiple clinical trials of EGFR-TKI-immunotherapy combination therapy have been terminated because of increased toxicity or high incidence of adverse events (AEs), and indicated the limitation of anti-PD-L1 in the current treatment of NSCLC.<sup>18-20</sup> Therefore, it is necessary to seek alternative strategy for immunotherapy in the treatment of these diseases with EGFR-TKIs.

Immune cells which with cytotoxicity to tumor cells attract attention in cancer therapy for past decades, such as CD8 T cells and NK cells. Generally, EGFR-mutant NSCLC has a non-inflamed tumor microenvironment (TME) and infiltrated with low frequency of CD8+ T cells. But, tumor EGFR-TKIs treatment enhanced the immune recognition and lysis to tumor cells which mediated by antigen-specific T cells and NK cells.<sup>21-23</sup> Moreover, published data showed that EGFR-TKIs treatment reversed immunosuppression by increasing cytotoxic CD8+ T cells and dendritic cells, which was accompanied with tumor shrinkage. Unfortunately, such EGFR-TKI treatment also consistently promoted the production of myeloid-derived suppressor cells (MDSCs), and increased the levels of IL-10 and

CCL-2.<sup>24</sup> Thus, EGFR-TKIs treatment induced the change in tumor immunological microenvironment might bring about a window of opportunity for combination treatment with cellular adoptive immunotherapy in advance NSCLC. Different autologous or allogeneic immune cells, such as CD8<sup>+</sup> tumour-infiltrating lymphocytes (TILs), NK cells, and genetically-modified immune cells (CAR-T and TCR-T cells) have been cultured *in vitro* for tumour immunotherapy since the 1980s.<sup>25–28</sup> In a previous study, we also identified one CD1d-independent CD8<sup>+</sup> NK-like T subset, also called NKT cell, that expresses both T-cell markers and NK-cell receptors, and has cototoxicity to tumor through killing MDSCs *in vivo*.<sup>29</sup> We also identified the equivalent CD8<sup>+</sup> NKT (CD8 + CD56<sup>+</sup>) in the peripheral blood in humans and confirmed its anti-tumor capability (data not shown). These data further indicated its potential usage in the combination with EGFR-TKIs treatment in the advance NSCLC.

Although various EGFR-TKIs of different generations are currently used in clinic, gefitinib is a representative drug of first-generation EGFR-TKIs, and has been proven to have superior efficacy in EGFR advanced NSCLC treatment than platinum-based chemotherapy, with a response rate (RR) of 73% and PFS of 10.8 months.<sup>30,31</sup> Thus, in this study, we designed a trial to explore the potential efficiency and safety of NKT cells adoptive immunotherapy in the combination with EGFR-TKIs in the treatment of the EGFR mutated advanced NSCLC.

## Materials and Methods

### Patients

In this clinical trial, we followed the guidelines of the Declaration of Helsinki, and it was approved by the institutional review board. Eligible male or female patients were aged between 18 and 75 years old, had a histopathological diagnosis of advanced NSCLC (clinical stage III/IV) with an EGFR exon 19 deletion mutation or EGFR exon 21 L858R substitution mutation, and one or more measurable lesions. In the trial period, the patients did not receive any additional chemotherapy or radiotherapy other than that specified in this study. Complete inclusion/exclusion criteria are reported in the trial protocol.<sup>32</sup> The trial was registered in the Chinese Clinical Trial Registry (ChiCTR-IIR-17013471) (<http://www.chictr.org.cn/index.aspx>). All patients signed informed consent forms before

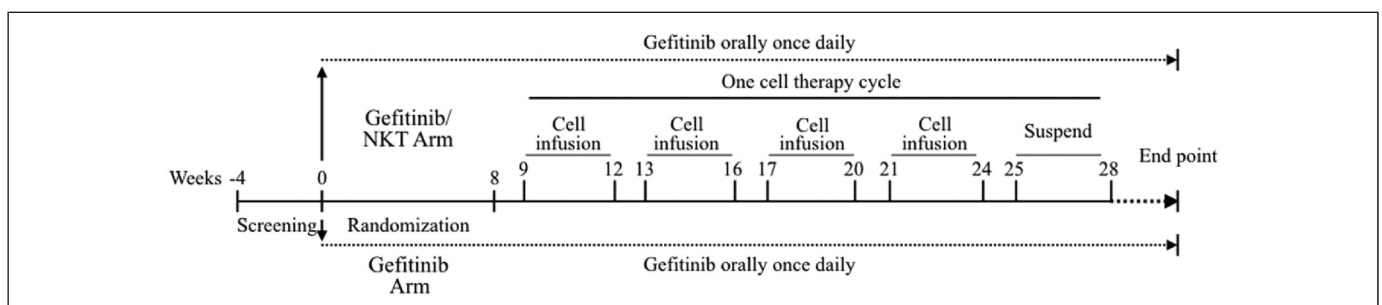
screening. The reporting of this study conforms to the CONSORT statements.<sup>33</sup>

### Study Design and Treatment

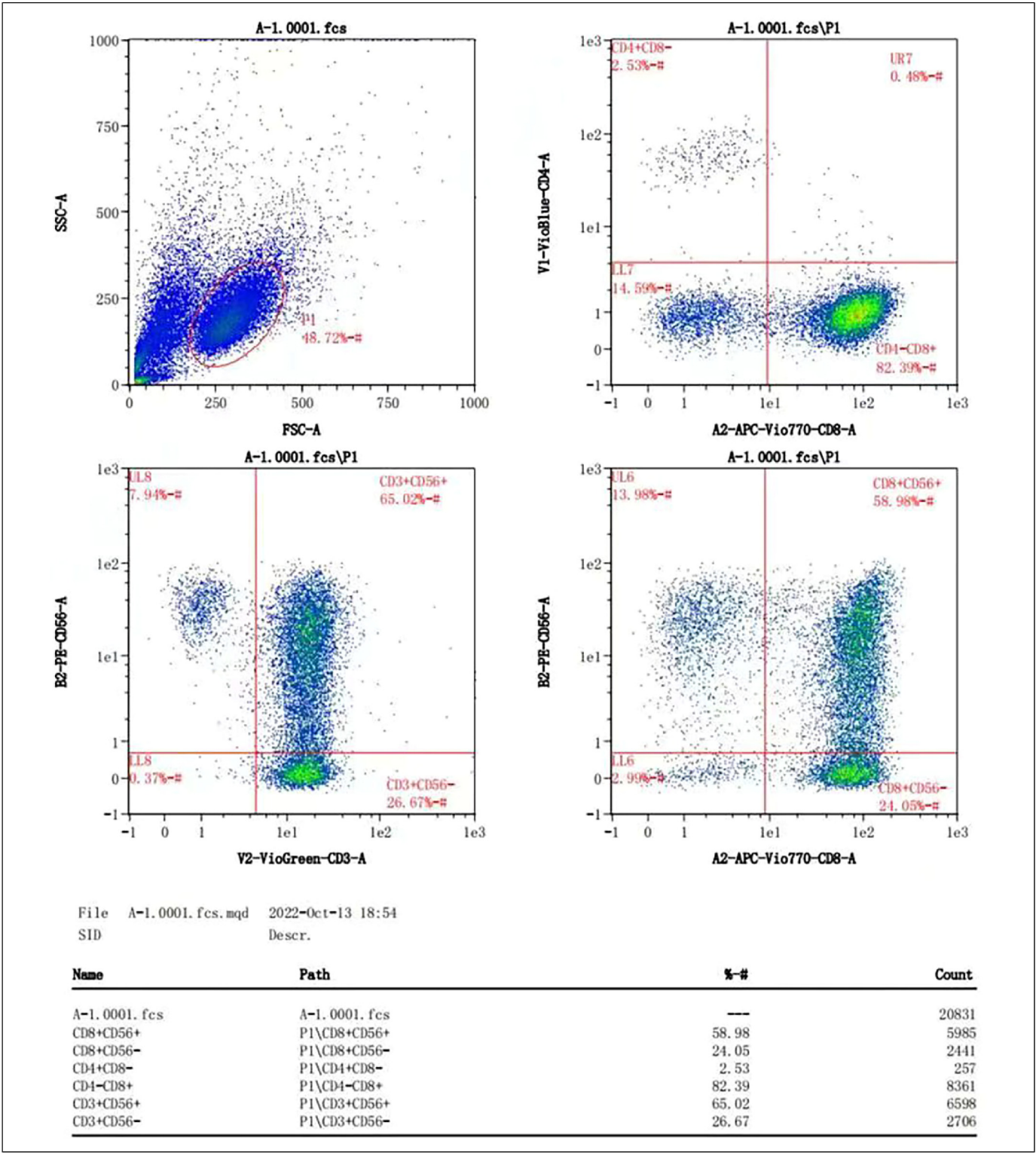
This study is prospective. Subjects were randomly divided into the gefitinib arm and gefitinib/NKT arm. After enrollment (and prior to any trial treatment), subjects were randomly assigned to the experimental group (gefitinib /NKT group) or the control group (gefitinib group) on a 1:1 ratio. And the group randomization method is used to randomly group the subjects. The subjects were divided into different block groups and then randomly assigned within each group, and tables of random numbers had been generated by statistical experts and recorded by the researchers. As the aim of this study was to explore the effects of adaptively transferred NKT cells on acquired gefitinib resistance in NSCLC, all volunteers were first treated with gefitinib for 8 weeks and then were screened the degrees of gefitinib response. Thus, the week-8 was designated as the baseline for this trial (Figure 1), and only the volunteers with no disease progression were enrolled in the study for further random allocation into the gefitinib arm and the gefitinib/NKT arm. Next, in the gefitinib arm, the patients were orally treated with 250 mg gefitinib once daily until disease progression. In the gefitinib/NKT arm, the patients were treated with gefitinib (as described in the gefitinib arm) but in combination with an infusion of a total of  $1 \times 10^{10}$  NKT cells every four weeks from the eighth week after gefitinib treatment. The diagram of the trial, the frequencies of gefitinib treatment and NKT-cell infusion are described in (Figure 1) and the protocol.<sup>32</sup>

### NKT-Cell Preparation

The immediate family members of the patient with consanguinity served as the blood donors. All donors signed informed consent forms and underwent medical examinations to ensure that they met the inclusion criteria for donors in the protocol.<sup>32</sup> peripheral blood mononuclear cells (PBMCs) from blood donors are collected using a Fresenius blood cell separator, model COM.TEC (COM). TEC cell separator for NKT-cell preparation in the trial (COM). The TEC cell separator (Fresenius Haemocare, Bad Homburg, Germany) had a total circulating blood volume of approximately 4000 ml. All PBMC samples were analyzed



**Figure 1.** The schedule for the gefitinib treatment and NKT cell adaptive transfer in the trial.



**Figure 2.** Flow analysis for cultured NKT cells. The percentage of CD8<sup>+</sup> cells among the expanded cells was analyzed by using flow cytometer, and the representative graphs were shown.

to determine the total cell number and the percentage of CD8<sup>+</sup> T cells and then cryopreserved in liquid nitrogen for further experiments.

CD8<sup>+</sup> NKT-cell preparation was accomplished by culturing allogeneic PBMCs from donors with CD8<sup>+</sup> NKT culture medium developed by the Institute of Cell Therapy of Tsinghua

University (Beijing, China). PBMCs were differentiated in T75 flasks and transferred to 2 L culture bags on Day 4 for CD8<sup>+</sup> NKT-cell expansion until cells were harvested on Days 12 and 15. The percentage of CD8<sup>+</sup> cells among the expanded cells was greater than 60%, and representative flow cytometry graphs are shown in (Figure 2) the supplementary data. The total number of cells used for patient infusion was up to 1 × 10<sup>10</sup>. The final cell product for infusion was prepared in saline with albumin.

### Clinical Evaluation

According to the improved version 1.1 of the evaluation standard for solid tumours,<sup>34</sup> imaging evaluations were performed at baseline and every 2 months thereafter until progression. Reaction, progression-free survival (PFS) and overall survival (OS) were assessed by investigators. Adverse events were graded using the National Cancer Institute CTCAE. After progression, survival information was collected every 6 months, including post-treatment information, until death.

### Study end Points

The primary end point was the time from randomization to the first recording of disease progression (imaging-based) or patient death comparing PFS between the gefitinib/NKT arm and the gefitinib arm. Secondary endpoints included OS, TTP, safety and adverse reactions. OS was defined as the time from randomization to death from any cause. The safety index is based on the CTCAE standard of the degree of adverse reactions, so adverse events are recorded regardless of the cause to ensure objective reporting.

### Statistical Analysis

The primary end point PFS was first compared to verify superiority between the two arms according to the protocol.<sup>32</sup> The test was considered effective only when it was demonstrated that the PFS of the gefitinib/NKT arm was greater than that of the gefitinib arm. The Kaplan–Meier method was used to estimate the survival curve of PFS, TTP and OS, and the log rank test was used to analyse the confidence interval. The research team considered that adverse reactions related to NKT-cell infusion, such as shivering, dizziness, fatigue, and fever, were not related to clinical outcomes. The median quartile interval was used to compare CEA

and ALT, and Fisher's exact test was used to compare the incidence of adverse reactions between the two arms.

## Results

### Patient Allocation

From July 2017 to June 2021, a total of 19 subjects were randomized. In both arms, 18 (94.7%) completed all the study visits, and 1 (5.3%) withdrew early from the study. Among them, 11 patients (57.9%, 11/19) were treated with gefitinib/NKT, and 8 patients (42.1%, 8/19) were treated with gefitinib alone. Of the 11 subjects in the cellular immunotherapy group, 10 (90.9%) completed all the study visits, and 1 (9.1%) withdrew from the trial for several reasons. Of the 10 subjects who completed the trial, 3 (30%) died of disease progression during the follow-up period after disease progression. Of the 8 subjects in the gefitinib arm, 8 (100.0%) completed all the study visits. During the follow-up period after disease progression, 1 (12.5%) died due to disease progression. Information on the patient registration and distribution is shown in (Table 1). The database was finalized on June 30, 2021.

### Demographic and Baseline Characteristics

The baseline characteristics of the patients are listed in (Table 2). There were 19 subjects, including 8 males and 11 females. In the gefitinib/NKT arm, there were 4 males (36.4%) and 7 females (63.6%). In the gefitinib arm, there were 4 males (50.0%) and 4

**Table 1.** Patient Accumulation.

N=19			
Subjects were randomly assigned			
*N=8		*N=11	
GEF Arm		GEF/NKT Arm	
N=8	N=0	N=10	N=1
Test	Fall by the	Test	Fall by the
complete	wayside	complete	wayside
*Follow-up period N=1 died		*Follow-up period N=3 died	

\*1. All patients were randomly divided into GEF Arm and GEF/NKT Arm.

2. During the follow-up period, 1 patient in the GEF Arm died and three patients in the GEF/NKT Arm died.

**Table 2.** Baseline Demographics and Clinical Characteristics.

Characteristic	GEF Arm	GEF/NKT Arm	P Value
No. of patients	8	11	0.6577
Sex			
Male	50.0%	36.4%	
Female	50.0%	63.6%	
Smoking status			0.0578
Never	62.5%	100.0%	
Previous or current smoker	37.5%	0.0%	
*KPS			0.0587
100	12.5%	63.6%	
90	87.5%	36.4%	
Bearing			0.1698
right	25.0%	63.6%	
left	75.0%	36.4%	
Histologic diagnosis			
Adenocarcinoma	100.0%	100.0%	
Squamous cell carcinomas	0.0%	0.0%	
Stage of disease			>0.9999
III	12.5%	9.1%	
IV	87.5%	90.9%	
Type of *EGFR mutation			>0.9999
Exon 19 deletion	25.0%	63.6%	
Exon 21 Leu858Arg mutation	75.0%	36.4%	

\*Abbreviations: KPS, Karnofsky, Physical condition rating sheet; EGFR, epidermal growth factor receptor.

females (50.0%). Among the smoking status, 11 people had no smoking history in GEF/NKT Arm (100%), 7 people had no smoking history in GEF Arm (62.5%), and 2 people had smoking history (37.5%). A uniform genetic test was taken before starting treatment. These patients had different histological subtypes, with exon19 mutation in GEF/NKT Arm in 7 patients (63.6%) and GEF Arm in 2 patients (25.0%). Exon 21 Leu858Arg mutation was found in 5 persons (36.4%) in GEF/NKT Arm and 6 persons (75.0%) in GEF Arm. Smoking status, KPS score, location, pathological type, stage and EGFR mutation type were compared, and Chi-square test was performed on the data, however, the difference was not statistically significant (all  $P > .05$ ).

### Treatment Process

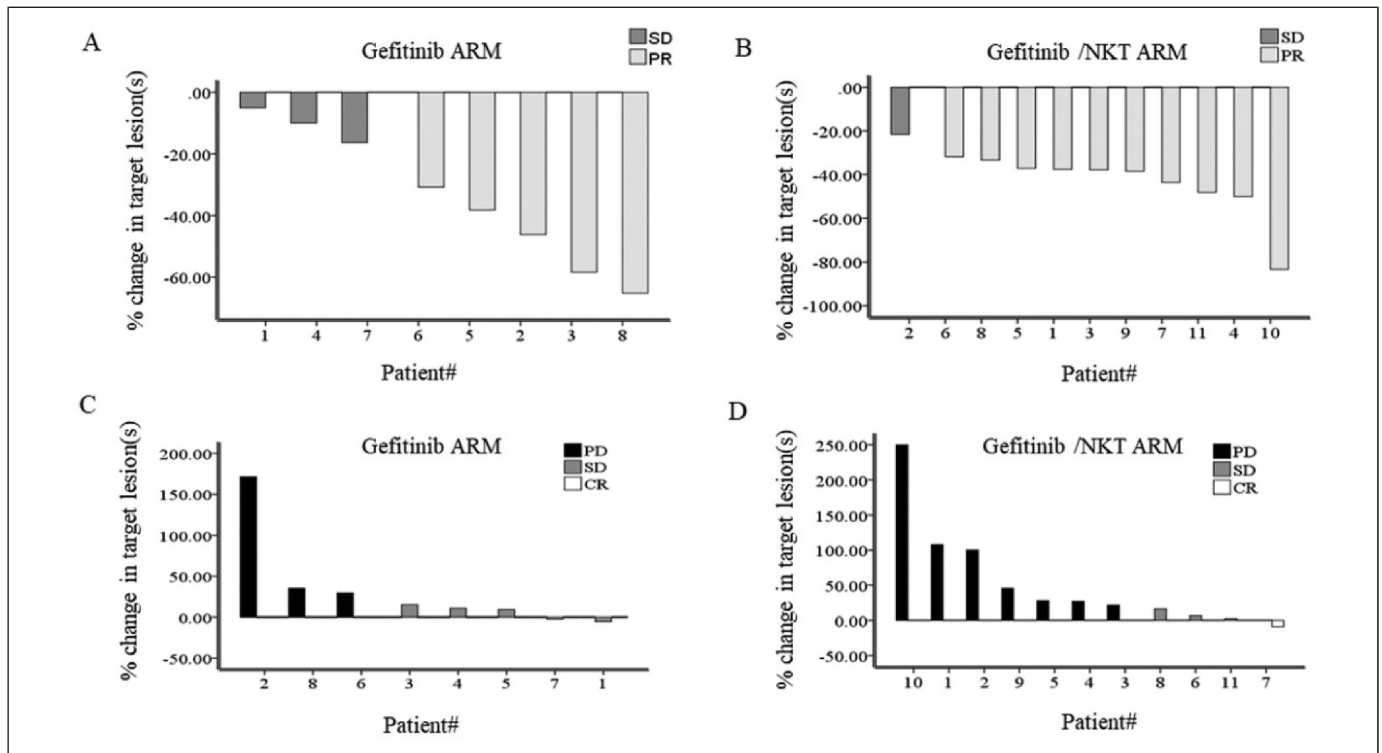
**Gefitinib arm** Of those exclusively receiving gefitinib treatment, 1 case of hypothyroidism was treated with thyroid drugs, 1 case was treated with oral antihypertensive drugs, 1 case was treated with human albumin infusion, and 1 case was treated with cisplatin (bronchial arteriography and perfusion chemoembolization).

**Gefitinib/NKT arm** According to the research flow chart as previously described,<sup>32</sup> 2 patients (18.1%) completed one

therapeutic cycle of NKT-cell infusion, four patients (36.3%) completed two therapeutic cycles, one patient (9.0%) completed three therapeutic cycles, two patients (18.1%) completed four therapeutic cycles, and two patients (18.1%) completed five therapeutic cycles. Eleven cases (100.0%) were treated with anti-allergic drugs due to NKT-cell infusion, 6 cases (54.5%) were treated with liver protection and enzyme lowering therapy with Sodium glutathione, Compound Glycyrrhizin, Silybin meglumine due to elevated ALT and abnormal liver function, 4 cases (36.3%) were treated with bone metastasis drugs with Zoledronic acid, Ibandronate due to bone metastasis, and 1 case was treated with montmorillonite powder due to diarrhoea.

### Efficiency

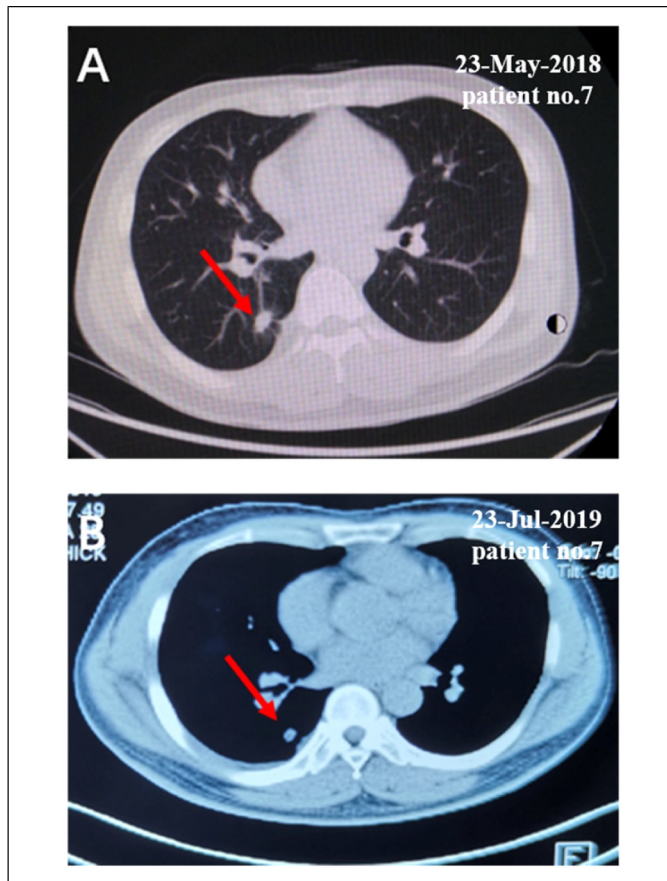
Although different degrees of response to gefitinib treatment were observed, all randomly assigned volunteers in both arms showed response to the gefitinib without significant difference in both arms in the first 8 weeks (Figure 3A and 3B). Next, the week-8 was designated as the baseline to explore the efficiency of adaptive transferred NKT cells in delaying the gefitinib resistance in the patients in this trial, and the efficiency was shown below.



**Figure 3.** The response of the volunteers to gefitinib treatment at different stages.

The maximum tumour change from baseline by the best overall response is shown, as per Response Evaluation Criteria in Solid Tumours (RECIST), version 1.1, in both the gefitinib arm and gefitinib/NKT-cell arm. (A-B) The degree of the response to the gefitinib after being treated for 8 weeks. The volunteer was screened in week 0 and then designated as baseline to initiate gefitinib treatment, and then the response to gefitinib was evaluated by comparing the lesions in week 8 with the baseline (week-0). The degrees of gefitinib response in the volunteers of (A) the gefitinib arm and (B) gefitinib/NKT arm after 8 weeks of treatment from week 0 are shown. The volunteers with no response to the gefitinib treatment were excluded. (C-D) The degree of the response to the gefitinib after being treated NKT cells. The week-8 was designated the baseline for NKT-cell treatment, and then all volunteers were divided into the gefitinib arm and gefitinib/NKT arm. The degrees of gefitinib response in the volunteers of both (C) the gefitinib arm and (D) gefitinib/NKT arm after accepting NKT-cell treatment from week 9 to the end point of the study were shown. Each bar represents the maximum change in the sum of the diameters of the target lesions of an individual patient. CR complete response, PR partial response, SD stable disease, PD progressive disease.





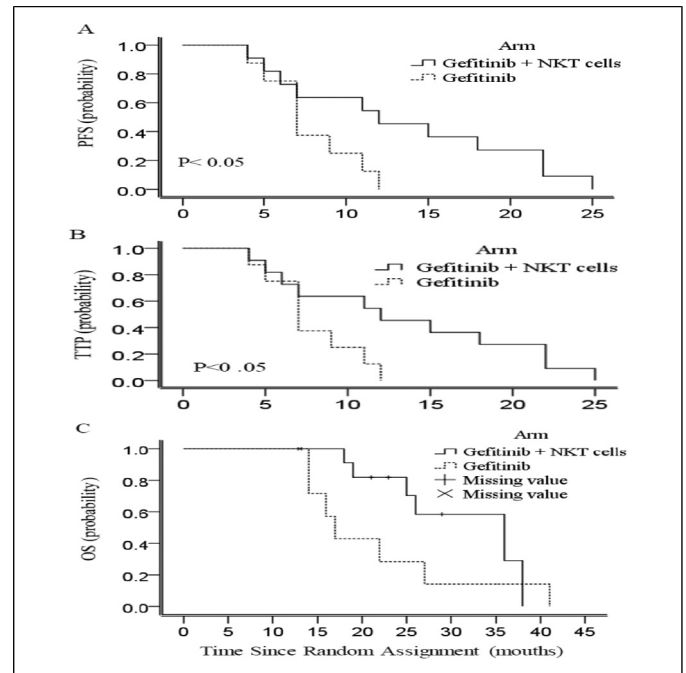
**Figure 4.** The CT scan data of the representative patient in the gefitinib/NKT arm after being treated with NKT-cell. The CT scan results at (A) the baseline and (B) three one-cell-therapy-cycles NKT cell treatment (Count up to 24 times NKT cells adaptive transfer in total 60 weeks) were shown.

**Gefitinib arm** Eight patients underwent disease assessment scans. SD was observed in 4 cases (50.0%), and PD was observed in 4 cases (50.0%) (Figure 3C).

**Gefitinib/NKT arm** Eleven patients were evaluated according to RECIST version 1.1.<sup>34</sup> All volunteers started to accept NKT treatment from week-9 except gefitinib. As shown in (Figure 3D), the best response was complete remission (CR) in 1 case (9.0%), surgical resection of the lesion, stable disease (SD) in 4 cases (36.3%), and progressive disease (PD) in 6 cases (54.5%). The CT scan images of one representative patient with partial response are shown in (Figure 4).

### Efficacy Evaluation

**Progression-free survival (PFS)** The primary endpoint was PFS. The target sample size was set to 30 patients (actually 19 patients in both arms), and withdrawal was allowed. The Kaplan–Meier method was used to estimate the median PFS, and the log rank test was used to compare between arms ( $P=.049$ ) (Figure 5A). The median PFS of the NKT combined with gefitinib arm was



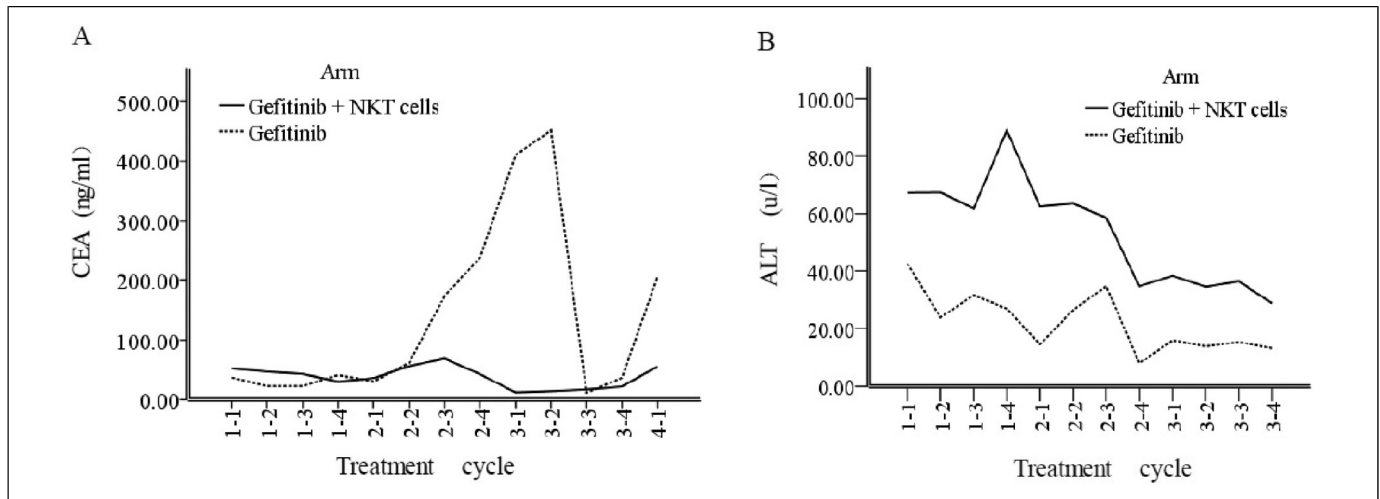
**Figure 5.** PFS, TTP and OS of the volunteers. Kaplan–Meier estimates of (A) PFS, (B) TTP and (C) OS in the full analysis set in the gefitinib arm and the gefitinib/NKT arm were shown.

12.0 months, and the median PFS of the gefitinib arm was 7.0 months. Thus, PFS was significantly prolonged in the NKT-cell treated arm.

**Time to progression (TTP)** The definition of tumour progression was the same as that of PFS. The Kaplan–Meier method was used to estimate the median TTP, and the log rank test was used to compare two arms ( $P=.049$ ) (Figure 5B). The time of disease progression in the gefitinib/NKT arm was significantly longer than that in the gefitinib arm. Moreover, the data also showed lower CEA protein levels in the NKT treatment arm (Figure 6A).

**Overall survival (OS)** Due to the limited number of cases and the immature OS data (Figure 5C), no available statistical analysis can be provided. At the end of the data collection period, 3 patients (27.2%) in the gefitinib/NKT arm died, and 1 patient (12.5%) in the gefitinib arm died.

**Security** According to the CTCAE standard, 7 cases (64%) in the gefitinib/NKT arm and 3 cases (39%) in the gefitinib arm had grade 3 adverse events. Among them, the most common grade 3 adverse events in the gefitinib/NKT arm were abnormal liver function in 8 cases (73%) and diarrhoea in 1 case (9%). The most common grade 1–2 adverse events were shivering in 2 cases (18%), dizziness in 3 cases (27%), fatigue in 8 cases (73%) and fever in 2 cases (18%). All of the adverse events were associated with NKT-cell reinfusion and relieved spontaneously after 48 h. In the gefitinib arm, there was 1 case (13%) of bleeding, 1 case (13%) of abnormal liver function, and 1 case (13%) of nausea and vomiting. Among the most common grade 1–2 adverse events, weight loss occurred in 2 cases



**Figure 6.** The levels of CEA and ALT in the serum of the volunteers.

Analysis using median quartile spacing of (A) CEA levels and (B) ALT levels in the full analysis set in the gefitinib arm and the gefitinib/NKT arm were shown.

**Table 3.** Common Adverse Events.

Toxicity	*No. of Patients (%)									
	GEF Arm (n=8)					GEF/NKT Arm (n=11)				
	ALL	Grade 1-2	Grade 3	Grade 4	Grade 5	ALL	Grade 1-2	Grade 3	Grade 4	Grade 5
Skin rash	0	0	0	0	0	1 (9%)	1 (9%)	0	0	0
Diarrhea	0	0	0	0	0	1 (9%)	0	1 (9%)	0	0
Dry skin	0	0	0	0	0	0	0	0	0	0
Paronychia	0	0	0	0	0	0	0	0	0	0
Bleeding	1 (13%)	1 (13%)	0	0	0	0	0	0	0	0
Liver function	1 (13%)	0	1 (13%)	0	0	8 (73%)	2 (18%)	6 (55%)	0	0
Skin rash	0	0	0	0	0	0	0	0	0	0
Weight loss	2 (25%)	2 (25%)	0	0	0	0	0	0	0	0
Nausea and vomiting	1 (13%)	0	1 (13%)	0	0	0	0	0	0	0
Intermittent lung disease	0	0	0	0	0	0	0	0	0	0
Shivering	0	0	0	0	0	2 (18%)	2 (18%)	0	0	0
Dizziness	0	0	0	0	0	3 (27%)	3 (27%)	0	0	0
Fatigue	0	0	0	0	0	8 (73%)	8 (73%)	0	0	0
Fever	0	0	0	0	0	2 (18%)	2 (18%)	0	0	0

\*1. Adverse events were classified according to the National Common Adverse Event Evaluation Criteria (CTCAE version 5.0).

2. Data are presented as number of patients and the percentage in the Arms (%).

(25%). No serious adverse events were reported in the gefitinib/NKT arm or gefitinib arm (Table 3).

The most common adverse event was abnormal liver function in the gefitinib/NKT arm, which mainly manifested as elevated transaminase levels (Figure 6B). Second, fatigue, dizziness and shivering appeared as immune activation reactions. The third most common adverse event included nausea, vomiting and diarrhoea in the digestive system. In the gefitinib arm, weight loss was the most common adverse event associated with the metabolic system. The next most common event was associated with the digestive system and involved liver

dysfunction and nausea and vomiting. The third most common event was bleeding. With the exception of the cases described in the adverse events section, no other obvious abnormal results with clinical significance were found.

## Discussion

In this study, we evaluate the safety and efficacy of allogenic cellular immunotherapy combined with EGFR-TKIs in the treatment of patients with advanced NSCLC and EGFR-sensitizing mutations compared with standard EGFR-TKIs monotherapy.



By treating with the first generation EGFR-TKI gefitinib simultaneously, the adoptive transfer of allogenic CD8<sup>+</sup> NKT cells dramatically increased the PFS time from 7 months in the gefitinib arm to 12 months in the gefitinib/NKT arm. Despite the lack of results regarding OS due to the limited numbers of cases, it should be noted that the patients in the gefitinib/NKT arm benefited from this immunotherapy without obvious or clinically relevant toxicity.

As a new kind of molecular targeted drug, EGFR-TKIs, including the first-generation gefitinib, represent a promising therapeutic option for NSCLC patients with EGFR mutations.<sup>35</sup> However, long-term treatment with EGFR-TKIs often results in drug resistance, which remains an unavoidable dilemma for NSCLC patients. Published data have shown that the median PFS times of patients treated with gefitinib vary from 8 to 9 months.<sup>36-38</sup> In this study, 19 subjects (8 patients in the gefitinib arm and 11 in the gefitinib/NKT arm) were assessed for more than three years, and the median PFS time was 7 months, which was slightly different from that previously reported. These variations in PFS might be attributed to the different patient populations and the small size of the samples in the studies.

It is well known that the occurrence and development of tumours are the result of the immune escape of newly mutated tumour cells. Similar mechanisms were also found in the resistance to EGFR-TKIs treatment in the advanced NSCLC. Therefore, immunotherapy is regarded as a potential effective therapy strategy in these diseases. However, low PD-1 and PD-L1 expression levels and increased B7-H4 protein levels in tumour cells result in poor therapeutic efficacy in patients with advanced EGFR-mutant NSCLC.<sup>39-42</sup> In this trial, we treated these patients by combining gefitinib with allogenic CD8<sup>+</sup> NKT cells, which were amplified from PBMCs *in vitro*. The results showed that the median PFS time was extended from 7 to 12 months in the patients (Figure 5A). Moreover, the TPP analysis results of the secondary analysis index also showed that the TPP of the gefitinib/NKT arm was significantly longer than that of the gefitinib arm (Figure 5B). Therefore, these results showed that NKT cells combined with gefitinib partially improved the progression of EGFR-TKIs drug resistance caused by treatment with gefitinib alone in EGFR-mutant NSCLC patients.

CEA is an important tumour marker in lung cancer.<sup>43</sup> In this trial, the dynamic evaluation results of CEA showed that the CEA values in the gefitinib arm fluctuated greatly. After treatment with gefitinib alone for 7 months, the levels of CEA increased significantly and peaked at the 10th month, indicating disease progression in the patients. Conversely, the CEA values were well controlled in the gefitinib/NKT arm, and the curve was smooth until the end of the trial (Figure 6A). Thus, this CEA tumour index also reflects the good control effect of CD8<sup>+</sup> NKT-cell immunotherapy on tumour load in patients.

To note, most patients in the gefitinib/NKT arm exhibited good medication compliance (80%) in the trial period. Unfortunately, during the COVID-19 epidemic in China, some patients could not go to the hospital to complete cell transfusions due to regional blockade policies in China from early

March 2020. Therefore, these patients maintained gefitinib monotherapy at home, resulting in rapid progression of the disease and even death in the follow-up period. Therefore, these unpredictable force and majeure factors affected the efficacy evaluation in this trial to some degree. Moreover, the insufficient number of recruited volunteers in the trial directly impacted the OS evaluation.

This pilot study was also designed to assess the safety of CD8<sup>+</sup> NKT-cell adoptive therapy. The clinical results indicated that this cellular immunotherapy offers potential clinical benefits with minimal adverse reactions. It has been reported that the most common side effects of gefitinib treatment include moderate degrees of rash, diarrhoea, nausea and vomiting, and these side effects typically occur in the first month of treatment. In this trial, no rash or diarrhoea occurred in the gefitinib arm, and only one patient developed nausea and vomiting. In the gefitinib/NKT arm, one patient developed skin rash and diarrhoea in the early gefitinib treatment period, and NKT-cell transfusion tended to aggravate diarrhoea and caused withdrawal from the trial. However, the direct effects and potential mechanisms of allogenic CD8<sup>+</sup> NKT-cell treatment on diarrhoea need further exploration in the future. Moreover, in this study, chills (2 cases), dizziness (3 cases), fatigue (8 cases) and fever (2 cases) were observed in some patients in the gefitinib/NKT arm.

Published data have shown that gefitinib treatment alone typically has no obvious liver damage effects,<sup>44</sup> which was also confirmed by the data (Figure 6B) in this trial. Generally, our preliminary clinical data revealed no abnormal or intolerable changes in liver and kidney function caused by CD8<sup>+</sup> NKT-cell infusion (data now shown). Herein, it should be noted that 8 patients in the gefitinib/NKT arm had elevated ALT levels (Figure 6B), indicating that this combined strategy caused mild liver injury. This might be due to the rich distribution of CD8<sup>+</sup> NKT cells in the liver after donor infusion, which can cause cytotoxic activation.

CD8<sup>+</sup> NKT cells are immune cells that express granzyme and perforin and exhibit high levels of IFN- $\gamma$  secretion. Therefore, transferred CD8<sup>+</sup> NKT cells exhibit direct cytotoxic effects to kill tumour cells and myeloid-derived suppressor cells (MDSCs) to inhibit tumour growth.<sup>45</sup> The impact of HLA allele mismatch on transplant outcomes is associated with an increased incidence of graft-versus-host disease in immunocompetent recipients and acute graft-versus-host disease (GVHD) in haploidentical stem cell transplantation.<sup>46</sup> These alloreactive reactions are mainly mediated by the activation of T cells from the donor and the host. Thus, allogenic CD8<sup>+</sup> NKT-cell transfer can also induce alloreactivity *in vivo* to upregulate the efficiency of antitumor immunity and contribute to the benefits and adverse reactions of patients with NSCLC. Of note, the mismatch between the donor and the recipient potentially leads to hypersensitivity in the patients. Therefore, to prevent hypersensitivity, anti-allergic drugs should be simultaneously administered during cell transfusion. Fortunately, in this trial, no patients experienced serious allergic reactions, and only a few patients experienced low to moderate adverse reactions, such as fever, chills, and dizziness.

Since the end of our study is in 2021, most of the patients who participated in our study were infected with COVID-19. Although we did not actually study the relationship between COVID-19 and lung cancer progression, there is still some literature supporting the relationship between COVID-19 and lung cancer progression. So these results of the study were also potentially affected by COVID-19.

There have been other case reports where EGFR TKI's and ALK TKI's were continued in patients with active COVID 19 infections, but patients in these reports had mild infection and did not require intensive care, unlike patients in this study.<sup>47,48</sup> The factors favoring the good response in patients could be the younger age and nonsmoker status in this study and results may be different for older patients who are smokers in many published works. However starting and continuing TKI therapy in driver mutated lung cancer warrants further studies.<sup>49</sup>

On the other hand, due to the complexity of cell preparation, rigorous individualization, and early stage of clinical trials, only a small number of patients can be recruited for trials to evaluate the initial effectiveness and safety, resulting in a small sample size bias.

Currently, though the first generation (gefitinib and erlotinib), the second generation (afatinib and dacomitinib) EGFR-TKIs and the third generation EGFR-TKIs, including osimertinib, have been developed, it is not easy to select the optimal therapy for the first-line treatment. Therefore, more and more attention has been paid to the orderly design of optimization strategies for various EGFR-TKI applications in clinical practice.<sup>50</sup> In oncology, a universally principle of "best therapy first" or "hit hard and early" have been accepted. At the earliest stage of EGFR-TKIs treatment in the advance NSCLC, these small molecular target drugs significantly induce tumor regression and gain disease control for a long period of time. Thus, such treatment provides one time window to receive efficacious combination therapy before patient status deteriorates. Considering the direct impact on the tumor immunological microenvironment after EGFR-TKIs treatment, a safe combination of immunotherapy and EGFR-TKIs is available in the first-line application. Unfortunately, a growing number of clinical trials for immunotherapy, such as using durvalumab and nivolumab, recruit the patients with well-tolerance to EGFR-TKIs, and such delaying treatment may limit overall benefit of the entire therapy.<sup>19,51</sup> In this study, we combined CD8+ NKT cell adoptive transfer with EGFR-TKIs at the opening phase, and the data declared that this synergistic action lifted benefit of the entire therapy, including the safety and efficacy. Thus, it indicated that the treatment using CD8+ NKT cells adoptive transfer combine with EGFR-TKIs may be a potential strategy for the advanced NSCLC with EGFR mutation.

## Conclusions

In summary, we combined allogenic CD8<sup>+</sup> NKT-cell immunotherapy with EGFR-TKI (gefitinib) for EGFR-mutant NSCLC treatment and demonstrated that this synergistic effect benefit patients with prolonged PFS, and no serious adverse reactions.

The data suggest the potential usage of this therapeutic regimen in EGFR-mutant NSCLC. However, more trials are still needed for confirmation, and the underlying mechanisms require further exploration.

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## Availability of Data and Materials

The raw data supporting the conclusions of this article will be made available by the authors with <https://www.jianguoyun.com/p/DQ-B3oQsJjtCRjOnqsE> and <https://www.jianguoyun.com/p/DQxRgdQQsJjtCRjVnqsE>

## Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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## Ethics Approval and Consent to Participate

All subjects gave their informed consent for inclusion before they participated in the study. The study was conducted in accordance with the Declaration of Helsinki. Institutional review board approval was obtained for the trial from the Biomedical Research Ethics Committee of affiliated hospital of Jiangsu University (Approved No. of ethic committee: SWYXLL20170904). This Trial was registered on 21/11/2017 with [www.chictr.org.cn](http://www.chictr.org.cn), ChiCTR-IIR-17013471. Written informed consent is obtained from all participants.


## Consent

All participants provide their written informed consent forms before their treatment of Gefitinib or Gefitinib/NKT cell. This study does not contain any individual person information of patients, and all patients have consented to participate in this study and publish this paper.

## Trial Registration Number

This trial (Phase I/II Trails of NKT Cell in Combination With Gefitinib For Non Small Cell Lung Cancer) was registered on 21/11/2017 with [www.chictr.org.cn](http://www.chictr.org.cn), ChiCTR-IIR-17013471.

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## Supplemental Material

Supplemental material for this article is available online.

## References

1. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2022. *CA Cancer J Clin*. 2022;72(1):7-33.
2. Xia C, Dong X, Li H, et al. Cancer statistics in China and United States, 2022: Profiles, trends, and determinants. *Chin Med J*. Feb 9 2022;135(5):584-590. doi:10.1097/cm9.0000000000002108
3. Planchard D, Popat S, Kerr K, et al. Metastatic non-small cell lung cancer: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Annals of Oncology : Official Journal of the European Society for Medical Oncology*. Oct. 1 2018;29(Suppl 4):iv192-iv237. doi:10.1093/annonc/mdy275
4. Zhao C, Christine M, Hammerman PS, et al. Non-small-cell lung cancers: a heterogeneous set of diseases. *Nat Rev Cancer*. 2014 Aug;14(8):535-546. doi: 10.1038/nrc3775
5. Cho J, Chen L, Sangji N, et al. Cetuximab response of lung cancer-derived EGF receptor mutants is associated with asymmetric dimerization. *Cancer Res*. Nov 15 2013;73(22):6770-6779. doi:10.1158/0008-5472.Can-13-1145
6. Han B, Tjulandin S, Hagiwara K, et al. EGFR mutation prevalence in Asia-Pacific and Russian patients with advanced NSCLC of adenocarcinoma and non-adenocarcinoma histology: The IGNITE study. *Lung Cancer*. 2017;113(37):37-44.
7. Shi Y, Au SK, Thongprasert S, et al. A prospective, molecular epidemiology study of EGFR mutations in Asian patients with advanced non-small-cell lung cancer of adenocarcinoma histology (PIONEER). *J Thorac Oncol*. 2014;9(2):154-162.
8. Shah R, Lester JF. Tyrosine kinase inhibitors for the treatment of EGFR mutation-positive non-small-cell lung cancer: A clash of the generations. *Clin Lung Cancer*. 2020;21(3):e216-e228.
9. Minari R, Bordi P, Tiseo M. Third-generation epidermal growth factor receptor-tyrosine kinase inhibitors in T790M-positive non-small cell lung cancer: Review on emerged mechanisms of resistance. *Transl Lung Cancer Res*. Dec 2016;5(6):695-708. doi:10.21037/tlcr.2016.12.02
10. Yang CY, Liao WY, Ho CC, et al. Association between programmed death-ligand 1 expression, immune microenvironments, and clinical outcomes in epidermal growth factor receptor mutant lung adenocarcinoma patients treated with tyrosine kinase inhibitors. *European Journal of Cancer (Oxford, England : 1990)*. Jan 2020;124:110-122. doi:10.1016/j.ejca.2019.10.019
11. Isomoto K, Haratani K, Hayashi H, et al. Impact of EGFR-TKI treatment on the tumor immune microenvironment in EGFR mutation-positive non-small cell lung cancer. *Clinical Cancer Research : An Official Journal of the American Association for Cancer Research*. Apr 15 2020;26(8):2037-2046. doi:10.1158/1078-0432.Ccr-19-2027
12. Hsu KH, Huang YH, Tseng JS, et al. High PD-L1 expression correlates with primary resistance to EGFR-TKIs in treatment naïve advanced EGFR-mutant lung adenocarcinoma patients. *Lung Cancer*. Jan 2019;127:37-43. doi:10.1016/j.lungcan.2018.11.021
13. Okita R, Maeda A, Shimizu K, Nojima Y, Saisho S, Nakata M. PD-L1 overexpression is partially regulated by EGFR/HER2 signaling and associated with poor prognosis in patients with non-small-cell lung cancer. *Cancer Immunology, Immunotherapy : CII*. Jul 2017; 66(7):865-876. doi:10.1007/s00262-017-1986-y
14. Passaro A, Atili I, Morganti S, Signore ED, Marinis FD. Clinical features affecting survival in metastatic NSCLC treated with immunotherapy: A critical review of published data. *Cancer Treat Rev*. 2020;89:102085. doi:10.1016/j.ctrv.2020.102085
15. Lee CK, Man J, Lord S, et al. Checkpoint inhibitors in metastatic EGFR-mutated non-small cell lung cancer—A meta-analysis. *J Thorac Oncol*. 2017;12(2):403-407. doi:10.1016/j.jtho.2016.10.007
16. Ji M, Liu Y, Li Q, et al. PD-1/PD-L1 expression in non-small-cell lung cancer and its correlation with EGFR/KRAS mutations. *Cancer Biol Ther*. 2016;17(4):407-413. doi:10.1080/15384047.2016.1156256
17. Michael O, Hira R, Megan T, et al. Tumor mutation burden and efficacy of EGFR-tyrosine kinase inhibitors in patients with EGFR-mutant lung cancers. *Clin Cancer Res*. 2019/02/01;25(3): 1063-1069. 2018. doi:10.1158/1078-0432.CCR-18-1102
18. Gourd K. Astrazeneca halts two lung cancer drug trials. *Lancet Respir Med*. Dec 2015;3(12):926. doi:10.1016/s2213-2600(15) 00464-6
19. Yang JC, Shepherd FA, Kim DW, et al. Osimertinib plus durvalumab versus osimertinib monotherapy in EGFR T790M-positive NSCLC following previous EGFR TKI therapy: CAURAL brief report. *Journal of Thoracic Oncology : Official Publication of the International Association for the Study of Lung Cancer*. May 2019;14(5):933-939. doi:10.1016/j.jtho.2019.02.001
20. Gettinger S, Hellmann MD, Chow LQM, et al. Nivolumab plus erlotinib in patients with EGFR-mutant advanced NSCLC. *Journal of Thoracic Oncology : Official Publication of the International Association for the Study of Lung Cancer*. Sep 2018;13(9):1363-1372. doi:10.1016/j.jtho.2018.05.015
21. Dominguez C, Tsang KY, Palena C. Short-term EGFR blockade enhances immune-mediated cytotoxicity of EGFR mutant lung cancer cells: Rationale for combination therapies. *Cell Death Dis*. Sep 29 2016;7(9):e2380. doi:10.1038/cddis.2016.297
22. He S, Yin T, Li D, et al. Enhanced interaction between natural killer cells and lung cancer cells: Involvement in gefitinib-mediated immunoregulation. *J Transl Med*. Aug 12 2013;11:186. doi:10.1186/1479-5876-11-186
23. Nishii K, Ohashi K, Tomida S, et al. CD8+ T-cell responses are boosted by dual PD-1/VEGFR2 blockade after EGFR inhibition in egfr-mutant lung cancer. *Cancer Immunol Res*. Sep 1 2022;10(9):1111-1126. doi:10.1158/2326-6066.Cir-21-0751
24. Jia Y, Li X, Jiang T, et al. EGFR-targeted therapy alters the tumor microenvironment in EGFR-driven lung tumors: Implications for combination therapies. *Int J Cancer*. Sep 1 2019;145(5):1432-1444. doi:10.1002/ijc.32191
25. Abel AM, Chao Y, Thakar MS, Subramaniam M. Natural killer cells: Development, maturation, and clinical utilization. *Front Immunol*. 2018;9:1869. doi:10.3389/fimmu.2018.01869
26. Dani?Lle K, Marianne H, Kuppen P. The role of natural killer T cells in cancer—A phenotypical and functional approach. *Front Immunol*. 2018;9:367. doi:10.3389/fimmu.2018.00367
27. Jena B, Dotti G, Cooper LJN. Redirecting T-cell specificity by introducing a tumor-specific chimeric antigen receptor. *Blood*. 2010;116(7):1035-1044. doi:10.1182/blood-2010-01-043737

28. Stevens D, Ingels J, Lint SV, Vandekerckhove B, Vermaelen K. Dendritic cell-based immunotherapy in lung cancer. *Front Immunol.* 2021;11:620374. doi:10.3389/fimmu.2020.620374
29. Li Z, Wu Y, Wang C, Zhang M. Mouse CD8(+)NKT-like cells exert dual cytotoxicity against mouse tumor cells and myeloid-derived suppressor cells. *Cancer Immunology, Immunotherapy : CII.* Aug 2019;68(8):1303-1315. doi:10.1007/s00262-019-02363-3
30. Gaafar RM, Surmont VF, Scagliotti GV, et al. A double-blind, randomised, placebo-controlled phase III intergroup study of gefitinib in patients with advanced NSCLC, non-progressing after first line platinum-based chemotherapy (EORTC 08021/ILCP 01/03). *Eur J Cancer.* 2011 Oct;47(15):2331-2340.
31. Maemondo M, Inoue A, Kobayashi K, et al. Gefitinib, chemotherapy, non-small-cell lung cancer, EGFR. *N Engl J Med.* 2010; 362(25):2380-2388.
32. Yu W, Ye F, Yuan X, Ma Y, Xia S. A phase I/II clinical trial on the efficacy and safety of NKT cells combined with gefitinib for advanced EGFR-mutated non-small-cell lung cancer. *BMC Cancer.* 2021;21(1):877. doi:10.1186/s12885-021-08590-1
33. Schulz KF, Altman DG, Moher D, et al. CONSORT 2010 statement: updated guidelines for reporting parallel group randomised trials. *Br Med J.* 2010 Mar 23;340:332.
34. Eisenhauer EA, Therasse P, Bogaerts J, et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). *Eur J Cancer.* 2009;45(2):0-247. doi:10.1016/j.ejca.2008.10.026
35. Pao W, Chmielecki J. Rational, biologically based treatment of EGFR-mutant non-small-cell lung cancer. *Nat Rev Cancer.* 2010;10(11):760-774. doi:10.1038/nrc2947
36. Patil VM, Noronha V, Joshi A, et al. Phase III study of gefitinib or pemetrexed with carboplatin in EGFR-mutated advanced lung adenocarcinoma. *Esmo Open.* 2017;2(1):e000168. doi:10.1136/esmoopen-2017-000168
37. Paz-Ares L, Soulières D, Moecks J, Bara I, Klughammer B. Pooled analysis of clinical outcome for EGFR TKI-treated patients with EGFR mutation-positive NSCLC. *Journal of Cellular & Molecular Medicine.* 2014;18(8):1519-1539. doi:10.1111/jcmm.12278
38. Sanford M, Scott LJ. Gefitinib: A review of its use in the treatment of locally advanced/metastatic non-small cell lung cancer. *Drugs.* 2009;69(16):2303-2328. doi:10.2165/10489100-000000000-00000
39. To KKW, Fong W, Cho WCS. Immunotherapy in treating EGFR-mutant lung cancer: Current challenges and new strategies. *Front Oncol.* 2021 May 25;11:635007. doi:10.3389/fonc.2021.635007
40. Chen N, Fang W, Zhan J, Hong S, Li Z. Upregulation of PD-L1 by EGFR activation mediates the immune escape in EGFR-driven NSCLC: Implication for optional immune targeted therapy for NSCLC patients with EGFR mutation. *J Thorac Oncol Official Publ Int Assoc Study Lung Cancer.* 2015;10(6):910-923. doi:10.1097/JTO.0000000000000500
41. D'Incecco MA A, Ludovini V, Rossi E, et al. PD-1 and PD-L1 expression in molecularly selected non-small-cell lung cancer patients. *Br J Cancer.* 2015;112(1):95-102. doi:10.1038/bjc.2014.555
42. Lu Y, Wu F, Cao Q, et al. B7-H4 is increased in lung adenocarcinoma harboring EGFR-activating mutations and contributes to immunosuppression. *Oncogene.* Jan 2022;41(5):704-717. doi:10.1038/s41388-021-02124-6
43. Grunnet M, Sorensen JB. Carcinoembryonic antigen (CEA) as tumor marker in lung cancer. *Lung Cancer.* 2012;76(2):138-143. doi:10.1016/j.lungcan.2011.11.012
44. Justyna Rawluk CFW. Gefitinib. *Recent Results Cancer Res.* 2018;211:235-246. doi:10.1007/978-3-319-91442-8\_16
45. Li Z, Wu Y, Wang C, Zhang M. Mouse CD8 + NKT-like cells exert dual cytotoxicity against mouse tumor cells and myeloid-derived suppressor cells. *Springer Open Choice.* 2019;68(8): 1303-1315. doi:10.1007/s00262-019-02363-3
46. Baumeister S, Rambaldi B, Shapiro RM, Romee R. Key aspects of the immunobiology of haploidentical hematopoietic cell transplantation. *Front Immunol.* 2020 Feb 14;11:191. doi:10.3389/fimmu.2020.00191
47. Zhang H, Xie C, Huang Y. Treatment and outcome of a patient with lung cancer infected with severe acute respiratory syndrome coronavirus-2. *J Thorac Oncol.* 2020 May;15(5): e63-e64.
48. Leonetti A, Facchinetti F, Zielli T, Brianti E, Tiseo M. COVID-19 in lung cancer patients receiving ALK/ROS1 inhibitors. *Eur J Cancer.* 2020 Jun;132:122-124.
49. Kolluri S, Panda SS, Panda A, Singh SK. Management and outcome of EGFR mutant lung cancer with SARS-CoV-2 infection. *J Cancer Res Ther.* 2023 Jul-Sep;19(5):1454-1456. doi: 10.4103/jcrt.JCRT\_1205\_21
50. Gelatti A, Drilon A, Santini FC. Optimizing the sequencing of tyrosine kinase inhibitors (TKIs) in epidermal growth factor receptor (EGFR) mutation-positive non-small cell lung cancer (NSCLC). *Lung Cancer.* 2019 Nov;137:113-122.
51. Spigel DR, Reynolds C, Waterhouse D, et al. Phase 1/2 study of the safety and tolerability of nivolumab plus crizotinib for the first-line treatment of anaplastic lymphoma kinase translocation - positive advanced non-small cell lung cancer (CheckMate 370). *Journal of Thoracic Oncology : Official Publication of the International Association for the Study of Lung Cancer.* May 2018;13(5):682-688. doi:10.1016/j.jtho.2018.02.022