### **REVIEW**



# Mesenchymal Stem Cells: A Therapeutic Approach in Fertility Restoration in Premature Ovarian Insufficiency

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#### **Abstract**

Primary ovarian insufficiency (POI) represents the cessation of ovarian function before age 40 due to follicular depletion or dysfunction. Affecting 1% of women, POI causes infertility and systemic health complications. Current treatments focus on symptom management rather than restoring ovarian function. Mesenchymal stem cells (MSCs) offer promising regenerative potential through paracrine activity, immunomodulation, and tissue repair mechanisms. MSCs from various sources have demonstrated the ability to improve ovarian function, increase follicular survival, and restore hormone production in preclinical models. MSC-derived extracellular vesicles are emerging as cell-free alternatives with similar therapeutic effects. This review examines MSC mechanisms in ovarian restoration, evaluates current evidence, and discusses challenges in clinical translation for POI treatment.

**Keywords** Premature ovarian dysfunction · Mesenchymal stem cells · Fertility restoration · Exosomes · Extracellular vesicles · Ovarian rejuvenation · Regenerative medicine

### Introduction

Globally, reproductive aging is increasing and is closely linked to aging in general [1]. Premature ovarian insufficiency (POI) is a severe form of reproductive aging that has primarily had an unexplained origin up to this point, which has limited its therapeutic applicability and resulted in significant personal and financial expenses [2, 3]. Before the age of 40, POI is a clinical illness marked by biochemical proof of ovarian insufficiency and loss of ovarian function,

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which is indicated by irregular menstruation periods. One to three% of women in the general population suffer from POI [3]. The incidence of POI is age-specific; it affects 1 in 250 women by the age of 35 and 1 in 100 by the age of 40 [4]. The clinical sequelae of POI extend far beyond infertility, impacting neurological, psychological (including increased risk of depression and anxiety), cardiovascular (due to estrogen deficiency), sexual, and bone health (leading to osteoporosis), thereby posing multifaceted challenges for both patients and healthcare providers (HCPs) [5]. These multifaceted complications pose significant challenges for both patients and healthcare providers [6].

Regenerative medicine shows promise for treating pathological illnesses that currently lack effective cures [7]. Stem cells form the foundation of regenerative medicine approaches. These cells are derived from multicellular organisms and possess the ability to differentiate into several cell types (potency) while also producing more of their own kind (self-renewal) [8, 9]. They represent groups of unspecialized cells with the capacity to develop into distinct cellular subtypes.

Mesenchymal stem cells (MSCs) have garnered significant interest in cell therapy and regenerative medicine due to their self-renewal capacity, differentiation potential, and



immunomodulatory properties [10]. MSCs primarily exert their therapeutic effects through paracrine mechanisms. They secrete bioactive molecules including growth factors, cytokines, chemokines, and extracellular vesicles that collectively modulate the tissue microenvironment, reduce inflammation, inhibit apoptosis, promote angiogenesis, and stimulate endogenous repair processes [11].

The application of MSCs in POI treatment has shown considerable promise in preclinical studies using various animal models. MSC transplantation has been demonstrated to improve ovarian reserve, enhance follicular development, restore hormone production, and even lead to successful pregnancies in these models [12, 13]. These beneficial effects are attributed to MSCs' ability to home to damaged ovarian tissue, reduce local inflammation, promote survival of existing follicles, stimulate angiogenesis, and potentially activate dormant primordial follicles [10, 14–17].

POI can result from various causes including genetic factors (e.g., Turner syndrome, fragile X premutation, mutations in genes involved in follicular development), autoimmune disorders, iatrogenic causes (chemotherapy, radiotherapy, ovarian surgery), environmental factors, and idiopathic causes which still account for the majority of cases [3, 18]. This review provides a comprehensive overview of the current understanding and therapeutic potential of MSCs in restoring fertility and ovarian function in women with POI. We explore the pathophysiology of POI, the diverse regenerative mechanisms of MSCs, and their potential to rejuvenate ovarian function. Additionally, we examine the existing preclinical and emerging clinical evidence supporting MSC-based therapies, discuss current challenges in their application, and highlight future directions for research.

Fig. 1 Pathophysiology of Premature ovarian insufficiency (POI). The figure shows the difference between a normal ovary and POI. The normal ovary has the potential to form active primordial follicles while in POI the ovary loses all primordial follicles (early eggs). Adopted from [30] (Created in https://Bio-Render.com)

### **Normal Ovary POI Ovary** Genetic Depleted follicular pool **Autoimmune** latrogenic **Environmental** Dysfunction primary follicle secondary increased follicle atresia

# Pathophysiology and Etiology of Premature Ovarian Insufficiency

Premature ovarian insufficiency (POI) is a heterogeneous disorder characterized by the loss of ovarian activity before the age of 40 (Fig. 1) [19]. While the clinical presentation often involves amenorrhea or oligomenorrhea, hypoestrogenism, and elevated gonadotropin levels (FSH>25–40 IU/L on two occasions at least 4 weeks apart), the underlying pathophysiology is complex and involves a premature depletion of the ovarian follicular pool or dysfunction of existing follicles (Fig. 1) [20, 21].

Normally, a woman is born with a finite number of primordial follicles, which gradually decline throughout her reproductive life until menopause [22, 23]. In POI, this process is drastically accelerated. The mechanisms leading to POI can be broadly categorized as [24, 25] (1) Accelerated follicular atresia, which is the most common underlying mechanism, where the rate of follicular death (atresia) is significantly increased, leading to a rapid exhaustion of the ovarian reserve. Factors contributing to accelerated atresia include genetic defects, autoimmune processes, and exposure to gonadotoxic agents [26, 27]. (2) Follicular Dysfunction: In some cases, follicles may be present in the ovaries but fail to respond appropriately to gonadotropin stimulation or to mature properly. This can be due to defects in gonadotropin receptors, signaling pathways, or oocytegranulosa cell communication [28, 29].

### **Etiological Factors**

The etiology of POI is diverse and often multifactorial. In a large percentage of cases (up to 75–90% in some series), the cause remains idiopathic (unknown) despite extensive



Table 1 Etiological factors of premature ovarian insufficiency

Genetic Factors: These account for a significant proportion of known causes [24].

Chromosomal Turner syndrome (45, X and its variants)
Abnormalities is a classic example. X chromosome dele-

lead to POI [31].

Single Gene Mutations Mutations in numerous genes involved in

ovarian development, follicle maturation, meiosis, DNA repair, and hormone synthesis/action have been implicated. Examples include mutations in FMR1 (Fragile X premutation), BMP15, GDF9, NOBOX, FIGLA, FSHR, LHCGR, NR5A1 (SF1), and genes involved in DNA repair pathways (e.g., BRCA1, BRCA2, MCM8,

tions, translocations, or mosaicism can also

MCM9) [32, 33].

Autoimmune Disorders: Autoimmune oophoritis, where the immune system mistakenly attacks ovarian tissue (targeting oocytes, granulosa cells, or theca cells), can lead to POI. This can occur as an isolated condition or in association with other autoimmune diseases, such as autoimmune thyroiditis, Addison's disease, type 1 diabetes mellitus, or systemic lupus erythematosus. The presence of anti-ovarian antibodies or lymphocytic infiltration in ovarian biopsies supports this diagnosis, though antibody testing has limitations in sensitivity and specificity [34, 35].

Iatrogenic Causes: Medical treatments can inadvertently damage the ovaries [36–38].

Chemotherapy Alkylating agents are particularly gonadotoxic, but other chemotherapeutic drugs

can also impair ovarian function.

Radiotherapy Pelvic irradiation can destroy ovarian fol-

licles, with the extent of damage depending on the radiation dose and the patient's age.

Ovarian Surgery Procedures such as bilateral oophorectomy, or even surgeries for benign conditions like endometriomas, can reduce ovarian reserve

or compromise blood supply, potentially leading to POI.

Environmental Factors and Infections [39, 40]:

Toxins Exposure to certain environmental toxins,

such as cigarette smoke, pesticides, and industrial chemicals, has been linked to ovarian damage and earlier menopause, potentially contributing to POI in suscep-

tible individuals.

Infections Viral infections like mumps oophoritis (though rare with vaccination) have been

implicated. The role of other infections is less clear but remains an area of research.

Metabolic Disorders: Galactosemia, a rare metabolic disorder, can lead to POI if not treated early with a galactose-restricted diet [41].

investigation [27]. However, several known causes have been identified as shown in Table 1.

However, the mechanisms by which MSCs exert their therapeutic effects in POI are multifaceted and extend beyond this single pathway [42, 43] (Table 2).

Understanding these diverse mechanisms is critical for optimizing MSC-based therapies for POI and will be explored in more detail in subsequent sections.

Table 2 Therapeutic mechanisms of mesenchymal stem cells in pre-

mature ovarian insufficiency	
Paracrine Signalling	MSCs secrete a wide array of growth factors (e.g., VEGF, HGF, IGF-1, FGF2), cytokines (e.g., IL-6, IL-10), and chemokines that can promote cell survival,
	proliferation, angiogenesis, and modulate immune responses within the ovarian microenvironment [44].
Immunomodulation	MSCs can suppress pro-inflammatory responses and promote an anti-inflammatory milieu, which is beneficial in cases of autoimmune oophoritis or chemotherapy-induced ovarian inflammation [42].
Anti-apoptotic Effects	MSCs can protect ovarian cells, particularly granulosa cells, from apoptosis induced by various stressors [45, 46].
Anti-fibrotic Effects	MSCs may reduce ovarian fibrosis, which can impair follicular development and ovarian function [47].
Mitochondrial Transfer	There is emerging evidence that MSCs can transfer healthy mitochondria to damaged cells, potentially restoring cellular function [48, 49].
Extracellular Vesicle (EV) Secretion	MSC-derived EVs, especially exosomes, carry proteins, lipids, mRNAs, and microRNAs that can mediate many of the paracrine effects of MSCs, offering a potential cell-free therapeutic approach [50–52].

# Very Small Embryonic-Like Stem Cells (VSELs) in Ovaries

Recent research has identified the presence of very small embryonic-like stem cells (VSELs) in adult mammalian ovaries, which may have significant implications for understanding and treating POI [53, 54]. VSELs are small (3–6  $\mu$ m) pluripotent stem cells that express markers of pluripotency such as OCT-4 A, SSEA-1, and NANOG.

Unlike the traditional understanding that female mammals are born with a fixed number of oocytes that cannot be replenished, evidence suggests that VSELs may represent a population of stem cells in the ovarian surface epithelium that can generate new oocytes throughout reproductive life [55, 56]. These cells remain relatively quiescent under normal conditions but can be activated in response to injury or disease.

Studies have demonstrated that VSELs survive chemotherapy in mouse ovaries while more mature follicular cells are destroyed [57]. This observation has important implications for fertility preservation and POI treatment, as it suggests that the ovary retains regenerative potential even after chemotherapy-induced damage. Stimulating the surviving VSELs could potentially restore ovarian function and fertility [58, 59].



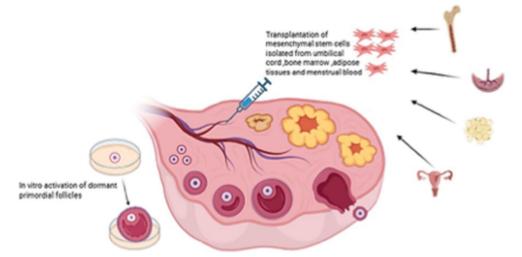
Furthermore, research indicates that VSELs may interact with ovarian somatic cells (particularly the ovarian surface epithelium) to initiate follicular development [37, 60]. This interaction appears to be regulated by various growth factors and cytokines, some of which are also secreted by MSCs. This suggests a potential synergistic relationship between endogenous VSELs and transplanted MSCs in ovarian regeneration [15, 43, 61].

The discovery of VSELs challenges the central dogma of fixed ovarian reserve and opens new avenues for POI treatment. Future research focusing on methods to activate endogenous VSELs, possibly in combination with MSC therapy, may lead to more effective regenerative approaches for women with POI [62, 63].

### Mesenchymal Stem Cell-Based Fertility Restoration in POI: Mechanisms and Evidence

The therapeutic potential of mesenchymal stem cells (MSCs) in restoring fertility for individuals with premature ovarian insufficiency (POI) represents one of the most dynamic and promising frontiers in regenerative medicine (Fig. 2) [13]. Conventional treatments for POI-related infertility, such as hormone replacement therapy (HRT) and assisted reproductive technologies (ART) with oocyte donation, address symptoms or bypass ovarian dysfunction but do not restore endogenous ovarian function [13, 64]. MSC-based therapies, in contrast, aim to rejuvenate the ovarian microenvironment, protect existing follicles, and potentially stimulate the activation of dormant primordial follicles, thereby offering a chance for natural conception or improved response to ART using autologous oocytes [13, 65].

Fig. 2 The therapeutic potential of mesenchymal stem cells isolated from different sources through their transplantation in dysfunctional ovaries due to POI in vivo and through activation of dormant primordial follicles in vitro in POI treatment. Adopted from [66]. Created in https://Bio-Render.com



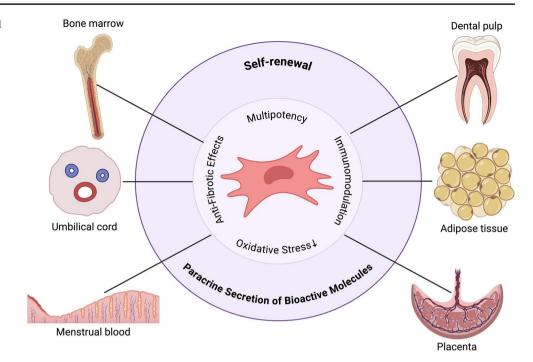
### **Sources of Mesenchymal Stem Cells for POI Therapy**

MSCs can be isolated from various adult and perinatal tissues, each with its own set of advantages and disadvantages for therapeutic application in POI (Fig. 3) [67]:

- Bone Marrow-Derived MSCs (BM-MSCs): Historically the most studied source, BM-MSCs have well-characterized regenerative properties. However, their isolation requires an invasive procedure, and their proliferation capacity and differentiation potential may decline with donor age [68].
- Adipose-Derived MSCs (AD-MSCs): AD-MSCs are abundant and can be obtained through a less invasive liposuction procedure. They exhibit robust proliferative capacity and potent immunomodulatory and angiogenic effects, making them an attractive option for POI [69].
- Umbilical Cord-Derived MSCs (UC-MSCs): These
  include MSCs from Wharton's jelly (WJ-MSCs) and
  umbilical cord blood. UC-MSCs are considered more
  primitive, possess higher proliferation rates, lower immunogenicity, and potent paracrine activity compared
  to adult MSCs. Their collection is non-invasive and
  ethically straightforward [70]. Several studies have
  highlighted the particular efficacy of UC-MSCs in POI
  models [71].
- Placenta-Derived MSCs (P-MSCs): The placenta is another rich source of young, highly proliferative MSCs with strong immunomodulatory properties. Similar to UC-MSCs, their collection is non-invasive [72].
- *Menstrual Blood-Derived MSCs (MenSCs)*: MenSCs can be easily and repeatedly collected non-invasively. They have shown promise in various regenerative applications, including endometrial regeneration and potentially ovarian rejuvenation, though research in POI is still emerging [73].



Fig. 3 Sources of Mesenchymal Stem Cells for POI Treatment. Adopted from [76]. Created in https://BioRender.com



Other Sources: MSCs have also been isolated from amniotic fluid, dental pulp, and induced pluripotent stem cells (iPSC-MSCs). iPSC-MSCs offer the potential for autologous therapy without invasive harvesting from adult tissues, but their generation and clinical translation face challenges related to safety and standardization [74, 75].

The choice of MSC source may influence therapeutic outcomes, and further research is needed to determine the optimal source for POI treatment.

### **Mechanisms of MSC-Mediated Ovarian Restoration**

MSCs exert their therapeutic effects in POI through a complex interplay of mechanisms, primarily driven by their paracrine activity rather than direct differentiation into ovarian cell types, which remains controversial and unlikely to be a major contributor to functional recovery [60, 77].

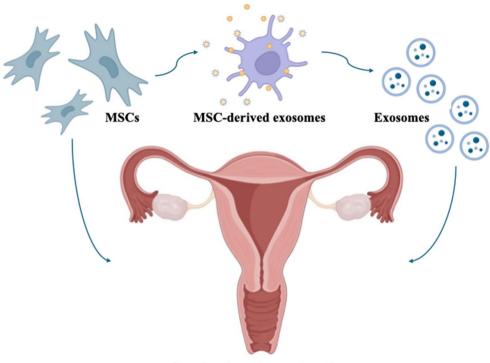
• Paracrine Secretion of Bioactive Molecules: MSCs release a plethora of growth factors (e.g., vascular endothelial growth factor (VEGF), hepatocyte growth factor (HGF), insulin-like growth factor 1 (IGF-1), basic fibroblast growth factor (bFGF)), cytokines (e.g., interleukin-10 (IL-10) and transforming growth factor-beta (TGF-β)), and chemokines [43, 44, 78, 79]. These factors collectively promote angiogenesis, where VEGF and bFGF stimulate the formation of new blood vessels, improving ovarian perfusion and nutrient supply, which is crucial for follicular survival and development [80].

They also inhibit apoptosis, where MSC-secreted factors can protect granulosa cells and oocytes from programmed cell death, a key feature of follicular atresia in POI [81], and stimulate cell proliferation and survival via growth factors like HGF and IGF-1, which can support the proliferation and survival of ovarian stromal and follicular cells [82, 83].

- *Immunomodulation*: MSCs possess potent immunomodulatory capabilities. They can suppress the activity of pro-inflammatory immune cells (e.g., Th1, Th17 lymphocytes, M1 macrophages) and promote an anti-inflammatory environment by inducing regulatory T cells (Tregs) and M2 macrophages. This is particularly relevant for autoimmune POI and for mitigating inflammation associated with chemotherapy-induced ovarian damage [42, 51, 72, 84].
- Anti-Fibrotic Effects: Chronic inflammation and tissue damage in POI can lead to ovarian fibrosis, impairing follicular development. MSCs can secrete anti-fibrotic factors and enzymes that degrade excess extracellular matrix, potentially reversing or limiting ovarian fibrosis [47, 85].
- Reduction of Oxidative Stress: Oxidative stress is a significant contributor to oocyte aging and follicular damage in POI. MSCs can enhance the antioxidant capacity of ovarian tissue by secreting antioxidant enzymes or by upregulating endogenous antioxidant pathways in ovarian cells [86, 87].
- Mitochondrial Transfer: Emerging evidence suggests that MSCs can transfer healthy mitochondria to damaged ovarian cells via tunneling nanotubes or extracellular



Fig. 4 MSCs stem cell-derived exosomes in POI treatment. Exosomes have the potential to markedly increase ovarian function and reproductive capacity in POI through stimulation of granulosa cells (GCs) inside the ovary. Adopted from [13] (created in https://BioRender.com)



### Ovarian function restoration

vesicles. This mitochondrial donation can rescue cells with mitochondrial dysfunction, improve cellular energy metabolism, and reduce apoptosis [48, 49].

Activation of Dormant Primordial Follicles: While the
exact mechanisms are still being elucidated, MSCs may
promote the activation of the remaining pool of dormant
primordial follicles, potentially through the secretion of
factors that influence the PI3K/AKT/mTOR pathway or
other signaling cascades involved in follicle awakening
[81, 88, 89].

### **Preclinical and Clinical Evidence**

Numerous preclinical studies using various animal models of POI (induced by chemotherapy, autoimmune mechanisms, or genetic factors) have demonstrated the efficacy of MSC transplantation from different sources [60, 81]. These studies consistently show improvements in ovarian morphology, increased numbers of healthy follicles at different developmental stages, restoration of hormone levels (e.g., increased estrogen and anti-Müllerian hormone (AMH), decreased FSH), reduced granulosa cell apoptosis, enhanced angiogenesis, and, in many cases, restoration of fertility with successful pregnancies [13, 80]. For instance, a recent meta-analysis of preclinical studies confirmed significant improvements in ovarian function markers following MSC therapy [15, 90].

Clinical translation of MSC therapy for POI is still in its early stages, but initial results from pilot studies and small clinical trials are encouraging. These studies have generally reported good safety and tolerability of MSC administration (often via intraovarian injection or systemic infusion) [66, 91, 92]. Reported outcomes in some patients include improvements in hormonal profiles, increased antral follicle counts, resumption of menstruation, and even spontaneous pregnancies [93, 94]. However, these trials are often limited by small sample sizes, lack of control groups, variability in MSC sources, dosage, and delivery methods, and short follow-up periods. Larger, well-controlled randomized clinical trials are crucial to definitively establish the efficacy and safety of MSC therapy for POI in humans [95, 96].

# The Emerging Role of MSC-Derived Extracellular Vesicles (Exosomes) in POI Therapy

In recent years, the therapeutic potential of mesenchymal stem cell (MSC)-derived extracellular vesicles (EVs), particularly exosomes, has garnered significant attention as a novel cell-free approach for treating various diseases, including premature ovarian insufficiency (POI) (Fig. 4) [97–99]. Exosomes are nano-sized (typically 30–150 nm) membrane-bound vesicles secreted by most cell types, including MSCs [98, 100]. They act as intercellular messengers, transferring a diverse cargo of bioactive molecules—such as proteins, lipids, mRNAs, and microRNAs (miRNAs)—from their parent cells to recipient cells, thereby modulating the function of the recipient cells [101, 102].



# Exosome-Based Therapy Over Whole-Cell MSC Therapy

Using MSC-derived exosomes instead of whole MSCs offers several potential advantages, including an improved safety profile, knowing that exosomes are non-living and cannot replicate, eliminating risks associated with wholecell therapy, such as uncontrolled proliferation, differentiation into undesirable cell types, or tumorigenicity (though the risk with MSCs themselves is very low) [103, 104]. Exosomes generally exhibit lower immunogenicity compared to their parent cells, potentially allowing for repeated administrations without eliciting strong immune responses [105, 106]. Exosomes can also be more easily stored and transported than live cells, facilitating off-the-shelf therapeutic products [107, 108]. Furthermore, characterizing and standardizing exosome preparations may be more straightforward than for complex cellular products [109]. Due to their small size, exosomes may be better able to penetrate tissues and cross biological barriers [110].

#### Mechanisms of Action of MSC-Exosomes in POI

MSC-derived exosomes are believed to mediate many of the regenerative effects previously attributed to MSCs themselves [98]. In the context of POI, MSC-exosomes have been shown in preclinical studies to promote granulosa cell proliferation and inhibit apoptosis, where exosomal cargo, particularly specific miRNAs and growth factors, can protect granulosa cells from damage and support their function [111]. Exosomes can also deliver pro-angiogenic factors (e.g., VEGF) or miRNAs that promote neovascularization in the ovary, improving blood supply and follicular health [112]. MSC-exosomes can carry immunomodulatory molecules that suppress inflammation and promote a tolerogenic microenvironment in the ovary [113, 114]. Exosomes may transfer antioxidant enzymes or molecules that enhance the antioxidant capacity of ovarian cells [115]. On the other hand, specific miRNAs within exosomes (e.g., miR-17-5p, miR-146a, miR-21) have been implicated in regulating follicular growth, atresia, and steroidogenesis. For example, studies have shown that MSC-exosomes can upregulate AMH expression and promote the transition from primordial to primary follicles [116–118]. By delivering a cocktail of beneficial molecules, exosomes can help restore homeostasis to the damaged ovarian microenvironment [119, 120].

### **Preclinical Evidence for MSC-Exosomes in POI**

A growing body of preclinical research supports the therapeutic potential of MSC-exosomes in POI [121, 122]. Studies using animal models of chemotherapy-induced or

age-related ovarian dysfunction have demonstrated that administration of MSC-exosomes can lead to (1) restoration of ovarian function, including regular estrous cycles and improved hormone levels (e.g., increased estrogen and AMH, decreased FSH) [123] (2) increased number of healthy follicles and reduced follicular atresia [124]; (3) enhanced ovarian angiogenesis and reduced ovarian fibrosis [141]; and (4) improved oocyte quality and even successful pregnancies after exosome treatment in some models [124].

### Challenges and Future Directions for Exosome Therapy in POI

Despite the promising preclinical data, several challenges need to be addressed before MSC-exosome therapy can become a clinical reality for POI, including (1) developing standardized and scalable methods for isolating and characterizing exosomes is crucial for ensuring product consistency and quality [125], (2) identifying the specific therapeutic components within the exosomal cargo and developing assays to measure their potency are ongoing research areas [126], (3) determining the optimal dose, delivery route (e.g., systemic vs. intraovarian injection), and timing of exosome administration requires further investigation [127], and establishing cost-effective methods for large-scale production of clinical-grade exosomes is necessary for widespread application [128]. Moreover, rigorous, well-designed clinical trials are needed to evaluate the safety and efficacy of MSC-exosome therapy in women with POI [71]. Nevertheless, MSC-derived exosomes represent a highly promising next-generation, cell-free therapeutic strategy for POI, potentially offering a safer and more practical alternative to whole-cell therapies.

## Challenges, Future Directions, and Conclusion

### Challenges in Translating MSC-Based Therapies for POI To the Clinic

Despite the promising preclinical data and early clinical observations, several challenges must be overcome to successfully translate MSC-based therapies (including cell-based and cell-free approaches like exosomes) into routine clinical practice for POI [129]. Standardization of MSC manufacturing remains a significant hurdle, with considerable variability in isolation protocols, culture conditions (e.g., use of fetal bovine serum vs. serum-free media, 2D vs. 3D culture), characterization methods, and expansion procedures, necessitating the establishment of standardized, Good Manufacturing Practice (GMP)-compliant



protocols to ensure product consistency, safety, and efficacy [130–133]. The optimal cell source (bone marrow, adipose, umbilical cord, etc.), dosage, delivery route (e.g., systemic intravenous infusion, direct intraovarian injection via laparoscopy or ultrasound guidance), and timing and frequency of treatment are yet to be definitively established, with intraovarian injection offering targeted delivery but being more invasive than systemic routes, highlighting the need for comparative studies to address these critical parameters [134-136]. Long-term data on the durability of therapeutic effects and potential late adverse events (e.g., immunogenicity with repeated doses, ectopic tissue formation, though rare for MSCs) are crucial, as most clinical studies to date have relatively short follow-up periods [125, 129]. Identifying the subset of POI patients most likely to benefit from MSC therapy is important [137], with early-onset POI (before age 40) and POI with preserved ovarian tissue potentially more responsive to treatment; factors such as age, etiology of POI, and remaining ovarian reserve may influence outcomes, while standardized, clinically meaningful efficacy measures (e.g., sustained restoration of menses, hormonal balance, antral follicle count, oocyte quality, live birth rates) need to be consistently applied across trials [138-140]. A more precise understanding of the specific molecular mediators (e.g., key growth factors, cytokines, exosomal miRNAs) and signaling pathways involved in MSC-mediated ovarian repair is needed to optimize therapies and develop targeted interventions, despite the wide acceptance of paracrine effects [43, 141]. Finally, evaluating the cost-effectiveness of these potentially expensive treatments compared to existing options will be important for their broader adoption [142, 143].

### **Future Directions in MSC Research for POI**

The field of MSC research for POI is rapidly evolving, with several exciting future directions. Advanced MSC engineering, including genetic modification to overexpress specific therapeutic factors (e.g., anti-inflammatory cytokines, pro-angiogenic factors) or to enhance ovarian homing, could improve efficacy, while preconditioning MSCs under hypoxic conditions may boost their paracrine activity [144, 145]. Cell-free therapies using MSC-derived exosomes or other EVs hold immense promise due to their potential safety, stability, and manufacturing advantages, with research focusing on optimizing exosome production, characterizing therapeutic cargo, and conducting clinical trials [145]. Combination therapies that merge MSC treatment with antioxidants, growth factors, or existing ovarian stimulation protocols may yield synergistic effects [146]. The use of biocompatible scaffolds to deliver MSCs or their products directly to the ovary could enhance retention, survival,

and local therapeutic effects, potentially creating a regenerative ovarian niche [147–149]. Personalized medicine approaches that tailor MSC therapies based on the specific etiology of POI in individual patients or their genetic background may improve outcomes [150]. While not a direct MSC therapy, advances in creating ovarian organoids from iPSCs or other cell sources, potentially supported by MSCs or their secretome, could offer future avenues for understanding ovarian biology and developing novel fertility restoration techniques [151, 152]. Finally, developing noninvasive methods to track MSCs post-transplantation and to monitor ovarian responses to therapy will be valuable for optimizing treatment protocols [153].

### **Conclusion**

Premature ovarian insufficiency is a challenging condition with profound implications for female reproductive and overall health [3, 20]. Current management strategies are largely supportive, failing to address the underlying loss of ovarian function [20]. Mesenchymal stem cell therapy, leveraging the unique regenerative and immunomodulatory properties of MSCs, has emerged as a highly promising therapeutic avenue [15, 42, 84]. A wealth of preclinical evidence demonstrates the ability of MSCs, and increasingly their derived exosomes, to improve ovarian function, enhance folliculogenesis, and restore fertility in various POI models [109, 145]. These effects are mediated through complex paracrine mechanisms, including the secretion of growth factors, cytokines, and miRNAs that collectively reduce inflammation and apoptosis, promote angiogenesis, modulate the immune system, and improve the ovarian microenvironment [43, 78, 122].

Early clinical trials have provided encouraging, albeit preliminary, evidence of the safety and potential efficacy of MSC therapy in women with POI [27, 121]. However, the field is still in its nascent stages, and significant challenges related to standardization, optimal treatment protocols, long-term outcomes, and regulatory approval must be addressed.

Future research focused on elucidating the precise mechanisms of action, optimizing MSC sources and delivery methods, exploring the potential of engineered MSCs and cell-free exosome-based therapies, and conducting rigorous, large-scale clinical trials will be critical for translating this promising therapeutic modality into a widely available and effective treatment for POI. The continued exploration of MSCs and their derivatives offers tangible hope for restoring ovarian function and improving the quality of life for the many women affected by this debilitating condition, moving



beyond symptomatic relief towards true ovarian rejuvenation and fertility restoration.

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Consent for Publication Not applicable.

Consent To Participate Not applicable.

**Competing Interests** The authors declare no competing interests.

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Clinical Trial Number Not applicable.

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### References

- Mason, J. B., Habermehl, T. L., Underwood, K. B., Schneider, A., Brieño-Enriquez, M. A., Masternak, M. M., & Parkinson, K. C. (2022). The interrelationship between female reproductive aging and survival. *The Journals of Gerontology, Series A: Biological Sciences and Medical Sciences*, 77(1), 75–83. https://doi.org/10.1 093/gerona/glab252
- Tucker, E. J., Baker, M. J., Hock, D. H., Warren, J. T., Jaillard, S., Bell, K. M., & Sinclair, A. H. (2022). Premature ovarian

- insufficiency in CLPB deficiency: Transcriptomic, proteomic and phenotypic insights. *The Journal of Clinical Endocrinology & Metabolism*, 107(12), 3328–3340. https://doi.org/10.1210/clinem/dgac528
- 3. Kapoor, E. (2023). Premature ovarian insufficiency. *Current Opinion in Endocrine and Metabolic Research*, 28, 100435. https://doi.org/10.1016/j.coemr.2023.100435
- Ulin, M., Cetin, E., Hobeika, E., Chugh, R. M., Park, H. S., Esfandyari, S., & Al-Hendy, A. (2021). Human mesenchymal stem cell therapy and other novel treatment approaches for premature ovarian insufficiency. *Reproductive Sciences*, 28(6), 1688–1696. https://doi.org/10.1007/s43032-021-00528-z
- Panay, N., Anderson, R. A., Bennie, A., Cedars, M., Davies, M., Ee, C.,... Flanagan, M. (2024). Evidence-based guideline: premature ovarian insufficiency, Human Reproduction Open, 2024(4). https://doi.org/10.1093/hropen/hoae065.
- Panay, N., Anderson, R. A., Nappi, R. E., Vincent, A. J., Vujovic, S., Webber, L., & Wolfman, W. (2020). Premature ovarian insufficiency: An International Menopause Society white paper. Climacteric: The Journal of the International Menopause Society, 23(5), 426–446. https://doi.org/10.1080/13697137.2020.1804 547
- Jarrige, M., Frank, E., Herardot, E., Martineau, S., Darle, A., Benabides, M., & Ben M'Barek, K. (2021). The future of regenerative medicine: Cell therapy using pluripotent stem cells and acellular therapies based on extracellular vesicles. *Cells*, 10(2), 240. https://doi.org/10.3390/cells10020240
- Bacakova, L., Zarubova, J., Travnickova, M., Musilkova, J., Pajorova, J., Slepicka, P., & Molitor, M. (2018). Stem cells: Their source, potency and use in regenerative therapies with focus on adipose-derived stem cells—a review. *Biotechnology Advances*, 36(4), 1111–1126. https://doi.org/10.1016/j.biotechadv.2018.03.0
- Zakrzewski, W., Dobrzyński, M., Szymonowicz, M., & Rybak, Z. (2019). Stem cells: Past, present, and future. Stem Cell Research & Therapy, 10(1), 68. https://doi.org/10.1186/s13287-019-1165
- Tan, S. S. H., Tjio, C. K. E., Wong, J. R. Y., Wong, K. L., Chew, J. R. J., Hui, J. H. P., & Toh, W. S. (2021). Mesenchymal stem cell exosomes for cartilage regeneration: A systematic review of preclinical in vivo studies. *Tissue Engineering, Part B, Reviews*, 27(1), 1–13. https://doi.org/10.1089/ten.teb.2019.0326
- Ullah, I., Subbarao, R. B., & Rho, G. J. (2015). Human mesenchymal stem cells - Current trends and future prospective. *Biosci*ence Reports. https://doi.org/10.1042/BSR20150025
- Li, J., Yu, Q., Huang, H., Deng, W., Cao, X., Adu-Frimpong, M., & Xu, X. (2018). Human chorionic plate-derived mesenchymal stem cells transplantation restores ovarian function in a chemotherapy-induced mouse model of premature ovarian failure. *Stem Cell Research & Therapy*, 9(1), 81. https://doi.org/10.1186/s13287-018-0819-7
- Ali, I., Padhiar, A. A., Wang, T., He, L., Chen, M., Wu, S., & Zhou, G. (2022). Stem cell-based therapeutic strategies for premature ovarian insufficiency and infertility: A focus on aging. Cells, 11(23), 3713. https://doi.org/10.3390/cells11233713
- Han, Y., Li, X., Zhang, Y., Han, Y., Chang, F., & Ding, J. (2019).
   Mesenchymal stem cells for regenerative medicine. *Cells*,8(8),
   Article 886. https://doi.org/10.3390/cells8080886
- Gao, G., Li, L., Li, C., Liu, D., Wang, Y., & Li, C. (2024). Mesenchymal stem cells: Guardians of women's health. *Regenerative Therapy*, 26, 1087–1098. https://doi.org/10.1016/j.reth.2024.10.0
- Song, N., Scholtemeijer, M., & Shah, K. (2020). Mesenchymal stem cell immunomodulation: Mechanisms and therapeutic potential. *Trends in Pharmacological Sciences*, 41(9), 653–664. https://doi.org/10.1016/j.tips.2020.06.009



- Li, J., Fan, H., Liu, W., Zhang, J., Xiao, Y., Peng, Y.,... Li, J. (2024). Mesenchymal stem cells promote ovarian reconstruction in mice. Stem Cell Research & Therapy, 15(1), 115. https://doi.org/10.1186/s13287-024-03718-z.
- Fortuño, C., & Labarta, E. (2014). Genetics of primary ovarian insufficiency: A review. *Journal of Assisted Reproduction and Genetics*, 31(12), 1573–1585. https://doi.org/10.1007/s10815-01 4-0342-9
- Orisaka, M., Mizutani, T., Miyazaki, Y., Shirafuji, A., Tamamura, C., Fujita, M., & Yoshida, Y. (2023). Chronic low-grade inflammation and ovarian dysfunction in women with polycystic ovarian syndrome, endometriosis, and aging. Frontiers in Endocrinology. https://doi.org/10.3389/fendo.2023.1324429
- Benetti-Pinto, C. L., Soares Júnior, J. M., Maciel, G. A., Nácul, A. P., Yela, D. A., & Silva, A. C. J. S. R. e. (2020). Premature ovarian insufficiency: A hormonal treatment approach. Revista Brasileira de Ginecologia e Obstetrícia / RBGO Gynecology and Obstetrics, 42(08), 511–518. https://doi.org/10.1055/s-0040-1716929
- Jiao, X., Meng, T., Zhai, Y., Zhao, L., Luo, W., Liu, P., & Qin, Y. (2021). Ovarian reserve markers in premature ovarian insufficiency: Within different clinical stages and different etiologies. Frontiers in Endocrinology. https://doi.org/10.3389/fendo.2021. 601752
- Moshfegh, F., Balanejad, S. Z., Shahrokhabady, K., & Attaranzadeh, A. (2022). Crocus sativus (saffron) petals extract and its active ingredient, anthocyanin improves ovarian dysfunction, regulation of inflammatory genes and antioxidant factors in testosterone-induced PCOS mice. Journal of Ethnopharmacology, 282, 114594. https://doi.org/10.1016/j.jep.2021.114594
- Park, S. U., Walsh, L., & Berkowitz, K. M. (2021). Mechanisms of ovarian aging. *Reproduction*, 162(2), R19–R33. https://doi.org/10.1530/REP-21-0022
- 24. Nie, L., Wang, X., Wang, S., Hong, Z., & Wang, M. (2024). Genetic insights into the complexity of premature ovarian insufficiency. *Reproductive Biology and Endocrinology*, 22(1), 94. htt ps://doi.org/10.1186/s12958-024-01254-2
- Wesevich, V., Kellen, A. N., & Pal, L. (2020). Recent advances in understanding primary ovarian insufficiency. F1000Research, 9, 1101. https://doi.org/10.12688/f1000research.26423.1
- Torrealday, S., Kodaman, P., & Pal, L. (2017). Premature ovarian insufficiency - an update on recent advances in understanding and management. F1000research, 6, 2069. https://doi.org/10.12688/f1 000research.11948.1
- Federici, S., Rossetti, R., Moleri, S., Munari, E. V., Frixou, M., Bonomi, M., & Persani, L. (2024). Primary ovarian insufficiency: Update on clinical and genetic findings. Frontiers in Endocrinology. https://doi.org/10.3389/fendo.2024.1464803
- Jinno, M. (2025). Ovarian stimulation by promoting basal follicular growth. *Reproductive Biology and Endocrinology*, 23(1), 35. https://doi.org/10.1186/s12958-025-01356-5
- Zhang, T., He, M., Zhang, J., Tong, Y., Chen, T., Wang, C.,... Xiao, Z. (2023). Mechanisms of primordial follicle activation and new pregnancy opportunity for premature ovarian failure patients. Frontiers in Physiology, 14. https://doi.org/10.3389/fphy s.2023.1113684.
- Chon, S. J., Umair, Z., & Yoon, M. S. (2021). Premature ovarian insufficiency: Past, present, and future. Frontiers in Cell and Developmental Biology. https://doi.org/10.3389/fcell.2021.6728
- Ibarra-Ramírez, M., Campos-Acevedo, L. D., & de Martínez, L. E. (2023). Chromosomal abnormalities of interest in Turner syndrome: An update. *Journal of Pediatric Genetics*, 12(04), 263–272. https://doi.org/10.1055/s-0043-1770982
- 32. Barasoain, M., Barrenetxea, G., Huerta, I., Télez, M., Criado, B., & Arrieta, I. (2016). Study of the genetic etiology of primary

- ovarian insufficiency: FMR1 gene. *Genes*, 7(12), 123. https://doi.org/10.3390/genes7120123
- Qin, Y., Jiao, X., Simpson, J. L., & Chen, Z. J. (2015). Genetics of primary ovarian insufficiency: New developments and opportunities. *Human Reproduction Update*, 21(6), 787–808. https://doi.or g/10.1093/humupd/dmv036
- Szeliga, A., Calik-Ksepka, A., Maciejewska-Jeske, M., Grymowicz, M., Smolarczyk, K., Kostrzak, A.,... Meczekalski, B. (2021).
   Autoimmune Diseases in Patients with Premature Ovarian Insufficiency—Our Current State of Knowledge. International Journal of Molecular Sciences, 22(5), 2594. https://doi.org/10.3390/ijms 22052594.
- Kirshenbaum, M., & Orvieto, R. (2019). Premature ovarian insufficiency (POI) and autoimmunity-an update appraisal. *Journal of Assisted Reproduction and Genetics*, 36(11), 2207–2215. https://doi.org/10.1007/s10815-019-01572-0
- Kim, S., Kim, S. W., Han, S. J., Lee, S., Park, H. T., Song, J. Y., & Kim, T. (2021). Molecular mechanism and prevention strategy of chemotherapy- and radiotherapy-induced ovarian damage. *International Journal of Molecular Sciences*, 22(14), 7484. https://doi.org/10.3390/ijms22147484
- Spears, N., Lopes, F., Stefansdottir, A., Rossi, V., De Felici, M., Anderson, R. A., & Klinger, F. G. (2019). Ovarian damage from chemotherapy and current approaches to its protection. *Human Reproduction Update*, 25(6), 673–693. https://doi.org/10.1093/humupd/dmz027
- Bedoschi, G., Navarro, P. A., & Oktay, K. (2016). Chemotherapy-induced damage to ovary: Mechanisms and clinical impact. Future Oncology, 12(20), 2333–2344. https://doi.org/10.2217/fo n-2016-0176
- Neff, A. M., Laws, M. J., Warner, G. R., & Flaws, J. A. (2022). The effects of environmental contaminant exposure on reproductive aging and the menopause transition. *Current Environmental Health Reports*, 9(1), 53–79. https://doi.org/10.1007/s40572-022-00334-v
- Hernández-Angeles, C., & Castelo-Branco, C. (2016). Early menopause. *Indian Journal of Medical Research*, 143(4), 420– 427. https://doi.org/10.4103/0971-5916.184283
- 41. Adam, M. P., Feldman, J., & Mirzaa, G. M. (2000). Classic Galactosemia and Clinical Variant Galactosemia.
- Shi, Y., Wang, Y., Li, Q., Liu, K., Hou, J., Shao, C., & Wang, Y. (2018). Immunoregulatory mechanisms of mesenchymal stem and stromal cells in inflammatory diseases. *Nature Reviews Nephrology*, 14(8), 493–507. https://doi.org/10.1038/s41581-018-0023-5
- Han, Y., Yang, J., Fang, J., Zhou, Y., Candi, E., Wang, J., & Shi, Y. (2022). The secretion profile of mesenchymal stem cells and potential applications in treating human diseases. *Signal Transduction and Targeted Therapy*, 7(1), 92. https://doi.org/10.1038/s41392-022-00932-0
- Maacha, S., Sidahmed, H., Jacob, S., Gentilcore, G., Calzone, R., Grivel, J.-C., & Cugno, C. (2020). Paracrine mechanisms of mesenchymal stromal cells in angiogenesis. *Stem Cells International*, 2020, 1–12. https://doi.org/10.1155/2020/4356359
- Chen, Z., Xia, X., Yao, M., Yang, Y., Ao, X., Zhang, Z.,... Xu, X. (2024). The dual role of mesenchymal stem cells in apoptosis regulation. Cell Death & Disease, 15(4), 250. https://doi.org/10.1038/s41419-024-06620-x.
- Elsherbiny, N. M., Abdel-Maksoud, M. S., Prabahar, K., Mohammedsaleh, Z. M., Badr, O. A. M., Dessouky, A. A., & Ebrahim, N. (2024). MSCs-derived EVs protect against chemotherapyinduced ovarian toxicity: Role of PI3K/AKT/mTOR axis. *Journal of Ovarian Research*, 17(1), 222. https://doi.org/10.1186/s13048-024-01545-7



- 47. Amargant, F., Vieira, C., Pritchard, M. T., & Duncan, F. E. (2024). Systemic low-dose anti-fibrotic treatment attenuates ovarian aging in the mouse. https://doi.org/10.1101/2024.06.21.600035
- Liu, Q., Zhang, X., Zhu, T., Xu, Z., Dong, Y., & Chen, B. (2024). Mitochondrial transfer from mesenchymal stem cells: Mechanisms and functions. *Mitochondrion*, 79, 101950. https://doi.org/10.1016/j.mito.2024.101950
- Li, C., Cheung, M. K. H., Han, S., Zhang, Z., Chen, L., Chen, J., & Qiu, J. (2019). Mesenchymal stem cells and their mitochondrial transfer: A double-edged sword. *Bioscience Reports*. https:// doi.org/10.1042/BSR20182417
- Yudintceva, N., Mikhailova, N., Fedorov, V., Samochernych, K., Vinogradova, T., Muraviov, A., & Shevtsov, M. (2022). Mesenchymal stem cells and MSCs-derived extracellular vesicles in infectious diseases: From basic research to clinical practice. *Bioengineering*, 9(11), 662. https://doi.org/10.3390/bioengineering9110662
- van Griensven, M., & Balmayor, E. R. (2024). Extracellular vesicles are key players in mesenchymal stem cells' dual potential to regenerate and modulate the immune system. *Advanced Drug Delivery Reviews*, 207, 115203. https://doi.org/10.1016/j.addr.2024.115203
- Park, H. S., Cetin, E., Siblini, H., Seok, J., Alkelani, H., Alkhrait, S., & Al-Hendy, A. (2023). Therapeutic potential of mesenchymal stem cell-derived extracellular vesicles to treat PCOS. *Interna*tional Journal of Molecular Sciences, 24(13), 11151. https://doi.org/10.3390/ijms241311151
- Bhartiya, D., Jha, N., Tripathi, A., & Tripathi, A. (2023). Very small embryonic-like stem cells have the potential to win the three-front war on tissue damage, cancer, and aging. Frontiers in Cell and Developmental Biology, 10. https://doi.org/10.3389/fcel 1.2022.1061022
- Bhartiya, D., Patel, H., & Parte, S. (2018). Improved understanding of very small embryonic-like stem cells in adult mammalian ovary. *Human Reproduction*, 33(5), 978–979. https://doi.org/10.1093/humrep/dev039
- Hanna, C. B., & Hennebold, J. D. (2014). Ovarian germline stem cells: An unlimited source of oocytes? Fertility and Sterility, 101(1), 20–30. https://doi.org/10.1016/j.fertnstert.2013.11.009
- Parte, S., Patel, H., Sriraman, K., & Bhartiya, D. (2015). Isolation and Characterization of Stem Cells in the Adult Mammalian Ovary (pp. 203–229). https://doi.org/10.1007/978-1-4939-1785-3 16
- Sriraman, K., Bhartiya, D., Anand, S., & Bhutda, S. (2015).
   Mouse ovarian very small embryonic-like stem cells resist chemotherapy and retain ability to initiate oocyte-specific differentiation. *Reproductive Sciences*, 22(7), 884–903. https://doi.org/10.1177/1933719115576727
- Sharma, D., & Bhartiya, D. (2021). Stem cells in adult mice ovaries form germ cell nests, undergo meiosis, neo-oogenesis and follicle assembly on regular basis during estrus cycle. Stem Cell Reviews and Reports, 17(5), 1695–1711. https://doi.org/10.1007/s12015-021-10237-4
- Sharma, D., & Bhartiya, D. (2022). Dysfunctional ovarian stem cells due to neonatal endocrine disruption result in PCOS and ovarian insufficiency in adult mice. *Stem Cell Reviews and Reports*, 18(8), 2912–2927. https://doi.org/10.1007/s12015-022-10414-z
- Li, Z., Zhang, M., Tian, Y., Li, Q., & Huang, X. (2021). Mesenchymal stem cells in premature ovarian insufficiency: Mechanisms and prospects. *Frontiers in Cell and Developmental Biology*. https://doi.org/10.3389/fcell.2021.718192
- Shan, Y., Zhang, M., Tao, E., Wang, J., Wei, N., Lu, Y.,... Wang,
   G. (2024). Pharmacokinetic characteristics of mesenchymal stem cells in translational challenges. Signal Transduction and

- Targeted Therapy, 9(1), 242. https://doi.org/10.1038/s41392-02 4-01936-8.
- Cui, X., & Jing, X. (2024). Stem cell-based therapeutic potential in female ovarian aging and infertility. *Journal of Ovarian Research*, 17(1), 171. https://doi.org/10.1186/s13048-024-0149 2-3
- Dunlop, C. E., Telfer, E. E., & Anderson, R. A. (2013). Ovarian stem cells—Potential roles in infertility treatment and fertility preservation. *Maturitas*, 76(3), 279–283. https://doi.org/10.1016/j.maturitas.2013.04.017
- 64. Makridakis, M., Roubelakis, M. G., & Vlahou, A. (2013). Stem cells: Insights into the secretome. *Biochimica Et Biophysica Acta Proteins And Proteomics*, 1834(11), 2380–2384. https://doi.org/10.1016/j.bbapap.2013.01.032
- Zafardoust, S., Kazemnejad, S., Fathi-Kazerooni, M., Darzi, M., Sadeghi, M. R., Tabar, S., & Sehat, Z. (2023). The effects of intraovarian injection of autologous menstrual blood-derived mesenchymal stromal cells on pregnancy outcomes in women with poor ovarian response. *Stem Cell Research & Therapy*, 14(1), 332. http s://doi.org/10.1186/s13287-023-03568-1
- 66. Umer, A., Khan, N., Greene, D. L., Habiba, U. E., Shamim, S., & Khayam, A. U. (2023). The therapeutic potential of human umbilical cord derived mesenchymal stem cells for the treatment of premature ovarian failure. Stem Cell Reviews and Reports, 19(3), 651–666. https://doi.org/10.1007/s12015-022-10493-y
- Semenova, E., P Grudniak, M., K Machaj, E., Bocian, K., Chroscinska-Krawczyk, M., Trochonowicz, M., & Rozwadowska, N. (2021). Mesenchymal stromal cells from different parts of umbilical cord: Approach to comparison & characteristics. *Stem Cell Reviews and Reports*, 17(5), 1780–1795. https://doi.org/10.1007/s12015-021-10157-3
- Arnhold, S. J., Goletz, I., Klein, H., Stumpf, G., Beluche, L. A., Rohde, C.,... Litzke, L. F. (2007). Isolation and characterization of bone marrow–derived equine mesenchymal stem cells. American Journal of Veterinary Research, 68(10), 1095–1105. https://d oi.org/10.2460/ajvr.68.10.1095.
- Mikłosz, A., Nikitiuk, B. E., & Chabowski, A. (2022). Using adipose-derived mesenchymal stem cells to fight the metabolic complications of obesity: Where do we stand? *Obesity Reviews*. h ttps://doi.org/10.1111/obr.13413
- Chen, M. Y., Lie, P. C., Li, Z. L., & Wei, X. (2009). Endothelial differentiation of wharton's jelly-derived mesenchymal stem cells in comparison with bone marrow-derived mesenchymal stem cells. *Experimental Hematology*, 37(5), 629–640. https://doi.org/10.1016/j.exphem.2009.02.003
- Elahi, N., Ai, J., & Makoolati, Z. (2023). A review on treatment of premature ovarian insufficiency: Characteristics, limitations, and challenges of stem cell versus exosometherapy. *Veterinary Medicine International*. https://doi.org/10.1155/2023/5760011
- Talwadekar, M. D., Kale, V. P., & Limaye, L. S. (2015). Placentaderived mesenchymal stem cells possess better immunoregulatory properties compared to their cord-derived counterparts–A paired sample study. *Scientific Reports*, 5(1), 15784. https://doi.or g/10.1038/srep15784
- Fu, X., Zhang, S., Li, T., Zhang, R., Lu, Y., Cheng, H., & Lin, J. (2022). Menstrual blood-derived endometrial stem cells ameliorate the viability of ovarian granulosa cells injured by cisplatin through activating autophagy. *Reproductive Toxicology*, 110, 39–48. https://doi.org/10.1016/j.reprotox.2022.03.012
- Jovic, D., Yu, Y., Wang, D., Wang, K., Li, H., Xu, F., & Luo, Y. (2022). A brief overview of global trends in MSC-based cell therapy. *Stem Cell Reviews and Reports*, 18(5), 1525–1545. https://doi.org/10.1007/s12015-022-10369-1
- 75. Thanaskody, K., Jusop, A. S., Tye, G. J., Kamarul Zaman, W., Dass, W. S., S. A., & Nordin, F. (2022). MSCs vs. iPSCs: Potential



- in therapeutic applications. Frontiers in Cell and Developmental Biology, 10. https://doi.org/10.3389/fcell.2022.1005926
- Brown, M. G., Brady, D. J., Healy, K. M., Henry, K. A., Ogunsola, A. S., & Ma, X. (2024). Stem cells and acellular preparations in bone regeneration/fracture healing: Current therapies and future directions. *Cells*, 13(12), 1045. https://doi.org/10.3390/cells13121045
- Hu, H. Q., Xin, X. Y., Zhu, Y. T., Fan, R. W., Zhang, H. L., Ye, Y., & Li, D. (2024). Application of mesenchymal stem cell therapy for premature ovarian insufficiency: Recent advances from mechanisms to therapeutics. World Journal of Stem Cells, 16(1), 1–6. https://doi.org/10.4252/wjsc.v16.i1.1
- González-González, A., García-Sánchez, D., Dotta, M., Rodríguez-Rey, J. C., & Pérez-Campo, F. M. (2020). Mesenchymal stem cells secretome: The cornerstone of cell-free regenerative medicine. World Journal of Stem Cells, 12(12), 1529–1552. https://doi.org/10.4252/wjsc.v12.i12.1529
- Park, S. R., Kim, J. W., Jun, H. S., Roh, J. Y., Lee, H. Y., & Hong, I. S. (2018). Stem cell secretome and its effect on cellular mechanisms relevant to wound healing. *Molecular Therapy*, 26(2), 606–617. https://doi.org/10.1016/j.ymthe.2017.09.023
- Cacciottola, L., Vitale, F., Donnez, J., & Dolmans, M. M. (2023).
   Use of mesenchymal stem cells to enhance or restore fertility potential: A systematic review of available experimental strategies. *Human Reproduction Open*. https://doi.org/10.1093/hropen/hoad040
- Sadeghi, S., Mosaffa, N., Huang, B., & Ramezani Tehrani, F. (2024). Protective role of stem cells in POI: Current status and mechanism of action, a review article. *Heliyon*, 10(1), e23271. htt ps://doi.org/10.1016/j.heliyon.2023.e23271
- Francés-Herrero, E., Bueno-Fernandez, C., Rodríguez-Eguren, A., Gómez-Álvarez, M., Faus, A., Soto-Prado, A., & Cervelló, I. (2024). Growth factor-loaded ovarian extracellular matrix hydrogels promote in vivo ovarian niche regeneration and enhance fertility in premature ovarian insufficiency preclinical models. *Acta Biomaterialia*, 186, 125–140. https://doi.org/10.1016/j.actbio.2024.07.056
- Youssef, A., Aboalola, D., & Han, V. K. M. (2017). The Roles of Insulin-Like Growth Factors in Mesenchymal Stem Cell Niche. Stem Cells International, 2017, 1–12. https://doi.org/10.1155/201 7/9453108
- Schu, S., Nosov, M., O'Flynn, L., Shaw, G., Treacy, O., Barry, F., & Ritter, T. (2012). Immunogenicity of allogeneic mesenchymal stem cells. *Journal of Cellular and Molecular Medicine*, 16(9), 2094–2103. https://doi.org/10.1111/j.1582-4934.2011.01509.x
- Gu, M., Wang, Y., & Yu, Y. (2024). Ovarian fibrosis: Molecular mechanisms and potential therapeutic targets. *Journal of Ovarian Research*, 17(1), 139. https://doi.org/10.1186/s13048-024-01448 -7
- Ra, K., Park, S. C., & Lee, B. C. (2023). Female reproductive aging and oxidative stress: Mesenchymal stem cell conditioned medium as a promising antioxidant. *International Journal of Molecular Sciences*, 24(5), 5053. https://doi.org/10.3390/ijms24 055053
- Yan, F., Zhao, Q., Li, Y., Zheng, Z., Kong, X., Shu, C.,... Shi, Y. (2022). The role of oxidative stress in ovarian aging: a review. Journal of Ovarian Research, 15(1), 100. https://doi.org/10.1186/s13048-022-01032-x.
- Bai, X., & Wang, S. (2022). Signaling pathway intervention in premature ovarian failure. Frontiers in Medicine. https://doi.org/ 10.3389/fmed.2022.999440
- Zhang, Y.-Y., Yang, W., Zhang, Y., Hu, Z., Chen, Y., Ma, Y.,... Zhang, S. (2023). HucMSC-EVs Facilitate In Vitro Development of Maternally Aged Preantral Follicles and Oocytes. Stem Cell Reviews and Reports, 19(5), 1427–1448. https://doi.org/10.1007/ s12015-022-10495-w.

- Na, J., & Kim, G. J. (2020). Recent trends in stem cell therapy for premature ovarian insufficiency and its therapeutic potential: A review. *Journal of Ovarian Research*, 13(1), 74. https://doi.org/1 0.1186/s13048-020-00671-2
- Song, Y., Wu, J., Liu, Y., Xu, N., Bai, H., Wang, L.,... Li, K. (2024). The remodeling of ovarian function: targeted delivery strategies for mesenchymal stem cells and their derived extracellular vesicles. Stem Cell Research & Therapy, 15(1), 90. https://doi.org/10.1186/s13287-024-03704-5.
- Yuan, Z., Zhang, Y., He, X., Wang, X., Wang, X., Ren, S.,... Xiao,
   Z. (2024). Engineering mesenchymal stem cells for premature ovarian failure: overcoming challenges and innovating therapeutic strategies. Theranostics, 14(17), 6487–6515. https://doi.org/10.7150/thno.102641.
- 93. Roof, K. A., Andre, K. E., Modesitt, S. C., & Schirmer, D. A. (2024). Maximizing ovarian function and fertility following chemotherapy in premenopausal patients: Is there a role for ovarian suppression? *Gynecologic Oncology Reports*, *53*, 101383. https://doi.org/10.1016/j.gore.2024.101383
- Seckin, S., Ramadan, H., Mouanness, M., Kohansieh, M., & Merhi, Z. (2022). Ovarian response to intraovarian platelet-rich plasma (PRP) administration: Hypotheses and potential mechanisms of action. *Journal of Assisted Reproduction and Genetics*, 39(1), 37–61. https://doi.org/10.1007/s10815-021-02385-w
- Galderisi, U., Peluso, G., & Di Bernardo, G. (2022). Clinical trials based on mesenchymal stromal cells are exponentially increasing: Where are we in recent years?? Stem Cell Reviews and Reports, 18(1), 23–36. https://doi.org/10.1007/s12015-021-10231-w
- Kabat, M., Bobkov, I., Kumar, S., & Grumet, M. (2020). Trends in mesenchymal stem cell clinical trials 2004–2018: Is efficacy optimal in a narrow dose range? *Stem Cells Translational Medicine*, 9(1), 17–27. https://doi.org/10.1002/sctm.19-0202
- Gowen, A., Shahjin, F., Chand, S., Odegaard, K. E., & Yelamanchili, S. V. (2020). Mesenchymal stem cell-derived extracellular vesicles: Challenges in clinical applications. Frontiers in Cell and Developmental Biology. https://doi.org/10.3389/fcell.2020.0014
- Roszkowski, S. (2024). Therapeutic potential of mesenchymal stem cell-derived exosomes for regenerative medicine applications. *Clinical and Experimental Medicine*, 24(1), 46. https://doi. org/10.1007/s10238-023-01282-z
- Katsuda, T., Kosaka, N., Takeshita, F., & Ochiya, T. (2013). The therapeutic potential of mesenchymal stem cell-derived extracellular vesicles. *Proteomics*, *13*(10–11), 1637–1653. https://doi.org/ 10.1002/pmic.201200373
- 100. Qu, Q., Liu, L., Wang, L., Cui, Y., Liu, C., Jing, X., & Xu, X. (2024). Exosomes derived from hypoxic mesenchymal stem cells restore ovarian function by enhancing angiogenesis. *Stem Cell Research & Therapy*, 15(1), 496. https://doi.org/10.1186/s13287-024-04111-6
- 101. Fernández-Messina, L., Gutiérrez-Vázquez, C., Rivas-García, E., Sánchez-Madrid, F., & de la Fuente, H. (2015). Immunomodulatory role of MicroRNAs transferred by extracellular vesicles. *Biology of the Cell*, 107(3), 61–77. https://doi.org/10.1111/boc.20 1400081
- 102. Kumar, M. A., Baba, S. K., Sadida, H. Q., Marzooqi, S. Al., Jerobin, J., Altemani, F. H.,... Bhat, A. A. (2024). Extracellular vesicles as tools and targets in therapy for diseases. Signal Transduction and Targeted Therapy, 9(1), 27. https://doi.org/10.1038/s 41392-024-01735-1.
- 103. Tang, P., Song, F., Chen, Y., Gao, C., Ran, X., Li, Y.,... Zhou, C. (2024). Preparation and characterization of extracellular vesicles and their cutting-edge applications in regenerative medicine. Applied Materials Today, 37, 102084. https://doi.org/10.1016/j.apmt.2024.102084.



- 104. Zhang, Y., Wu, D., Zhou, C., Bai, M., Wan, Y., Zheng, Q.,... Yang, C. (2024). Engineered extracellular vesicles for tissue repair and regeneration. Burns & Trauma, 12. https://doi.org/10.1093/burnst/tkae062.
- 105. Ye, J., Li, D., Jie, Y., Luo, H., Zhang, W., & Qiu, C. (2024). Exosome-based nanoparticles and cancer immunotherapy. *Biomedicine & Pharmacotherapy*, 179, 117296. https://doi.org/10.1016/j.biopha.2024.117296
- 106. Zeng, H., Guo, S., Ren, X., Wu, Z., Liu, S., & Yao, X. (2023). Current strategies for exosome cargo loading and targeting delivery. *Cells*, 12(10), 1416. https://doi.org/10.3390/cells12101416
- 107. Zhang, Y., Bi, J., Huang, J., Tang, Y., Du, S., & Li, P. (2020). Exosome: A review of its classification, isolation techniques, storage, diagnostic and targeted therapy applications. *International Journal of Nanomedicine*, 6917–6934. https://doi.org/10.2147/IJN.S264498
- 108. Sun, T., Li, M., Liu, Q., Yu, A., Cheng, K., Ma, J.,... Zhang, Y. (2024). Insights into optimizing exosome therapies for acute skin wound healing and other tissue repair. Frontiers of Medicine, 18(2), 258–284. https://doi.org/10.1007/s11684-023-1031-9.
- 109. Lai, J. J., Chau, Z. L., Chen, S., Hill, J. J., Korpany, K. V., Liang, N.,... Shen, W. (2022). Exosome Processing and Characterization Approaches for Research and Technology Development. Advanced Science, 9(15). https://doi.org/10.1002/advs.2021032 22.
- 110. Li, J., Wang, J., & Chen, Z. (2025). Emerging role of exosomes in cancer therapy: Progress and challenges. *Molecular Cancer*, 24(1), 13. https://doi.org/10.1186/s12943-024-02215-4
- 111. Fan, W., Qi, Y., Wang, Y., Yan, H., Li, X., & Zhang, Y. (2023). Messenger roles of extracellular vesicles during fertilization of gametes, development and implantation: Recent advances. Frontiers in Cell and Developmental Biology. https://doi.org/10.3389/fcell.2022.1079387
- 112. Olejarz, W., Kubiak-Tomaszewska, G., Chrzanowska, A., & Lorenc, T. (2020). Exosomes in angiogenesis and anti-angiogenic therapy in cancers. *International Journal of Molecular Sciences*, 21(16), 5840. https://doi.org/10.3390/ijms21165840
- 113. Awadasseid, A., Wu, Y., & Zhang, W. (2021). Extracellular vesicles (Exosomes) as immunosuppressive mediating variables in tumor and chronic inflammatory microenvironments. *Cells*, 10(10), 2533. https://doi.org/10.3390/cells10102533
- 114. Zubair, M., Abouelnazar, F. A., Iqbal, M. A., Pan, J., Zheng, X., Chen, T.,... Chu, Y. (2025). Mesenchymal stem cell-derived exosomes as a plausible immunomodulatory therapeutic tool for inflammatory diseases. Frontiers in Cell and Developmental Biology, 13. https://doi.org/10.3389/fcell.2025.1563427.
- 115. Zhang, W., Liu, R., Chen, Y., Wang, M., & Du, J. (2022). Cross-talk between Oxidative Stress and Exosomes. *Oxidative Medicine and Cellular Longevity*, 2022, 1–11. https://doi.org/10.1155/2022/3553617
- 116. Asgarpour, K., Shojaei, Z., Amiri, F., Ai, J., Mahjoubin-Tehran, M., Ghasemi, F.,... Mirzaei, H. (2020). Exosomal microRNAs derived from mesenchymal stem cells: cell-to-cell messages. Cell Communication and Signaling, 18(1), 149. https://doi.org/10.1186/s12964-020-00650-6.
- 117. Yu, X., Odenthal, M., & Fries, J. (2016). Exosomes as miRNA carriers: Formation–function–future. *International Journal of Molecular Sciences*, 17(12), Article 2028. https://doi.org/10.3390/ijms17122028
- Dalmizrak, A., & Dalmizrak, O. (2022). Mesenchymal stem cellderived exosomes as new tools for delivery of miRNAs in the treatment of cancer. Frontiers in Bioengineering and Biotechnology. https://doi.org/10.3389/fbioe.2022.956563
- 119. Worzfeld, T., von Strandmann, P., Huber, E., Adhikary, M., Wagner, T., Reinartz, U., S., & Müller, R. (2017). The unique

- molecular and cellular microenvironment of ovarian cancer. *Frontiers in Oncology*, 7. https://doi.org/10.3389/fonc.2017.000
- 120. Li, X., Liu, Y., Zheng, S., Zhang, T., Wu, J., Sun, Y.,... Liu, G. (2021). Role of exosomes in the immune microenvironment of ovarian cancer (Review). Oncology Letters, 21(5), 377. https://doi.org/10.3892/ol.2021.12638.
- 121. Tan, F., Li, X., Wang, Z., Li, J., Shahzad, K., & Zheng, J. (2024). Clinical applications of stem cell-derived exosomes. *Signal Transduction and Targeted Therapy*, 9(1), 17. https://doi.org/10.1038/s41392-023-01704-0
- 122. Zhou, A. K., Jou, E., Lu, V., Zhang, J., Chabra, S., Abishek, J.,... Guo, B. (2023). Using Pre-Clinical Studies to Explore the Potential Clinical Uses of Exosomes Secreted from Induced Pluripotent Stem Cell-Derived Mesenchymal Stem cells. Tissue Engineering and Regenerative Medicine, 20(6), 793–809. https://doi.org/10.1007/s13770-023-00557-6.
- 123. Cavalcante, M. B., Sampaio, O. G. M., Câmara, F. E. A., Schneider, A., de Ávila, B. M., Prosczek, J.,... Campos, A. R. (2023). Ovarian aging in humans: potential strategies for extending reproductive lifespan. GeroScience, 45(4), 2121–2133. https://doi.org/10.1007/s11357-023-00768-8.
- 124. Yang, W., Zhang, J., Xu, B., He, Y., Liu, W., Li, J., ... Li, J. (2020). HucMSC-Derived Exosomes Mitigate the Age-Related Retardation of Fertility in Female Mice. Molecular Therapy, 28(4), 1200–1213. https://doi.org/10.1016/j.ymthe.2020.02.003.
- 125. Dilsiz, N. (2024). A comprehensive review on recent advances in exosome isolation and characterization: Toward clinical applications. *Translational Oncology*, 50, 102121. https://doi.org/10.101 6/j.tranon.2024.102121
- 126. Lee, Y. J., Shin, K. J., & Chae, Y. C. (2024). Regulation of cargo selection in exosome biogenesis and its biomedical applications in cancer. *Experimental and Molecular Medicine*, *56*(4), 877–889. https://doi.org/10.1038/s12276-024-01209-y
- 127. Park, H., Seok, J., Cetin, E., Ghasroldasht, M. M., Liakath Ali, F., Mohammed, H.,... Al-Hendy, A. (2024). Fertility protection: a novel approach using pretreatment with mesenchymal stem cell exosomes to prevent chemotherapy—induced ovarian damage in a mouse model. American Journal of Obstetrics and Gynecology, 231(1), 111.e1-111.e18. https://doi.org/10.1016/j.ajog.2024.02.0
- 128. Wang, Y., Xiong, J., Ouyang, K., Ling, M., Luo, J., Sun, J.,... Zhang, Y. (2025). Extracellular vesicles: From large-scale production and engineering to clinical applications. Journal of Tissue Engineering, 16. https://doi.org/10.1177/20417314251319474.
- 129. Zhou, T., Yuan, Z., Weng, J., Pei, D., Du, X., He, C., & Lai, P. (2021). Challenges and advances in clinical applications of mesenchymal stromal cells. *Journal of Hematology & Oncology*, 14(1), 24. https://doi.org/10.1186/s13045-021-01037-x
- 130. Barekzai, J., Refflinghaus, L., Okpara, M., Tasto, L., Tertel, T., Giebel, B.,...Salzig, D. (2024). Process development for the production of mesenchymal stromal cell-derived extracellular vesicles in conventional 2D systems. Cytotherapy, 26(9), 999–1012. https://doi.org/10.1016/j.jcyt.2024.04.071.
- 131. Hoang, V. T., Trinh, Q.-M., Phuong, D. T. M., Bui, H. T. H., Hang, L. M., Ngan, N.T. H.,... Hoang, D. M. (2021). Standardized xeno- and serum-free culture platform enables large-scale expansion of high-quality mesenchymal stem/stromal cells from perinatal and adult tissue sources. Cytotherapy, 23(1), 88–99. htt ps://doi.org/10.1016/j.jcyt.2020.09.004.
- 132. Naeem, A., Gupta, N., Naeem, U., Khan, M. J., Elrayess, M. A., Cui, W., & Albanese, C. (2022). A comparison of isolation and culture protocols for human amniotic mesenchymal stem cells. Cell Cycle, 21(15), 1543–1556. https://doi.org/10.1080/1538410 1.2022.2060641



- 133. Adlerz, K., Patel, D., Rowley, J., Ng, K., & Ahsan, T. (2020). Strategies for scalable manufacturing and translation of MSC-derived extracellular vesicles. *Stem Cell Research*, 48, 101978. https://doi.org/10.1016/j.scr.2020.101978
- 134. Kang, H., Feng, J., Peng, Y., Liu, Y., Yang, Y., Wu, Y.,... He, Y. (2023). Human mesenchymal stem cells derived from adipose tissue showed a more robust effect than those from the umbilical cord in promoting corneal graft survival by suppressing lymphangiogenesis. Stem Cell Research & Therapy, 14(1), 328. https://doi.org/10.1186/s13287-023-03559-2.
- 135. Yasumura, Y., Teshima, T., Taira, Y., Saito, T., Yuchi, Y., Suzuki, R., & Matsumoto, H. (2022). Optimal intravenous administration procedure for efficient delivery of canine adipose-derived mesenchymal stem cells. *International Journal of Molecular Sciences*, 23(23), 14681. https://doi.org/10.3390/ijms232314681
- 136. Bagno, L. L., Salerno, A. G., Balkan, W., & Hare, J. M. (2022). Mechanism of action of mesenchymal stem cells (MSCs): Impact of delivery method. *Expert Opinion on Biological Therapy*, 22(4), 449–463. https://doi.org/10.1080/14712598.2022.2016695
- 137. Silvén, H., Savukoski, S. M., Pesonen, P., Pukkala, E., Ojaniemi, M., Gissler, M.,... Niinimäki, M. (2023). Association of genetic disorders and congenital malformations with premature ovarian insufficiency: a nationwide register-based study. Human Reproduction, 38(6), 1224–1230. https://doi.org/10.1093/humrep/dead 066.
- 138. Umer, A., Ahmad, K., Khan, N., Greene, D. L., Shamim, S., & Habiba, U. E. (2024). Meta-analysis highlight the therapeutic potential of stem cells for premature ovarian failure. *Regenerative Therapy*, 26, 478–488. https://doi.org/10.1016/j.reth.2024.07001
- 139. Yan, L., Wu, Y., Li, L., Wu, J., Zhao, F., Gao, Z.,... Wang, H. (2020). Clinical analysis of human umbilical cord mesenchymal stem cell allotransplantation in patients with premature ovarian insufficiency. Cell Proliferation, 53(12). https://doi.org/10.1111/cpr.12938.
- 140. Wang, Y., Jiang, J., Zhang, J., Fan, P., & Xu, J. (2023). Research progress on the etiology and treatment of premature ovarian insufficiency. *Biomedicine Hub*, 8(1), 97–107. https://doi.org/10.1159/000535508
- 141. Reza-Zaldivar, E. E., Hernández-Sapiéns, M. A., Minjarez, B., Gutiérrez-Mercado, Y. K., Márquez-Aguirre, A. L., & Canales-Aguirre, A. A. (2018). Potential effects of MSC-derived exosomes in neuroplasticity in Alzheimer's disease. Frontiers in Cellular Neuroscience. https://doi.org/10.3389/fncel.2018.00317
- 142. Nagpal, A., Milte, R., Kim, S. W., Hillier, S., Hamilton-Bruce, M. A., Ratcliffe, J., & Koblar, S. A. (2019). Economic evaluation of stem cell therapies in neurological diseases: A systematic review. *Value in Health*, 22(2), 254–262. https://doi.org/10.1016/j.jval.2018.07.878
- 143. Thavorn, K., van Katwyk, S., Krahn, M., Mei, S. H. J., Stewart, D. J., Fergusson, D.,... McIntyre, L. (2020). Value of mesenchymal

- stem cell therapy for patients with septic shock: an early health economic evaluation. International Journal of Technology Assessment in Health Care, 36(5), 525–532. https://doi.org/10.1017/S0266462320000781.
- 144. Li, M., Jiang, Y., Hou, Q., Zhao, Y., Zhong, L., & Fu, X. (2022). Potential pre-activation strategies for improving therapeutic efficacy of mesenchymal stem cells: Current status and future prospects. Stem Cell Research & Therapy, 13(1), 146. https://doi.org/10.1186/s13287-022-02822-2
- 145. Cheng, L., Zhang, K., Wu, S., Cui, M., & Xu, T. (2017). Focus on mesenchymal stem cell-derived exosomes: Opportunities and challenges in cell-free therapy. *Stem Cells International*, 2017, 1–10. https://doi.org/10.1155/2017/6305295
- 146. Mokhtari, R. B., Homayouni, T. S., Baluch, N., Morgatskaya, E., Kumar, S., Das, B., & Yeger, H. (2017). Combination therapy in combating cancer. *Oncotarget*, 8(23), 38022–38043. https://doi.org/10.18632/oncotarget.16723
- 147. Zhao, X., Hu, D. A., Wu, D., He, F., Wang, H., Huang, L.,... Athiviraham, A. (2021). Applications of Biocompatible Scaffold Materials in Stem Cell-Based Cartilage Tissue Engineering. Frontiers in Bioengineering and Biotechnology, 9. https://doi.org/10.3389/fbioe.2021.603444.
- 148. Li, S., Dan, X., Chen, H., Li, T., Liu, B., Ju, Y.,... Fan, X. (2024). Developing fibrin-based biomaterials/scaffolds in tissue engineering. Bioactive Materials, 40, 597–623. https://doi.org/10.1016/j.bioactmat.2024.08.006.
- 149. Hong, I. S. (2022). Enhancing stem cell-based therapeutic potential by combining various bioengineering technologies. Frontiers in Cell and Developmental Biology. https://doi.org/10.3389/fcell. 2022.901661
- Akhondzadeh, S. (2014). Personalized medicine: A tailor made medicine. Avicenna Journal of Medical Biotechnology, 6(4), 191.
- 151. Zhang, T., Zhang, M., Zhang, S., & Wang, S. (2024). Research advances in the construction of stem cell-derived ovarian organoids. Stem Cell Research & Therapy, 15(1), 505. https://doi.org/1 0.1186/s13287-024-04122-3
- 152. Maenhoudt, N., Defraye, C., Boretto, M., Jan, Z., Heremans, R., Boeckx, B.,... Vankelecom, H. (2020). Developing Organoids from Ovarian Cancer as Experimental and Preclinical Models. Stem Cell Reports, 14(4), 717–729. https://doi.org/10.1016/j.stemcr.2020.03.004
- 153. Ashmore-Harris, C., Iafrate, M., Saleem, A., & Fruhwirth, G. O. (2020). Non-invasive reporter gene imaging of cell therapies, including T cells and stem cells. *Molecular Therapy*, 28(6), 1392–1416. https://doi.org/10.1016/j.ymthe.2020.03.016

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