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## CAR-T 疗法在未分化甲状腺癌中靶点探索的研究进展

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**摘要:** 目的 探讨嵌合抗原受体 T 细胞 (Chimeric Antigen Receptor T-cell, CAR-T) 疗法在未分化甲状腺癌 (Anaplastic Thyroid Cancer, ATC) 治疗中的靶点研究与应用进展。方法 综合分析近年来国内外关于 CAR-T 疗法在 ATC 中的研究, 归纳其主要靶点 (如 *ICAM-1*、*TSHR*、*CSPG4*、*B7-H3*、*ROR1* 等) 及作用机制, 并分析其临床应用中的关键问题。结果 CAR-T 可通过靶向多种 ATC 相关抗原发挥抗肿瘤作用, 但治疗仍受到肿瘤异质性、免疫抑制微环境及安全风险等因素限制。结论 深入开展 ATC 相关抗原的探索与 CAR 结构优化, 结合联合治疗策略, 有望提升 CAR-T 疗法的治疗效能, 为 ATC 的精准免疫治疗提供新的方向与理论基础。

**关键词:** 未分化甲状腺癌; 嵌合抗原受体 T 细胞 (CAR-T); 免疫治疗

### Research Progress in Target Exploration of CAR-T Therapy for Anaplastic Thyroid Cancer

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**Abstract: Objective** To explore the advances in targets and applications of chimeric antigen receptor T-cell (CAR-T) therapy in the treatment of anaplastic thyroid cancer (ATC). **Methods** Recent literature on studies on CAR-T therapy in anaplastic thyroid cancer (ATC) was retrieved to summarize the principal therapeutic targets, such as *ICAM-1*, *TSHR*, *CSPG4*, *B7-H3*, and *ROR1*, and their underlying mechanisms of action, and to identify major problems encountered in clinical applications. **Results** CAR-T cells exerted antitumor activity by targeting multiple ATC-associated antigens. However, their therapeutic efficacy was still limited by such factors as tumor heterogeneity, immunosuppressive tumor microenvironments, and potential safety concerns. **Conclusion** Insights into ATC-associated antigens and optimization of CAR design, in combination with integrated therapeutic strategies, may enhance the efficacy of CAR-T therapy and offer clues to precision immunotherapy in ATC.

**Keywords:** Anaplastic Thyroid Cancer; Chimeric Antigen Receptor T-Cell (CAR-T); Immunotherapy

甲状腺癌 (Thyroid Cancer, TC) 是常见的内分泌恶性肿瘤, 2022 年全球甲状腺癌新发病例约 821 173 例<sup>[1]</sup>。未分化甲状腺癌 (Anaplastic Thyroid Cancer, ATC) 虽仅占甲状腺癌的 1% ~ 2%, 却是最具侵袭性的亚型, 导致约 80% 的甲状腺癌相关死亡, 患者中位生存期通常为数月<sup>[2-3]</sup>。临床上, ATC 常表现为肿瘤快速生长、局部浸润及压迫症状; 约 50% 病例确诊时已发生远处转移, 肺为最常见受累器官之一<sup>[4-5]</sup>。ATC

目前以手术、放疗、化疗、靶向治疗、免疫治疗等综合治疗为主, 治疗效果欠佳。过继细胞疗法 (Adoptive T-Cell Therapy, ACT) 中发展出的嵌合抗原受体 T 细胞 (Chimeric Antigen Receptor T-cell, CAR-T) 为难治性 ATC 提供了新的研究方向<sup>[7]</sup>。本文旨在综述 CAR-T 疗法在 ATC 中的研究进展, 重点关注靶点探索、临床疗效、现有挑战及应对策略。

#### 1 CAR-T 疗法的概述

嵌合抗原受体 (Chimeric Antigen Receptor, CAR) 于 1989 年提出<sup>[8]</sup>, 其设计将 T 细胞受体 (T-Cell Receptor, TCR) 的细胞内信号域与抗体可变区融合, 赋予 T 细胞识别肿瘤抗原并发挥细胞毒性的能力。基于此概念发展出 CAR-T 疗法是一种革新的免疫治疗

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方法,其核心原理是从患者体内分离 T 细胞,通过基因工程将识别肿瘤抗原的 CAR 导入细胞,扩增后回输给患者,使 T 细胞可非依赖主要组织相容性复合体(Major Histocompatibility Complex, MHC)识别并杀伤目标细胞<sup>[9]</sup>。典型 CAR 结构包括 4 个关键模块:细胞外抗原识别域(Single-Chain Variable Fragment, scFv)、铰链区、跨膜域及细胞内信号域<sup>[10]</sup>。自提出以来, CAR 设计经历多代优化:第 1 代 CAR 仅包含 CD3 信号域<sup>[8]</sup>;第 2 代增加共刺激结构域(如 CD28 或 CD137)以增强 T 细胞活性<sup>[11]</sup>;第 3 代进一步添加第 2 个共刺激结构域(如 CD28、CD137 或 ICOS)<sup>[12]</sup>;第 4 代 CAR(通用细胞因子介导杀伤重定向 T 细胞, TRUCKs)整合额外功能模块<sup>[13]</sup>;第 5 代在第 2 代基础上引入可激活 JAK-STAT 信号的结构域(如 IL-2R 片段),以增强 T 细胞增殖、存活及抗肿瘤能力<sup>[14]</sup>。CAR-T 疗法已成为癌症免疫治疗的重要里程碑,尤其在血液系统恶性肿瘤中表现突出。2017 年,美国食品药品监督管理局(FDA)批准 Kymriah,用于治疗难治性及复发性 B 细胞急性淋巴细胞白血病<sup>[15]</sup>。目前, CAR-T 疗法的研究正在向实体瘤拓展, ATC 亦是潜在应用方向。

## 2 ATC 的治疗现状

随着分子生物学与免疫学的深入发展, ATC 的治疗策略已从传统手术、放疗和化疗拓展至靶向治疗、免疫治疗及纳米医学。SMALLRIDGE 等<sup>[16]</sup>总结 BRAF、RAS、PIK3CA 及 TP53 等基因突变及染色体异常,为靶向治疗提供了理论基础。基于此,靶向治疗取得突破:2018 年, FDA 批准达拉菲尼(Dabrafenib)联合曲美替尼(Trametinib)用于 BRAF V600E 突变阳性 ATC 患者,临床显示客观缓解率 69%, 12 个月生存率达 80%<sup>[17]</sup>。在免疫治疗方面,有研究表明 ATC 肿瘤微环境中 CD8<sup>+</sup>T 细胞存在浸润及耗竭特征,为免疫检查点抑制剂(Immune Checkpoint Inhibitors, ICIs)的应用奠定基础<sup>[18]</sup>。CAR-T 疗法则聚焦于靶向细胞间黏附分子-1(Intercellular Adhesion Molecule-1, ICAM-1),该分子在 ATC 中高表达,既促进肿瘤细胞增殖、迁移和抗凋亡,又增强肿瘤对 CAR-T 细胞的敏感性<sup>[19]</sup>。靶向治疗联合免疫治疗显示出潜在前景,但现有临床研究多为单臂 II 期试验<sup>[20-21]</sup>。未来研究需纳入对照组并延长随访,以获得更可靠的疗效和安全性数据。此外,纳米医学在

药物递送系统中表现出应用潜力。有团队开发的碘化钾修饰纳米黏土载体,通过特异性甲状腺组织靶向及调控细胞自噬/内吞,在动物模型中使 ATC 原发灶肿瘤抑制率达 90% 以上<sup>[22]</sup>。虽然尚未有纳米药物获批临床,但其在化疗-靶向联合治疗中具备转化潜力。综上, ATC 治疗正朝向个体化、多模式整合方向发展。未来研究应聚焦关键肿瘤特异性靶点机制(包括突变基因及免疫逃逸通路)、优化靶向-免疫联合方案,并推动纳米技术临床转化,以期改善患者预后。

## 3 CAR-T 在 ATC 治疗中的靶点

### 3.1 ICAM-1

ICAM-1 属于免疫球蛋白超家族,广泛分布于多种细胞表面<sup>[23]</sup>。在 ATC 及其他恶性肿瘤中, ICAM-1 呈过表达状态,通过促进肿瘤细胞增殖、迁移并抑制凋亡,驱动肿瘤侵袭与转移<sup>[24-25]</sup>。高表达 ICAM-1 可显著增强肿瘤对 CAR-T 疗法的敏感性。2017 年, MIN 等<sup>[26]</sup>报道靶向 ICAM-1 的 CAR-T 疗法,体外实验显示其对 ICAM-1 高表达的 ATC 细胞系具有高效特异性杀伤作用; NSG 小鼠转移瘤模型进一步证明该疗法可显著抑制肿瘤生长、延长生存并延缓疾病进展,同时对非肿瘤细胞毒性有限,无明显关键器官损伤。随后,该团队开发第 3 代 ICAM-1 CAR-T,通过优化靶向结合能力,实现对高表达肿瘤细胞的选择性杀伤,同时降低对低表达正常组织的影响<sup>[27]</sup>。2025 年,美国癌症研究协会年会上,公布 AIC100 临床研究结果:第 3 代 CAR-T 采用亲和力优化技术靶向高表达 ICAM-1,并共表达生长抑素受体 2 以实现 DOT-ATATE PET 实时监测。I 期临床试验(NCT04420754)显示,在 DL2/DL3 剂量组( $1 \times 10^8 \sim 5 \times 10^8$  CAR-T 细胞)中, ATC 患者客观缓解率达 50%, 疾病控制率为 56%;治疗相关毒性总体可控,而 DL4 组出现 3 级肺炎提示剂量上限。基于安全性与疗效, DL3 ( $5 \times 10^8$ ) 被确立为 II 期推荐剂量,扩展研究预计于 2025 年启动<sup>[28]</sup>。综上,靶向 ICAM-1 的 CAR-T 疗法已进入临床验证阶段,其良好的疗效和安全性表明,该策略有望成为复发/难治性 ATC 治疗的新突破。

### 3.2 促甲状腺激素受体(Thyroid Stimulating Hormone Receptor, TSHR)

TSHR 是甲状腺组织特异性抗原,在分化型甲状腺癌(Differentiated Thyroid Carcinoma, DTC)中高

表达,但在 ATC 中普遍失活<sup>[29]</sup>。*TSHR* 靶点的一个挑战是其在 ATC 中因表观遗传变化而下调,使其失去疗效。研究发现,ATC 中 *TSHR* 基因启动子常被高度甲基化<sup>[30]</sup>,并伴随组蛋白去乙酰化及其他抑制性表观遗传修饰,从而阻断 TSH 依赖的甲状腺分化基因表达<sup>[31-32]</sup>。如 ATC 细胞中过表达的 HN1 蛋白可募集 *HDAC2* 去乙酰化 *H3K27*,降低甲状腺分化基因的可及性<sup>[32]</sup>。此外,*SWI/SNF* 染色体重塑复合体的突变可导致染色质压缩,使得 *MAPK* 抑制剂 (*MAPKi*) 无法完全逆转分化基因的沉默<sup>[33]</sup>。已有的临床研究多集中在非 ATC 类型,如 DING 等<sup>[34]</sup>报道分化不良甲状腺癌病例。值得注意的是,*MAPKi* 可在 ATC PDX 模型中诱导 *TSHR* 再表达,且需持续治疗维持<sup>[35]</sup>。动物实验提示,*MAPKi* 联合 *TSHR CAR-T* 可协同抑瘤,而单用 *MAPKi* 仅能部分恢复甲状腺特异性基因表达<sup>[33]</sup>。LI 等<sup>[29]</sup>开发的 *TSHR CAR-T* 在 DTC 模型中显示出强效杀伤作用且未见明显毒性。因此,在 ATC 中,通过 *MAPKi* “重编程”表观遗传状态以恢复 *TSHR* 表达,再结合 *TSHR CAR-T*,有望实现精准且时间窗口依赖的治疗,为未来联合治疗提供思路。

### 3.3 硫酸软骨素蛋白聚糖 4 (Chondroitin Sulfate Proteoglycan 4, *CSPG4*) 和 B7 同系物 3 (B7 homolog 3, *B7-H3*)

*CSPG4* 和 *B7-H3* 均在多种实体瘤(如黑色素瘤、胶质瘤)中过表达<sup>[36-37]</sup>,且在 ATC 组织及细胞系中显著上调,而正常甲状腺组织表达有限<sup>[38-39]</sup>。*CSPG4* 可能通过激活整合素 *FAK* 通路促进肿瘤侵袭和血管生成,而 *B7-H3* 则通过抑制 T 细胞功能促进免疫逃逸。CATTANEO 等<sup>[39]</sup>比较两者的 *CAR-T*

效果发现:*B7-H3 CAR-T* 在 ATC 小鼠模型中表现出更高的肿瘤清除率(5/5 小鼠完全缓解),而 *CSPG4 CAR-T* 仅在 2/5 小鼠中实现完全缓解;体外实验亦显示 *B7-H3 CAR-T* 诱导更强的细胞因子分泌和杀伤活性。然而,*CSPG4 CAR-T* 在 40% 动物中仍能清除 ATC 灶,尤其在 *BRAF V600E* 阴性或靶向治疗后复发病例中具有潜在价值<sup>[40]</sup>。综上,*B7-H3* 是更具潜力的 ATC *CAR-T* 靶点,而 *CSPG4* 则可能为特定分子亚型的 ATC 提供补充选择。

### 3.4 其他潜在靶点

除前述靶点外,胚胎型酪氨酸激酶受体 1 (Receptor Tyrosine Kinase Like Orphan Receptor 1, *ROR1*) 在多种恶性肿瘤中高表达,而在正常组织中几乎不表达。*ROR1* 能够同时激活 *PI3K-AKT*、*MEK-ERK*、*STAT3*、*c-Met* 和 *EGFR* 等多条信号通路,从而促进肿瘤细胞的增殖、转移及存活<sup>[41]</sup>。体外实验表明,*ROR1 CAR-T* 可特异性识别并裂解 *ROR1<sup>+</sup>ATC* 细胞,同时分泌大量 *IFN-γ*、*TNF-α* 等关键效应因子,显示出明显的靶点依赖性杀伤作用<sup>[42]</sup>。值得注意的是,*ROR1* 的表达不受常用激酶抑制剂的影响,这提示其在联合治疗或耐药后的后续治疗中可能具有独特优势。虽然目前仍处于临床前研究阶段,但凭借高度特异性及多通路调控潜力,*ROR1 CAR-T* 有望成为 ATC 精准免疫治疗的重要新兴靶点。关于 *ICAM-1*、*TSHR*、*CSPG4*、*B7-H3* 及 *ROR1* 等 *CAR-T* 靶点研究进展见表 1。

## 4 *CAR-T* 治疗 ATC 的挑战与策略

*CAR-T* 疗法在血液系统肿瘤中已取得突破性进展<sup>[43-44]</sup>,但在实体瘤领域的应用仍受到免疫抑制

表 1 不同靶点 *CAR-T* 疗法的研究进展  
Table 1 Research Progress in *CAR-T* Therapies Targeting Different Antigens in Anaplastic Thyroid Cancer

靶点	表达特点	主要作用机制	研究进展	临床试验注册编号
<i>ICAM-1</i>	在 ATC 细胞表面高表达,正常组织低表达	促进细胞增殖、迁移并抑制凋亡	体外和小鼠模型显示显著抗肿瘤活性; I 期临床试验 ( <i>AIC100</i> ) 客观缓解率 50%, 疾病控制率 56%, DL3 剂量推荐用于 II 期扩展研究	NCT04420754
<i>TSHR</i>	DTC 中高表达, ATC 中常下调或沉默	表观遗传修饰(甲基化、去乙酰化)致下调; 可通过 <i>MAPK</i> 抑制剂部分恢复	DTC 模型中 <i>CAR-T</i> 显示有效; ATC 中需联合 <i>MAPKi</i> 诱导再表达方可发挥疗效	NCT04925206
<i>CSPG4</i>	在 ATC 组织及细胞系显著上调, 正常甲状腺组织有限表达	激活整合素 <i>FAK</i> 通路, 促进侵袭和血管生成	临床前研究中 <i>CAR-T</i> 具杀伤作用, 部分小鼠模型完全缓解	暂无
<i>B7-H3</i>	ATC 中高表达, 正常组织有限	抑制 T 细胞功能, 促进免疫逃逸	临床前研究显示抗肿瘤效果优于 <i>CSPG4 CAR-T</i> , 小鼠模型中实现完全缓解	暂无
<i>ROR1</i>	多种恶性肿瘤及 ATC 中高表达, 正常组织不表达	激活 <i>PI3K-AKT</i> 、 <i>MEK-ERK</i> 、 <i>STAT3</i> 、 <i>c-Met</i> 、 <i>EGFR</i> 等通路, 促进增殖与转移	体外研究显示 <i>CAR-T</i> 可特异性杀伤 <i>ROR1<sup>+</sup>ATC</i> 细胞并分泌效应因子, 处于临床前阶段	暂无

性肿瘤微环境 (Tumor Microenvironment, TME) 的显著限制<sup>[45-47]</sup>。ATC 的 TME 不仅存在高度异质性的肿瘤细胞群, 还伴随多种抑制性因子和免疫调节细胞, 限制 CAR-T 细胞在局部的扩增和效应功能。其中, *IL-10* 和 *TGF- $\beta$*  是关键的抑制分子。*IL-10* 可通过激活 *JAK1/TYK2-STAT3* 通路, 降低炎症性细胞因子 (如 *IFN- $\gamma$* 、*TNF- $\alpha$* ) 的表达, 并削弱树突状细胞的抗原呈递, 从而限制 T 细胞的活化与增殖<sup>[48]</sup>。*TGF- $\beta$*  则通过 *SMAD* 依赖性信号抑制细胞毒分子生成, 并促进 *FOXP3*<sup>+</sup> 调节性 T 细胞扩展, 进一步维持局部免疫抑制环境。研究显示, 阻断 *TGF- $\beta$*  信号通路有助于增强 CAR-T 细胞的持久性与抗肿瘤效应<sup>[49]</sup>。代谢失衡亦是影响 CAR-T 疗效的重要障碍。肿瘤微环境常伴随缺氧、乳酸积聚和低葡萄糖环境, 这些因素导致 CAR-T 能量代谢受限, 线粒体功能下降, 并易进入耗竭状态<sup>[50]</sup>。由此可见, TME 中的细胞因子与代谢应激共同削弱了 CAR-T 的效应功能。为突破这些限制, 多种优化策略被提出。如利用工程化细菌在肿瘤局部定植并释放人工抗原, 从而实现“肿瘤再标记”, 提高 CAR-T 的识别效率<sup>[51]</sup>; 纳米递送平台能够将抗原或免疫刺激分子精确释放于肿瘤内, 以激活机体已有的免疫记忆, 从而间接增强 CAR-T 活性<sup>[52]</sup>; 联合免疫检查点抑制剂或代谢调控手段也被认为具有潜力, 可部分逆转 T 细胞耗竭并改善能量代谢状态<sup>[53]</sup>。

尽管针对 *ICAM-1*、*TSHR*、*CSPG4*、*B7-H3* 和 *ROR1* 等抗原的 CAR-T 在临床前研究中已展现出一定的抗 ATC 效果, 但如何突破免疫抑制性 TME、降低脱靶毒性并确保长期安全性仍是亟需解决的问题。在此背景下, 新型基于 CAR 的细胞疗法 (如 *CAR-NK*、*CAR* 巨噬细胞和 *CAR-NKT* 等) 逐渐受到重视<sup>[54-56]</sup>。这些策略通过多样化的免疫机制, 可能在增强靶向性、克服耐药性及提升持久性方面展现优势。虽然仍处于探索阶段, 但为 ATC 的治疗提供新的研究思路。

## 5 小结

CAR-T 疗法为侵袭性极强且传统疗法效果有限的 ATC 带来了新的治疗希望。当前研究已识别多种潜力靶点: 其中 *ICAM-1* 因其在 ATC 中高度表达且可增强肿瘤对免疫细胞的敏感性, 成为最有前景的临床转化靶标。第 3 代亲和力和优化的 *ICAM-1 CAR-T* (*AICI100*) 在 I 期试验中显示较好疗效和安全性。尽

管 *TSHR* 在 ATC 中表达下调限制了其靶向价值, 但通过 *MAPKi* 暂时恢复 *TSHR* 表达并联用 *TSHR CAR-T* 的策略在临床前已证实可协同增效。新靶点如 *ROR1*、*CSPG4* 也展现出显著的靶点依赖性抗肿瘤活性, 尤其对缺乏 *BRAF V600E* 突变的难治性 ATC 患者提供了新方向。然而, CAR-T 疗法仍面临复杂的肿瘤微环境抑制、靶点异质表达及免疫逃逸等多重挑战。未来优化策略可包括: 重塑 TME、与免疫检查点抑制剂或靶向药物联用以提升疗效、研发 *CAR-NK/CAR-Ms* 等替代平台以增强稳定性并降低毒性等。同时, 应加速新靶点 (如 *ROR1*) 和组合疗法的临床验证, 设计多模式治疗方案以整合微环境调控与抗原激活, 实现对 ATC 的多层次打击, 并在更大规模临床试验中评估疗效与安全性, 以期最终改善 ATC 患者的生存预后。

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